



# Effects of PSA Removal from NCAM on the Critical Period Plasticity Triggered by the Antidepressant Fluoxetine in the Visual Cortex

Ramon Guirado<sup>1\*</sup>, Danilo La Terra<sup>1</sup>, Mathieu Bourguignon<sup>2,3</sup>, Hector Carceller<sup>4</sup>, Juzoh Umemori<sup>1</sup>, Pia Sipilä<sup>1,5</sup>, Juan Nacher<sup>4</sup> and Eero Castrén<sup>1\*</sup>

<sup>1</sup> Neuroscience Center, University of Helsinki, Helsinki, Finland, <sup>2</sup> Department of Neuroscience and Biomedical Engineering, Aalto University, Helsinki, Finland, <sup>3</sup> Basque Center on Cognition, Brain and Language, Donostia, Spain, <sup>4</sup> Departamento de Biología Celular, Spanish National Network for Research in Mental Health, CIBERSAM, Fundación Investigación Hospital Clínico de Valencia, INCLIVA, Universitat de Valencia, Valencia, Spain, <sup>5</sup> Max Planck Institute for Neurobiology, Martinsried, Germany

## OPEN ACCESS

### Edited by:

Chao Deng,  
University of Wollongong, Australia

### Reviewed by:

Graziella DiCristo,  
University of Montreal, Canada  
Herbert Hildebrandt,  
Hannover Medical School, Germany

### \*Correspondence:

Ramon Guirado  
ramon.guirado@helsinki.fi;  
Eero Castrén  
eero.castrén@helsinki.fi

**Received:** 13 November 2015

**Accepted:** 22 January 2016

**Published:** 05 February 2016

### Citation:

Guirado R, La Terra D, Bourguignon M, Carceller H, Umemori J, Sipilä P, Nacher J and Castrén E (2016) Effects of PSA Removal from NCAM on the Critical Period Plasticity Triggered by the Antidepressant Fluoxetine in the Visual Cortex. *Front. Cell. Neurosci.* 10:22. doi: 10.3389/fncel.2016.00022

Neuronal plasticity peaks during critical periods of postnatal development and is reduced towards adulthood. Recent data suggests that windows of juvenile-like plasticity can be triggered in the adult brain by antidepressant drugs such as Fluoxetine. Although the exact mechanisms of how Fluoxetine promotes such plasticity remains unknown, several studies indicate that inhibitory circuits play an important role. The polysialylated form of the neural cell adhesion molecules (PSA-NCAM) has been suggested to mediate the effects of Fluoxetine and it is expressed in the adult brain by mature interneurons. Moreover, the enzymatic removal of PSA by neuroaminidase-N not only affects the structure of interneurons but also has been shown to play a role in the onset of critical periods during development. We have here used ocular dominance plasticity in the mouse visual cortex as a model to investigate whether removal of PSA might influence the Fluoxetine-induced plasticity. We demonstrate that PSA removal in the adult visual cortex alters neither the baseline ocular dominance, nor the fluoxetine-induced shift in the ocular dominance. We also show that both chronic Fluoxetine treatment and PSA removal independently increase the basal FosB expression in parvalbumin (PV) interneurons in the primary visual cortex. Therefore, our data suggest that although PSA-NCAM regulates inhibitory circuitry, it is not required for the reactivation of juvenile-like plasticity triggered by Fluoxetine.

**Keywords:** fluoxetine, critical period plasticity, PSA-NCAM, parvalbumin interneurons, visual plasticity

## INTRODUCTION

Critical periods during development have been characterized as time-windows of high plasticity during which the external milieu shapes networks and behavior (Katz and Shatz, 1996; Berardi et al., 2000). In the visual cortex, the classical studies of Hubel and Wiesel showed how ocular dominance columns are established during a critical period in postnatal development of the visual system in cats (Wiesel and Hubel, 1963). After that time-window of enhanced plasticity, the established ocular dominance will remain permanent (Hubel and Wiesel, 1970). Disturbance of normal binocular vision during such critical period, in animal models usually by monocular deprivation,

leads to disrupted integration of visual information in the cortex. The altered balance of information arriving to the visual cortex from the occluded eye results in reduced visual acuity and contrast sensitivity (Dews and Wiesel, 1970; Fagiolini et al., 1994; Prusky et al., 2000), a condition known in humans as amblyopia (Webber and Wood, 2005).

Although the visual system of rodents lacks characteristic columns described in higher mammals, critical periods for ocular dominance have nevertheless been extensively characterized in rodents as well (Kirkwood and Bear, 1994; Kirkwood et al., 1995; Huang et al., 1999). Interestingly, recent reports have shown that a critical period-like plasticity can be reopened in the adult visual cortex through different experimental manipulations (Sale et al., 2009; Bavelier et al., 2010; Beurdeley et al., 2012) including antidepressant drugs such as Fluoxetine (Maya-Vetencourt et al., 2008; Tiraboschi et al., 2013). The combination of Fluoxetine with monocular deprivation of one eye in adult rats produces a shift in the ocular dominance towards the open eye. This shift resembles that achieved by the same paradigm of monocular deprivation during the developmental critical period (Maya-Vetencourt et al., 2008).

Cellular and molecular mechanisms that have been connected with the Fluoxetine-induced opening of critical periods in adult brain include rearrangement of inhibitory circuits (Maya-Vetencourt et al., 2008), signaling by Brain-derived neurotrophic factor (BDNF; Maya-Vetencourt et al., 2008), serotonergic transmission (Maya-Vetencourt et al., 2010; Ferrés-Coy et al., 2015) and the transcription factor NPAS4 (Maya-Vetencourt et al., 2012), but detailed molecular mechanisms through which Fluoxetine reopens critical periods are still unclear (Maya-Vetencourt et al., 2012; Sharp, 2013; Tiraboschi et al., 2013). A promising candidate to mediate this mechanism is the polysialylated form of neural cell adhesion molecules (PSA-NCAM), a molecule widely expressed during development but subsequently restricted, in the adult neocortex, to a subpopulation of interneurons (Sandi, 2004; Rutishauser, 2008). PSA-NCAM is a negatively charged hydrophilic molecule that creates a steric impediment affecting membrane receptors and signaling (Burgess and Aubert, 2006; Gascon et al., 2007). NCAM is the major polysialylated molecule in the adult brain and its polysialylation has been linked with adult brain plasticity (Bonfanti, 2006; Nacher et al., 2013). Particularly, chronic antidepressant treatments have been reported to increase PSA-NCAM expression in different brain regions (Sairanen et al., 2007; Varea et al., 2007; Guirado et al., 2014b). Conversely, enzymatic removal of PSA has been shown to disrupt plastic mechanisms such as LTP (Muller et al., 1996; Dityatev et al., 2004); effects that can be counteracted by BDNF supplementation (Muller et al., 2000). In addition, PSA removal has also been suggested to trigger an earlier onset of critical periods in the visual cortex through the precocious maturation of the perisomatic innervation of pyramidal neurons by basket cells and enhancing inhibitory synaptic transmission (Di Cristo et al., 2007). In fact, PSA depletion during adulthood also induces an increase in this perisomatic inhibitory input (Castillo-Gómez et al., 2011). All these studies suggest

that regulation of PSA-NCAM expression is a possible mechanism mediating the effects of Fluoxetine in the adult brain plasticity.

