

Asymmetric Synthesis of N-Fmoc-(*S*)-7-aza-tryptophan via Alkylation of Chiral Nucleophilic Glycine Equivalent

Yupiao Zou,^[a] Ryosuke Takeda,^[b] Jianlin Han,*^[a] Hiroyuki Konno,^[c] Hiroki Moriwaki,^[b] Hidenori Abe,*^[b] Kunisuke Izawa,^[b] and Vadim A. Soloshonok*^[d, e]

In Memory of Professor Ferenc Fülöp.

Ni(II)-complexes, derived from glycine Schiff bases with chiral tridentate ligands, have been used as powerful tools for the synthesis of structurally diverse tailor-made amino acids. In this manuscript, asymmetric alkylation reaction between chiral nucleophilic glycine derived Ni-complex and 3-(chloromethyl)-1*H*-pyrrolo[2,3-*b*]pyridine has been developed under convenient conditions, which affords the corresponding alkylated Ni-complex in 74% yield and excellent diastereoselectivity (only one isomer). This reaction features convenient conditions and completely controlled diastereoselectivity, which provides a highly valuable approach for asymmetric synthesis of 7-aza-tryptophan.

Amino acids (AAs) belong to the most ubiquitous class of naturally occurring compounds,^[1] which have been used as an

important type of structural units in the design of modern drugs due to several advantageous features, such as high structural diversity, wide scope of functional groups, good solubility in aqueous media, and few safety concerns.^[2] Currently, a number of small-molecule, peptidomimetics, and peptide containing residues of tailor-made AAs or their derivatives serve as key structural features in many blockbuster drugs.^[3] Among these natural and non-natural AAs, 7-aza-tryptophan occupies an important position, which has been widely used as biological fluorescent probes,^[4] antiplasmodial compounds,^[5] and inhibitors of checkpoint kinase 1.^[6,7] Although there have been several reports on the synthesis of aza-tryptophanes,^[8] the synthesis of 7-aza-tryptophan still remained less developed, in particular in the asymmetric mode. One of the first asymmetric methods was based on alkylation reaction between (1*R,4R*)-camphor imine and *tert*-butylglycinate, which proceeded with poor chemical yield (Scheme 1a).^[9] Chemoenzymatic synthesis represents an alternative strategy for the preparation of optical pure 7-aza-tryptophan using *Aspergillus* genus acylase,^[10] L-amino acid oxidase^[11] or tryptophan synthase from *Salmonella typhimurium*^[12] as biocatalyst

[a] Y. Zou, Prof. Dr. J. Han

Jiangsu Co – Innovation Center of Efficient Processing and Utilization of Forest Resources,
International Innovation Center for Forest Chemicals and Materials, College
of Chemical Engineering
Nanjing Forestry University
159 Longpan Road, 210037 Nanjing, China
E-mail: hanjl@njfu.edu.cn
http://www.hg.njfu.edu.cn

[b] Dr. R. Takeda, Dr. H. Moriwaki, Dr. H. Abe, Dr. K. Izawa
Hamari Chemicals Ltd.,
1-4-29 Kunijima, Higashi-Yodogawa-ku, 533-0024 Osaka, Japan
E-mail: hidenori-abe@hamari.co.jp

[c] Prof. Dr. H. Konno
Department of Biological Engineering, Graduate School of Science and
Engineering
Yamagata University
Yonezawa, 992-8510 Yamagata, Japan

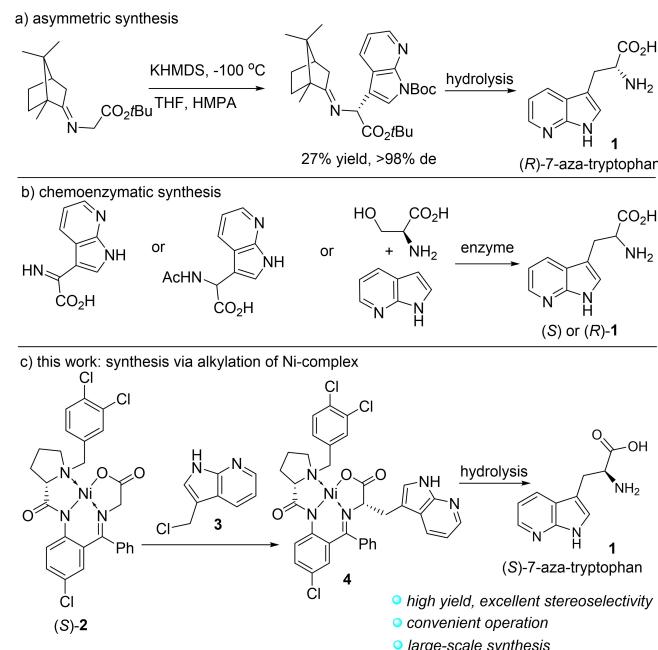
[d] Prof. Dr. V. A. Soloshonok
Department of Organic Chemistry I, Faculty of Chemistry
University of the Basque Country UPV/EHU
Paseo Manuel Lardizábal 3, 20018 San Sebastián, Spain
E-mail: vadym.soloshonok@ehu.es
http://www.ikerbasque.net/vadym.soloshonok

[e] Prof. Dr. V. A. Soloshonok
IKERBASQUE, Basque Foundation for Science
Alameda Urquijo 36–5, Plaza Bizkaia, 48011 Bilbao, Spain

Supporting information for this article is available on the WWW under
https://doi.org/10.1002/ejoc.202100485

Part of the Special Collection in Memory of Ferenc Fülöp.