To investigate the interaction between PSA-NCAM and Fluoxetine, we studied the effects of enzymatic removal of the polysialic acid moiety of PSA-NCAM on a critical period plasticity paradigm: the shifts of ocular dominance in the visual cortex during Fluoxetine chronic treatment.

## MATERIALS AND METHODS

### Animals

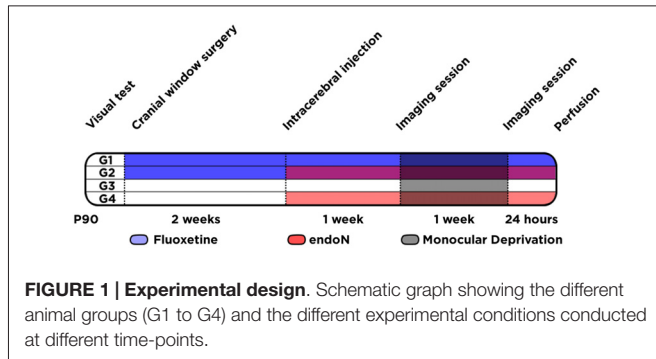
Forty-eight C57BL/6 female mice (Harlan Laboratories, Netherlands) were used. Animals were divided into 4 different groups of 12 to control two experimental manipulations: Fluoxetine treatment and Endoneuraminidase-N (endoN) intracerebral injections (group 1: no manipulation; group 2: Fluoxetine only; group 3: EndoN only; group 4: Fluoxetine and EndoN). The animals were caged in groups of four and kept under standard housing conditions (21°C, 12 h light/12 h dark cycle, lights on at 6 AM). Mice were 3 months old at the beginning of the experiment, ensuring that the shift in the ocular dominance took place after P110 when the extended critical period has been suggested to end (Greifzu et al., 2014). All animals had free access to food and water for the entire duration of the experimentation. All procedures were carried out according to the Directive 2010/63/EU of the European Parliament and were approved by the Animal Ethical Committee of Southern Finland.

### Visual Abilities

Following a previously described methodology (Drapeau et al., 2014), general visual acuity was tested by using the visual placing test, which takes advantage of the forepaw-reaching reflex: the mouse was held by its tail approximately 20 cm above the surface and was then progressively lowered. As it approaches the surface, the mouse should expand its forepaws to reach the floor. The test was repeated three times with a 30 s interval, and the distance from the ground when the animals show the forepaw reflex without using their whiskers was quantified.

### Fluoxetine Treatment

Mice received Fluoxetine via drinking water. Drinking water was supplemented with 0.16 mg/mL Fluoxetine and 0.5% sucrose. Solutions were prepared fresh twice a week, and drinking bottles protected from light. We started the treatment immediately after the cranial window surgery, and it lasted for 4 weeks until the end of the experiment (**Figure 1**). The level of Fluoxetine/water consumed and animal bodyweight were constantly monitored to evaluate drug dose, which was approximately 20 mg/Kg/day. This protocol of chronic Fluoxetine administration to adult mice was reported to result in clinical relevant plasma levels of the drug (Dulawa et al., 2004).



## Chronic Cranial Window Surgical Procedure

Three weeks prior to the first imaging session animals were implanted flush with cranial windows in their right hemisphere, following a protocol adapted from Holtmaat et al. (2009). In short: anesthesia was induced with 3–4% Isoflurane (Vetflurane), then continued with intraperitoneal (IP) injection of sleeping cocktail: Fentanyl (opioid receptor agonist; 0.05 mg/kg), Midazolam (GABA-A agonist; 5.0 mg/Kg) and Medetomidine (adrenergic receptor agonist; 0.5 mg/Kg). Animals were placed in a stereotaxic frame, the skin and periosteum were removed and the right temporalis muscles separated from the bone. With this procedure a large area of the skull was exposed. Using a small amount of cyanoacrylate (Vetbond) we seal the skin on the sides. A circular groove was drawn using a surgical drill. Then, the bone piece was lifted leaving the dura intact. A 5 mm diameter circular cover glass (EMS) was flushed on. Glue gel (Loctite) was applied around the borders and under a metal head bar to glue it to the skull. Finally, dentist cement was applied around the craniotomy to cover all tissues exposed, except a small area ipsilateral to the cranial window, covered with putty for the following intracerebral injection.

## Stereotaxic Injection of EndoN

As previously described, mice were anesthetized first with isoflurane and then with a sleeping cocktail. Additionally, animals were injected subcutaneously with carprofen (Rimadyl; 5 mg/Kg) and intramuscularly with dexamethasone (0.6 mg/Kg), for pre-emptive analgesia and anti-inflammatory effects. Then placed in a stereotaxic frame, the piece of putty was removed for the intracerebral injection. The enzyme endoN (0.7 U/ $\mu$ L in glycerol; AbCys) or the vehicle solution (1  $\mu$ L; saline and glycerol 1:1) was injected using a Hamilton syringe in the ipsilateral hemisphere to the cranial window within the somatosensory cortex (antero-posterior  $-0.5$  mm, medio-lateral  $-1.5$  mm, dorso-ventral  $-0.7$  mm, relative to Bregma). The needle was left in position for 1 min and then 1  $\mu$ L of endoN or vehicle was injected during another minute. After the injection was completed, the needle was left in place for 3 min to reduce reflux of the solution into the track of the injection needle and then slowly withdrawn.

After all surgical procedures, animals received an IP injection with a wake-up cocktail containing antagonists of the same receptors affected by the sleeping cocktail: Naloxone (1.2 mg/Kg), Flumazenil (0.5 mg/Kg) and Atipamezole (2.5 mg/Kg).

## In Vivo Imaging of Intrinsic Signals

We adapted our methodology from a previous protocol described by the laboratory of Dr Stryker (Kalatsky and Stryker, 2003; Cang et al., 2005) to measure saturation of hemeoglobin as an indicator of cortical activity in the mouse visual cortex. Mice were injected IP with a lower dose (2/3 of the surgical dose) of the sleeping cocktail to produce a shallow anesthesia. Then, animals were placed in the setup, under a high frequency CCD camera (Brain Imager 3001/S) 25 cm in front of a screen. The cranial window was illuminated with red light using a broad pass filter ( $630 \pm 20$  nm), and the camera focused 500  $\mu$ m below the surface area with a field of view roughly matching the full extent of the cranial window. We produced a visual stimulus consisting in a two degrees-thick horizontal bar in a position between  $-15$  and  $+5$  degrees azimuth (relative to the field of vision of the animal). The bar flickered at 6 Hz and drifted vertically with cycles corresponding to 0.11 Hz to elicit rhythmic activity at the same frequency in the right hemisphere from both ipsilateral and contralateral eyes. Four stimulation protocols were performed: two for each eye, and within one eye one moving upwards and one downwards. In each protocol, images of dimensions  $336 \times 336$  pixels were sampled at 10 Hz for 5 min.

## Monocular Deprivation

After the first imaging session animals still under light anesthesia were implanted with eye occluders in the left eye (contralateral to the cranial window). These occluders were made as previously described (Park et al., 2005). In short, they were made from 0.25 mm thick black vinyl sheets that were heated and molded over a steel ball similar in size to the mice eyes. Mice from all groups were subjected to monocular deprivation during 7 days, after which they underwent a second imaging session. In a separate pilot experiment we ensured that there are no responses in the visual cortex when the animals wearing occluders are exposed to the same visual stimulus.

## Analysis of Imaging Data

Images were analyzed offline with custom-made scripts in matlab (MATLAB 7.8, The MathWorks, Inc). These scripts are freely available upon request. Images were stacked in the time-domain and Fourier-transformed in that dimension to extract a map of signal amplitude and phase at 0.11 Hz (the stimulation frequency). Amplitude images for stimuli moving upwards and downwards were averaged together, leaving us with one amplitude image per eye (contralateral and ipsilateral). Images were further smoothed with a 10-pixels-wide full-width-at-half-maximum Gaussian kernel in order to decrease the noise level. A region of interest that in principle matches with the location of the primary visual cortex was then defined

as the pixels for which the image for the contralateral eye take values above 70% of the maximum of that same image. Finally, the ocular dominance index (ODI) was defined as  $ODI = (A_{contra} - A_{ipsi}) / (A_{contra} + A_{ipsi})$ , where  $A_{contra}$  (and  $A_{ipsi}$  respectively) is the mean amplitude of the image for the contralateral (resp. ipsilateral) eye within the region of interest. Phase maps masked outside the region of interest were used to confirm that cortical activity is entrained according to the known retinotopy; i.e., with a phase in each cortical areas directly related to its most responsive polar angle. Mice who failed to demonstrate systematic activity at 0.11 Hz and expected retinotopy were excluded from further analysis of the ODI.

## Perfusion

One day after the second *in vivo* imaging session, animals were perfused transcardially under deep pentobarbital anesthesia, first with saline and then with PFA 4% in PB 0.1 M. After perfusions, brains were extracted and postfixed in the same fixative solution for 2 h, then stored in PB 0.1 M with sodium azide 0.05%. Then brains were cut into 50  $\mu\text{m}$  sections using a vibratome (VT 1000E, Leica). Sections were collected in six different subseries and stored at 4°C in PB 0.1 M and sodium azide.

## Immunohistochemistry

Tissue was processed free-floating for fluorescence immunohistochemistry as follows. Sections were washed in PBS, then slices were incubated in 10% normal donkey serum (NDS; Gibco), 0.2% Triton-X100 (Sigma) in PBS for 1 h. Sections were then incubated for 48 h at 4°C with IgM mouse anti-PSA-NCAM (1:500; DSHB) or with a cocktail containing IgG mouse anti-parvalbumin (1:2000; Swant) and IgG rabbit anti-FosB (1:500; Santa Cruz) antibodies diluted in PBS 0.2% Triton-X100. After washing again, sections were incubated for 2 h at room temperature with secondary antibody anti-mouse IgM conjugated with A488 (1:200; Life Technologies) or with a cocktail containing: anti-mouse IgG conjugated with Alexa 647 and anti-rabbit IgG conjugated with Alexa 488 (1:200; Life Technologies). Sections were then washed in PB 0.1 M, mounted on slides and coverslipped using fluorescence mounting medium (Dako).

## Confocal Imaging

Sections were analyzed using a confocal microscope (Zeiss LSM 700). We evaluated PSA-NCAM expression and quantified PV and FosB fluorescence intensity in two different sections containing primary visual cortex (between +3 and +3.7 mm in the Antero-Posterior relative to Bregma). Stacks were obtained at an optimal penetration depth in the tissue (1–6  $\mu\text{m}$  deep). We used same acquiring conditions (as any other previous condition) to compare then the intensity of fluorescence between the samples. For PSA-NCAM quantification, small square areas of 500  $\mu\text{m}^2$  were analyzed in deep layers of two different sections within the primary visual cortex of each animal. For FosB and PV quantification, all PV neurons were counted in that same optimal penetration depth in the tissue, then the nucleus stained with

FosB was profiled and its fluorescence intensity quantified for both channels: FosB and PV. A total of 639 PV neurons were analyzed, although for statistical purposes we always used the number of animals as “*n*”. All animals were coded and the code was not broken until the end of the study.

## RESULTS

### EndoN Efficiently Depletes PSA-NCAM in the Visual Cortex

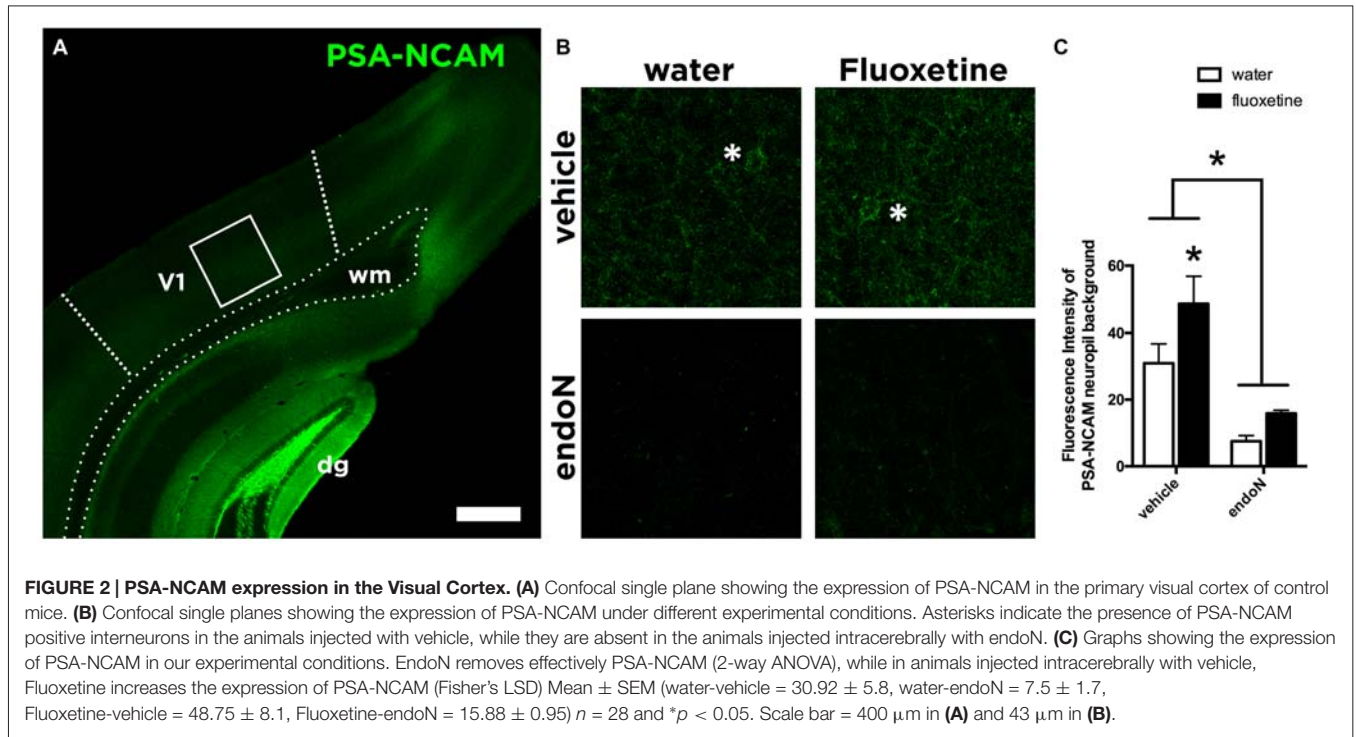
We analyzed the expression of the PSA-NCAM in the primary visual cortex of all the animals (**Figure 2A**). Intracerebral injections of the enzyme endoN produced a clear depletion of PSA-NCAM. Two weeks after the injection of the enzyme, the depletion covered most parts of the telencephalon, including always the visual cortex of the right hemisphere where the cranial windows were placed (**Figure 2B**). In these areas the enzyme effectively removed PSA-NCAM (2-way ANOVA; endoN effect:  $F_{(1,24)} = 17.02$  and  $p = 0.0004$ ; **Figure 2C**). In addition, Fluoxetine treatment increased the expression of PSA-NCAM (Fisher's LSD;  $p = 0.0004$ ) in control animals as previously described (Varea et al., 2007).

### Fluoxetine Produces a Shift in the Ocular Dominance but EndoN Does Not

Before the start of the experiment, all animals were evaluated for gross visual acuity. Three animals failed to show the forepaw reflex without using their whiskers and were removed from the study. The rest of the animals, showing at least one successful forepaw reflex without using the whiskers, were therefore used for the paradigm of visual plasticity (**Figure 3B**).

We then analyzed the effects of Fluoxetine and endoN in a paradigm of visual plasticity. By measuring the activity during visual stimulation in each eye, we calculated values of ODI as previously described (Cang et al., 2005) for each animal before and after monocular deprivation (**Figure 3A**). Briefly, we calculated the shift in ODI for each animal that occurred between the first imaging session (before monocular deprivation) and the second imaging session (after monocular deprivation). Of the 45 animals analyzed, 14 failed to reveal systematic activity at stimulation frequency (0.11 Hz) and were therefore not included in further analyses, leaving us with 31 animals (no manipulation: 8; Fluoxetine only: 6; EndoN only: 8; Fluoxetine and EndoN: 9). We found that animals not treated with Fluoxetine did not undergo any shift in ODI, regardless of whether they were intracerebrally injected with endoN (paired *t*-test before vs. after MD;  $p = 0.29$ ), or with vehicle (paired *t*-test;  $p = 0.49$ ), indicating that PSA removal did not influence basal visual responses. Fluoxetine treatment induced a significant shift of the ODI towards the non-occluded eye in the control (vehicle-injected) mice, consistent with previous findings in rats (Maya-Vetencourt et al., 2008; paired *t*-test;  $p = 0.035$ , **Figure 3C**). Similarly, Fluoxetine also produced a similar significant shift in the ODI in mice intracerebrally injected with endoN (paired *t*-test;  $p = 0.036$ , **Figure 3C**). This effect is also observed when comparing the shift of ODI





with a 2-way ANOVA (factors: Fluoxetine and EndoN). We indeed found a significant effect of Fluoxetine ( $F_{(1,27)} = 5.102$  and  $p = 0.032$ ), no effect of endoN ( $F_{(1,27)} = 0.332$  and  $p = 0.063$ ), and no Fluoxetine-endoN interaction ( $F_{(1,27)} = 0.089$  and  $p = 0.76$ ). These results suggest that depletion of PSA-NCAM influenced neither the basal properties nor the fluoxetine-induced plasticity of the ocular dominance in the mouse visual cortex.

### Both Fluoxetine and EndoN Alter the Basal Activity of Parvalbumin Interneurons

Parvalbumin (PV) expressing interneurons are considered key regulators of the critical period plasticity and their axon terminals are modulated by PSA-NCAM expression (Di Cristo et al., 2007; Castillo-Gómez et al., 2011). We hypothesize that PSA-NCAM modulation on the axon terminals might affect the synaptic activity of PV interneurons. We therefore analyzed the effects of chronic Fluoxetine treatment and intracerebral injection of endoN on the expression of the basal activity marker FosB (Nestler et al., 2001; Robison and Nestler, 2011) in PV interneurons in the primary visual cortex (Figures 4A,B). Fluoxetine treatment produced a significant increase of FosB expression in PV positive interneurons in animals injected with vehicle (Fisher's LSD;  $p = 0.020$ ; Figure 4C). In addition, endoN injection also increased FosB intensity in PV positive interneurons compared to control animals (Fisher's LSD;  $p = 0.0146$ ; Figure 4C). However, it is interesting to note that the combination of chronic Fluoxetine and the removal of PSA returned the expression of FosB to the basal control levels,

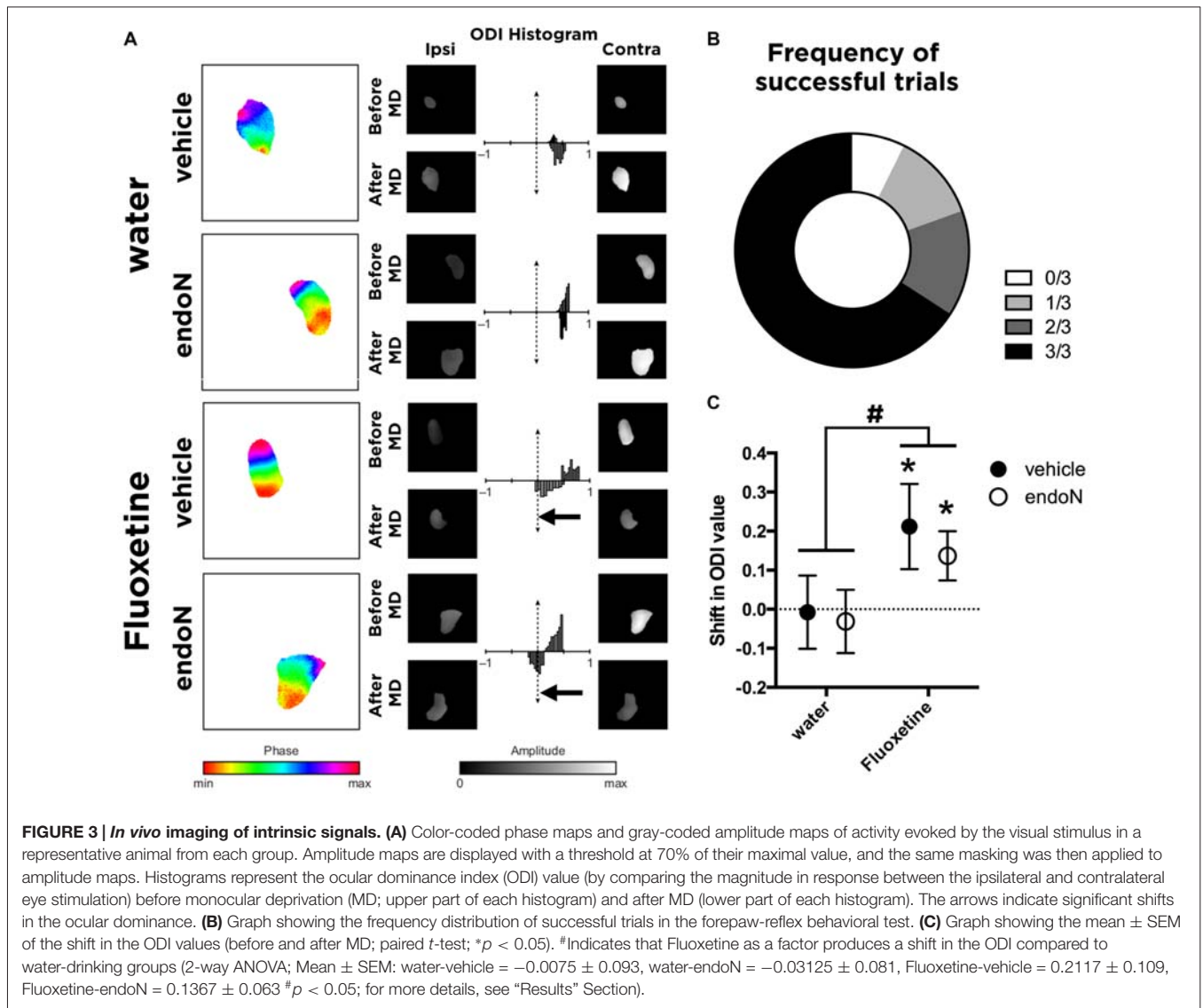
indicating an interaction (2-way ANOVA;  $F_{(1,31)} = 11.36$  and  $p = 0.0020$ ).

Finally, we analyzed more in detail the expression of PV. By measuring the fluorescence intensity of PV, we found a trend in the interaction between both factors: endoN and Fluoxetine (2-way ANOVA;  $F_{(1,31)} = 3.908$  and  $p = 0.0570$ ; Figure 4D). In the multiple comparisons test, we found that in animals injected intracerebrally with vehicle, Fluoxetine treatment increased the fluorescence intensity of PV interneurons (Fisher's LSD;  $p = 0.0362$ ). However, the total number of neurons in the visual cortex was unaffected (2-way ANOVA; Fluoxetine:  $F_{(1,31)} = 0.2996$  and  $p = 0.5881$ ; endoN  $F_{(1,31)} = 0.7434$  and  $p = 0.3952$ ; Figure 4E).

## DISCUSSION

In the present study, we have used ocular dominance plasticity in the mouse visual cortex as a model to investigate whether removal of PSA might influence the Fluoxetine-induced reactivation of critical period-like plasticity in the adult brain. We show that injection of the enzyme endoN produces a clear removal of PSA that after 2 weeks extends to the visual cortex and most parts of the telencephalon, indicating that the enzymatic treatment reliably disrupts the PSA side chains of NCAM in adult brain.

As previously reported in rats (Maya-Vetencourt et al., 2008), we found that chronic Fluoxetine treatment reopens a period of plasticity in the adult mouse visual cortex that resembles the original critical period of postnatal development. Specifically, chronic Fluoxetine treatment has also been demonstrated to



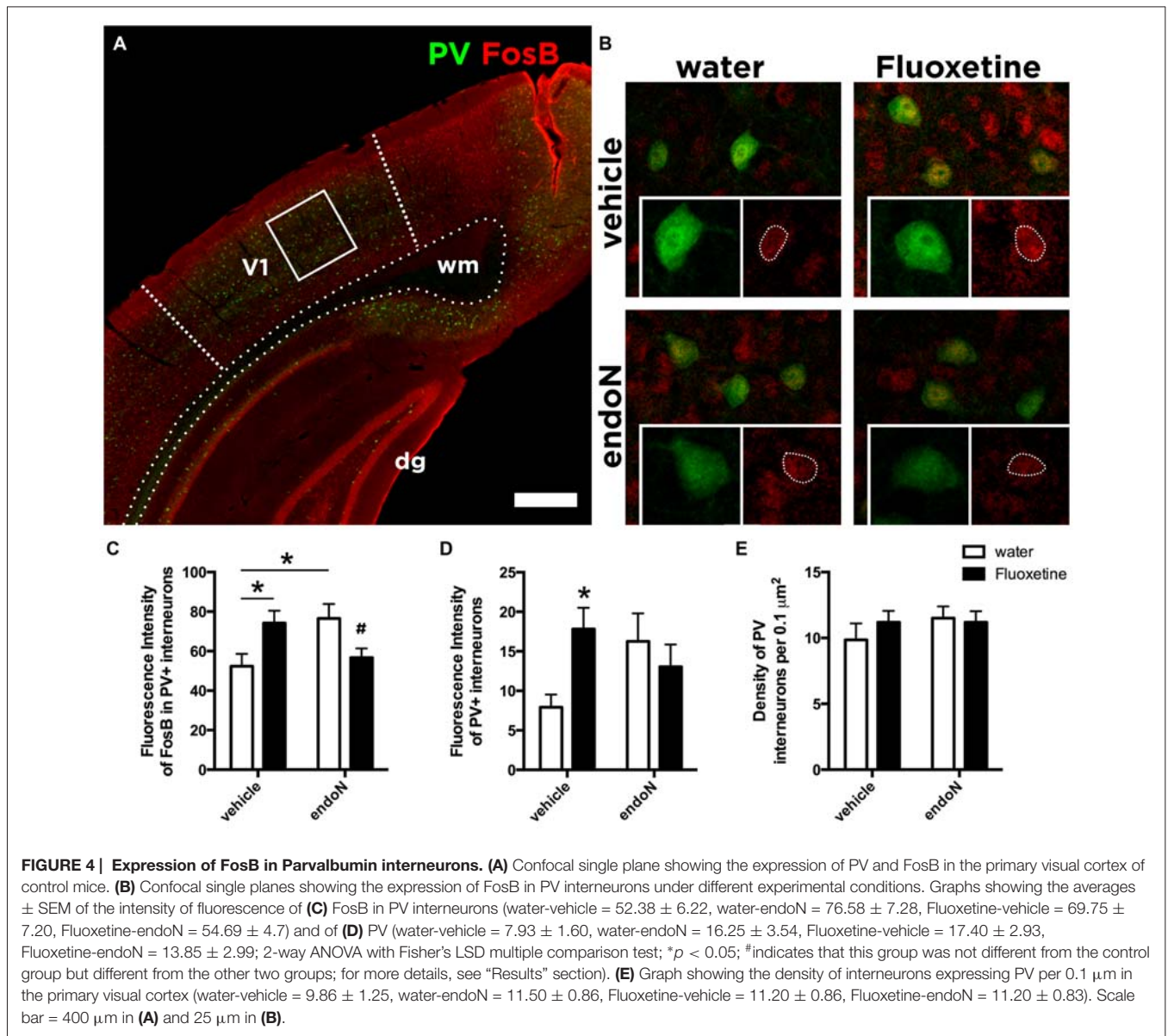
produce an increase in PSA-NCAM expression and other molecules involved in inhibitory neurotransmission (Varea et al., 2007; Karpova et al., 2011; Ohira et al., 2013; Guirado et al., 2014a). On the other hand, removal of PSA by intracerebral infusion of endoN in the basolateral amygdala has been shown to enhance extinction of fear memories (Markram et al., 2007). At the cellular level, removal of PSA produces specific structural plasticity changes in different subpopulations of interneurons (Saarelainen et al., 2003; Varea et al., 2007; Guirado et al., 2014b).

Regarding visual plasticity, the decline of PSA-NCAM expression after early development has been linked to the earlier onset of critical period plasticity through an enhanced perisomatic innervation by PV interneurons in the visual cortex (Di Cristo et al., 2007). We hypothesized that the increased PSA-NCAM expression produced by Fluoxetine, resembling that high PSA-NCAM expression during development, might be necessary for the structural remodeling underlying the shifts in the ocular dominance. However, we have found that depletion of

PSA by the injection of endoN influenced neither the basal ocular dominance conditions, nor the reactivation of juvenile-like plasticity induced by Fluoxetine. Therefore, our results suggest that PSA-NCAM is not required for the plasticity triggered by Fluoxetine in the visual cortex.

The opening and closure of critical periods in the visual cortex has been suggested to be dependent on the remodeling of inhibitory circuits (Yazaki-Sugiyama et al., 2009; Harauzov et al., 2010), and therefore the disruption the excitatory/inhibitory balance (Hensch, 2005). Specifically, modulation of PV interneurons activity has been suggested to play a role in triggering critical period plasticity (Hensch, 2005; Kuhlman et al., 2013).

Therefore we decided to study the expression of FosB as a marker of basal long-term activity (Nestler et al., 2001; Robison and Nestler, 2011; Nestler, 2015) in PV interneurons. While we have found that Fluoxetine increases the expression of PV in control animals, we also show that both chronic



Fluoxetine treatment and PSA removal, independently produce an increase in the expression of FosB in PV interneurons. This is in line with a very recent study that showed an increased FosB expression throughout the brain after Fluoxetine treatment (Vialou et al., 2015). It is also in agreement with a previous study that indicated an increase in the density of PV and PV/synaptophysin expressing puncta on the perisomatic region of pyramidal neurons after PSA removal (Castillo-Gómez et al., 2011), what suggests an increased activity of PV interneurons. In fact, PSA removal has previously been suggested to lead to increased activity via BDNF (Burgess and Aubert, 2006). However, it is interesting to note the negative interaction between both factors as well, suggesting they act through different mechanisms on PV interneurons. Further experiments including control animals in which no monocular

deprivation is carried out, might help to understand this negative interaction.

Since we observe an increase in the FosB expression in PV interneurons in animals only injected with endoN, these results indicate that the average changes of activity of PV interneurons are not sufficient to trigger adult brain plasticity. As it has been suggested previously, brain plasticity might depend on the activity of few selected PV hub interneurons (Bonifazi et al., 2009; Buzsáki, 2010; Royer et al., 2012; Cossart, 2014). Therefore, it might be that our study masks the specific changes in the activity of those few selected hub PV interneurons. Nevertheless, further studies will be necessary to understand the mechanisms through which antidepressants, such as Fluoxetine, trigger critical period plasticity in the adult brain.



## AUTHOR CONTRIBUTIONS

RG, JU, PS and EC designed research; RG, DLT and HC performed research; RG and MB analyzed data; JN, MB, RG and EC wrote the article.

## REFERENCES

- Bavelier, D., Levi, D., Li, R., Dan, Y., and Hensch, T. (2010). Removing brakes on adult brain plasticity: from molecular to behavioral interventions. *J. Neurosci.* 30, 14964–14971. doi: 10.1523/JNEUROSCI.4812-10.2010
- Berardi, N., Pizzorusso, T., and Maffei, L. (2000). Critical periods during sensory development. *Curr. Opin. Neurobiol.* 10, 138–145. doi: 10.1016/s0959-4388(99)00047-1
- Beurdeley, M., Spatzza, J., Lee, H. H., Sugiyama, S., Bernard, C., Di Nardo, A. A., et al. (2012). Otx2 binding to perineuronal nets persistently regulates plasticity in the mature visual cortex. *J. Neurosci.* 32, 9429–9437. doi: 10.1523/JNEUROSCI.0394-12.2012
- Bonfanti, L. (2006). PSA-NCAM in mammalian structural plasticity and neurogenesis. *Prog. Neurobiol.* 80, 129–164. doi: 10.1016/j.pneurobio.2006.08.003
- Bonifazi, P., Goldin, M., Picardo, M. A., Jorquera, I., Cattani, A., Bianconi, G., et al. (2009). GABAergic hub neurons orchestrate synchrony in developing hippocampal networks. *Science* 326, 1419–1424. doi: 10.1126/science.1175509
- Burgess, A., and Aubert, I. (2006). Polysialic acid limits choline acetyltransferase activity induced by brain-derived neurotrophic factor. *J. Neurochem.* 99, 797–806. doi: 10.1111/j.1471-4159.2006.04110.x
- Buzsáki, G. (2010). Neural syntax: cell assemblies, synapsembles and readers. *Neuron* 68, 362–385. doi: 10.1016/j.neuron.2010.09.023
- Cang, J., Kalatsky, V. A., Löwel, S., and Stryker, M. P. (2005). Optical imaging of the intrinsic signal as a measure of cortical plasticity in the mouse. *Vis. Neurosci.* 22, 685–691. doi: 10.1017/s0952523805225178
- Castillo-Gómez, E., Varela, E., Blasco-Ibáñez, J., Crespo, C., and Nacher, J. (2011). Polysialic acid is required for dopamine D2 receptor-mediated plasticity involving inhibitory circuits of the rat medial prefrontal cortex. *PLoS One* 6:e29516. doi: 10.1371/journal.pone.0029516
- Cossart, R. (2014). Operational hub cells: a morpho-physiologically diverse class of GABAergic neurons united by a common function. *Curr. Opin. Neurobiol.* 26, 51–56. doi: 10.1016/j.conb.2013.12.002
- Dews, P. B., and Wiesel, T. N. (1970). Consequences of monocular deprivation on visual behaviour in kittens. *J. Physiol. (Lond)* 206, 437–455. doi: 10.1113/jphysiol.1970.sp009023
- Di Cristo, G., Chattopadhyaya, B., Kuhlman, S., Fu, Y., Bélanger, M.-C., Wu, C., et al. (2007). Activity-dependent PSA expression regulates inhibitory maturation and onset of critical period plasticity. *Nat. Neurosci.* 10, 1569–1577. doi: 10.1038/nn2008
- Dityatev, A., Dityateva, G., Sytnyk, V., Dellling, M., Toni, N., Nikonenko, I., et al. (2004). Polysialylated neural cell adhesion molecule promotes remodeling and formation of hippocampal synapses. *J. Neurosci.* 24, 9372–9382. doi: 10.1523/JNEUROSCI.1702-04.2004
- Drapeau, E., Dorr, N. P., Elder, G. A., and Buxbaum, J. D. (2014). Absence of strong strain effects in behavioral analyses of Shank3-deficient mice. *Dis. Model. Mech.* 7, 667–681. doi: 10.1242/dmm.013821
- Dulawa, S. C., Holick, K. A., Gundersen, B., and Hen, R. (2004). Effects of chronic fluoxetine in animal models of anxiety and depression. *Neuropsychopharmacology* 29, 1321–1330. doi: 10.1038/sj.npp.1300433
- Fagiolini, M., Pizzorusso, T., Berardi, N., Domenici, L., and Maffei, L. (1994). Functional postnatal development of the rat primary visual cortex and the role of visual experience: dark rearing and monocular deprivation. *Vision Res.* 34, 709–720. doi: 10.1016/0042-6989(94)90210-0
- Ferrés-Coy, A., Galofré, M., Pilar-Cuellar, F., Vidal, R., Paz, V., Ruiz-Bronchal, E., et al. (2015). Therapeutic antidepressant potential of a conjugated siRNA silencing the serotonin transporter after intranasal administration. *Mol. Psychiatry* doi: 10.1038/mp.2015.80 [Epub ahead of print].
- Gascon, E., Vutskits, L., and Kiss, J. (2007). Polysialic acid-neural cell adhesion molecule in brain plasticity: from synapses to integration of new neurons. *Brain Res. Rev.* 56, 101–118. doi: 10.1016/j.brainresrev.2007.05.014
- Greifzu, F., Pielecka-Fortuna, J., Kalogeraki, E., Krempler, K., Favaro, P., Schlüter, O., et al. (2014). Environmental enrichment extends ocular dominance plasticity into adulthood and protects from stroke-induced impairments of plasticity. *Proc. Natl. Acad. Sci. U S A* 111, 1150–1155. doi: 10.1073/pnas.1313385111
- Guirado, R., Perez-Rando, M., Sanchez-Matarredona, D., Castillo-Gómez, E., Liberia, T., Rovira-Esteban, L., et al. (2014a). The dendritic spines of interneurons are dynamic structures influenced by PSA-NCAM expression. *Cereb. Cortex* 24, 3014–3024. doi: 10.1093/cercor/bht156
- Guirado, R., Perez-Rando, M., Sanchez-Matarredona, D., Castrén, E., and Nacher, J. (2014b). Chronic fluoxetine treatment alters the structure, connectivity and plasticity of cortical interneurons. *Int. J. Neuropsychopharmacol.* 17, 1635–1646. doi: 10.1017/s1461145714000406
- Harauzov, A., Spolidoro, M., DiCristo, G., De Pasquale, R., Cancedda, L., Pizzorusso, T., et al. (2010). Reducing intracortical inhibition in the adult visual cortex promotes ocular dominance plasticity. *J. Neurosci.* 30, 361–371. doi: 10.1523/JNEUROSCI.2233-09.2010
- Hensch, T. (2005). Critical period plasticity in local cortical circuits. *Nat. Rev. Neurosci.* 6, 877–888. doi: 10.1038/nrn1787
- Holtmaat, A., Bonhoeffer, T., Chow, D. K., Chuckowree, J., De Paola, V., Hofer, S. B., et al. (2009). Long-term, high-resolution imaging in the mouse neocortex through a chronic cranial window. *Nat. Protoc.* 4, 1128–1144. doi: 10.1038/nprot.2009.89
- Huang, Z. J., Kirkwood, A., Pizzorusso, T., Porciatti, V., Morales, B., Bear, M. F., et al. (1999). BDNF regulates the maturation of inhibition and the critical period of plasticity in mouse visual cortex. *Cell* 98, 739–755. doi: 10.1016/s0092-8674(00)81509-3
- Hubel, D., and Wiesel, T. (1970). The period of susceptibility to the physiological effects of unilateral eye closure in kittens. *J. Physiol. (Lond)* 206, 419–436. doi: 10.1113/jphysiol.1970.sp009022
- Kalatsky, V., and Stryker, M. (2003). New paradigm for optical imaging: temporally encoded maps of intrinsic signal. *Neuron* 38, 529–545. doi: 10.1016/S0896-6273(03)00286-1
- Karpova, N., Pickenhagen, A., Lindholm, J., Tiraboschi, E., Kuleskaya, N., Agústsdóttir, A., et al. (2011). Fear erasure in mice requires synergy between antidepressant drugs and extinction training. *Science* 334, 1731–1734. doi: 10.1126/science.1214592
- Katz, L., and Shatz, C. (1996). Synaptic activity and the construction of cortical circuits. *Science* 274, 1133–1138. doi: 10.1126/science.274.5290.1133
- Kirkwood, A., and Bear, M. F. (1994). Hebbian synapses in visual cortex. *J. Neurosci.* 14, 1634–1645.
- Kirkwood, A., Lee, H. K., and Bear, M. F. (1995). Co-regulation of long-term potentiation and experience-dependent synaptic plasticity in visual cortex by age and experience. *Nature* 375, 328–331. doi: 10.1038/375328a0
- Kuhlman, S. J., Olivas, N. D., Tring, E., Ikrar, T., Xu, X., and Trachtenberg, J. T. (2013). A disinhibitory microcircuit initiates critical-period plasticity in the visual cortex. *Nature* 501, 543–546. doi: 10.1038/nature12485
- Markram, K., Lopez Fernandez, M. A., Abrós, D. N., and Sandi, C. (2007). Amygdala upregulation of NCAM polysialylation induced by auditory fear conditioning is not required for memory formation, but plays a role in fear extinction. *Neurobiol. Learn. Mem.* 87, 573–582. doi: 10.1016/j.nlm.2006.11.007
- Maya-Vetencourt, J. F., Sale, A., Viegi, A., Baroncelli, L., De Pasquale, R., O’Leary, O., et al. (2008). The antidepressant fluoxetine restores plasticity in the adult visual cortex. *Science* 320, 385–388. doi: 10.1126/science.1150516



- Maya-Vetencourt, J. F., Tiraboschi, E., Greco, D., Restani, L., Cerri, C., Auvinen, P., et al. (2012). Experience-dependent expression of NPAS4 regulates plasticity in adult visual cortex. *J. Physiol. (Lond)* 590, 4777–4787. doi: 10.1113/jphysiol.2012.234237
- Maya-Vetencourt, J. F., Tiraboschi, E., Spolidoro, M., Castrén, E., and Maffei, L. (2010). Serotonin triggers a transient epigenetic mechanism that reinstates adult visual cortex plasticity in rats. *Eur. J. Neurosci.* 33, 49–57. doi: 10.1111/j.1460-9568.2010.07488.x
- Muller, D., Djebbara-Hannas, Z., Jourdain, P., Vutskits, L., Durbec, P., Rougon, G., et al. (2000). Brain-derived neurotrophic factor restores long-term potentiation in polysialic acid-neural cell adhesion molecule-deficient hippocampus. *Proc. Natl. Acad. Sci. U S A* 97, 4315–4320. doi: 10.1073/pnas.070022697
- Muller, D., Wang, C., Skibo, G., Toni, N., Cremer, H., Calaora, V., et al. (1996). PSA-NCAM is required for activity-induced synaptic plasticity. *Neuron* 17, 413–422. doi: 10.1016/s0896-6273(00)80174-9
- Nacher, J., Guirado, R., and Castillo-Gómez, E. (2013). Structural plasticity of interneurons in the adult brain: role of PSA-NCAM and implications for psychiatric disorders. *Neurochem. Res.* 38, 1122–1133. doi: 10.1007/s11064-013-0977-4
- Nestler, E. J. (2015).  $\Delta$ FosB: a transcriptional regulator of stress and antidepressant responses. *Eur. J. Pharmacol.* 753, 66–72. doi: 10.1016/j.ejphar.2014.10.034
- Nestler, E. J., Barrot, M., and Self, D. W. (2001).  $\Delta$ FosB: a sustained molecular switch for addiction. *Proc. Natl. Acad. Sci. U S A* 98, 11042–11046. doi: 10.1073/pnas.191352698
- Ohira, K., Takeuchi, R., Iwanaga, T., and Miyakawa, T. (2013). Chronic fluoxetine treatment reduces parvalbumin expression and perineuronal nets in gamma-aminobutyric acidergic interneurons of the frontal cortex in adult mice. *Mol. Brain* 6:43. doi: 10.1186/1756-6606-6-43
- Park, J. H., Paul, M. J., Butler, M. P., and Zucker, I. (2005). Binocular interactions in the entrainment and phase shifting of locomotor activity rhythms in Syrian hamsters. *J. Biol. Rhythms* 20, 49–59. doi: 10.1177/0748730404272967
- Prusky, G. T., West, P. W., and Douglas, R. M. (2000). Experience-dependent plasticity of visual acuity in rats. *Eur. J. Neurosci.* 12, 3781–3786. doi: 10.1046/j.1460-9568.2000.00236.x
- Robison, A. J., and Nestler, E. J. (2011). Transcriptional and epigenetic mechanisms of addiction. *Nat. Rev. Neurosci.* 12, 623–637. doi: 10.1038/nrn3111
- Royer, S., Zemelman, B. V., Losonczy, A., Kim, J., Chance, F., Magee, J. C., et al. (2012). Control of timing, rate and bursts of hippocampal place cells by dendritic and somatic inhibition. *Nat. Neurosci.* 15, 769–775. doi: 10.1038/nn.3077
- Rutishauser, U. (2008). Polysialic acid in the plasticity of the developing and adult vertebrate nervous system. *Nat. Rev. Neurosci.* 9, 26–35. doi: 10.1038/nrn2285
- Saarelainen, T., Hendolin, P., Lucas, G., Koponen, E., Sairanen, M., MacDonald, E., et al. (2003). Activation of the TrkB neurotrophin receptor is induced by antidepressant drugs and is required for antidepressant-induced behavioral effects. *J. Neurosci.* 23, 349–357.
- Sairanen, M., O'Leary, O. F., Knuutila, J. E., and Castrén, E. (2007). Chronic antidepressant treatment selectively increases expression of plasticity-related proteins in the hippocampus and medial prefrontal cortex of the rat. *Neuroscience* 144, 368–374. doi: 10.1016/j.neuroscience.2006.08.069
- Sale, A., Berardi, N., and Maffei, L. (2009). Enrich the environment to empower the brain. *Trends Neurosci.* 32, 233–239. doi: 10.1016/j.tins.2008.12.004
- Sandi, C. (2004). Stress, cognitive impairment and cell adhesion molecules. *Nat. Rev. Neurosci.* 5, 917–930. doi: 10.1038/nrn1555
- Sharp, T. (2013). Molecular and cellular mechanisms of antidepressant action. *Curr. Top. Behav. Neurosci.* 14, 309–325. doi: 10.1007/7854\_2012\_216
- Tiraboschi, E., Guirado, R., Greco, D., Auvinen, P., Maya-Vetencourt, J. F., Maffei, L., et al. (2013). Gene expression patterns underlying the reinstatement of plasticity in the adult visual system. *Neural Plast.* 2013:605079. doi: 10.1155/2013/605079
- Varea, E., Blasco-Ibáñez, J., Gómez-Climent, M. A., Castillo-Gómez, E., Crespo, C., Martínez-Guijarro, F., et al. (2007). Chronic fluoxetine treatment increases the expression of PSA-NCAM in the medial prefrontal cortex. *Neuropsychopharmacology* 32, 803–812. doi: 10.1038/sj.npp.1301183
- Vialou, V., Thibault, M., Kaska, S., Cooper, S., Gajewski, P., Eagle, A., et al. (2015). Differential induction of FosB isoforms throughout the brain by fluoxetine and chronic stress. *Neuropharmacology* 99, 28–37. doi: 10.1016/j.neuropharm.2015.07.005
- Webber, A. L., and Wood, J. (2005). Amblyopia: prevalence, natural history, functional effects and treatment. *Clin. Exp. Optom.* 88, 365–375. doi: 10.1111/j.1444-0938.2005.tb05102.x
- Wiesel, T., and Hubel, D. (1963). Single-cell responses in striate cortex of kittens deprived of vision in one eye. *J. Neurophysiol.* 26, 1003–1017.
- Yazaki-Sugiyama, Y., Kang, S., Câteau, H., Fukai, T., and Hensch, T. K. (2009). Bidirectional plasticity in fast-spiking GABA circuits by visual experience. *Nature* 462, 218–221. doi: 10.1038/nature08485

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

EC is an advisor and shareholder in Herantis Pharma, Inc.

Copyright © 2016 Guirado, La Terra, Bourguignon, Carceller, Umemori, Sipilä, Nacher and Castrén. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution and reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.