© 2021 The Authors. European Journal of Organic Chemistry published by Wiley-VCH GmbH. This is an open access article under the terms of the Creative Commons Attribution Non-Commercial NoDerivs License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made.



Scheme 1. Asymmetric synthesis of 7-aza-tryptophan.

(Scheme 1b). Thus, there is a room for the development of convenient, efficient, and easy scalable methods for preparation of chiral 7-aza-tryptophan.

Consistent with our long-standing interest in preparation of tailor-made AAs, in particular phosphorus-^[13] and fluorine-containing derivatives,^[14] we were developing general approach for asymmetric synthesis of AAs via Ni(II) complexes of chiral Schiff bases.^[15] Typically, chiral ligands (*S*)-5–7 are used for the preparation of nucleophilic glycine-based Ni-complex for the introduction of the desired side-chain(s) (Figure 1).^[16] Several types of reactions of these nucleophilic Ni-complexes have been developed, such as alkyl halide alkylation^[17] aldol,^[18] Mannich^[19] and Michael^[20] addition reactions. Based on the literature data and our own experience in application of Ni(II)-complexes of Schiff bases for the synthesis of "Tailor-Made Amino Acids™",^[21,22] we eversion that reaction between Ni-complex and suitable alkyl halide would provide a method to chiral 7-aza-tryptophan. Herein, report an asymmetric method for the synthesis of chiral 7-aza-tryptophan via an alkylation of chiral Ni-complex with corresponding alkyl chloride (Scheme 1c).

Ligand (*S*)-6 derived glycine Ni-complex **2** was chosen as the starting material for this alkylation reaction (Scheme 2), which usually shows enhanced control in stereoselectivity due

to a parallel displaced type of aromatic interactions between the selectively chlorinated o-amino-benzophenone and Pro *N*-benzyl rings.^[23] The desired alkyl chloride **3**, Boc-protected 3-(chloromethyl)-1*H*-pyrrolo[2,3-*b*]pyridine, was prepared with 1*H*-pyrrolo[2,3-*b*]pyridine-3-carbaldehyde as the starting reagent (for details, see SI file).

Having prepared alkyl chloride **3**, we then studied its alkylation reaction with Ni(II)-complex **2** (Scheme 2). To find an optimal reaction condition to achieve high selectivity and clean conversion to the desired Ni (II) complex **4a**, several screens were carried out. The optimization study led to the very specific condition using DMSO as a reaction solvent, which had rarely been used in the past applications of this methodology.^[21] Using DMSO showed a significant difference in purity of the product **4a** during the reaction, exhibiting the high diastereomeric selectivity as well as the lowering impurities formation. After more refining studies were conducted to find an optimal stoichiometry of base and alkyl chloride minimizing the key bis-alkylation byproduct, we found that the reaction with DMSO as a solvent in the presence of NaOH (1.5 equiv.) and alkyl chloride (0.8 equiv.) provided the best yield and diastereoselectivity. To address this question, we added the following sentence: The beneficial effect of DMSO on the reaction outcome is likely multifactorial. However, what could be clearly noticed is a significantly reduced amount of the oxidation products that usually observed under strongly basic reaction conditions.^[24]

During the workup procedure, surprisingly, the Boc group was partially removed during the reaction even under basic conditions providing a mixture of compounds (**4a** and **4b**). Thus, we developed a one-pot procedure for the synthesis of pure alkylated Ni-complex **4a**. The Boc remaining protected product **4b** was completely converted to compound **4a** by the treatment of the crude reaction mixture with MeOH.

Thus, the reaction was conducted in 2.0 g scale of (*S*)-2, 0.8 equiv. of alkyl chloride **3**, 1.5 equiv. of NaOH, in DMSO (40 mL) under nitrogen atmosphere at room temperature for 1.5 h. The resulted reaction mixture was treated with MeOH (40 mL) and stirred at 20 °C for 2 h, and then heated to 60 °C for 1 h, which afforded the corresponding product **4a** in 74% yield and completely controlled stereochemical outcome.

The alkylated Ni (II) complex (*S,2S*)-**4a** was then disassembled in the presence of 3 N HCl (aqueous) with 1,2-dimethoxyethane (DME) as a solvent, which afforded the corresponding free amino acid, (*S*)-7-aza-tryptophan (1) hydrochloric acid salt in 95% yield (Scheme 3). DME was chosen as a solvent, instead of methanol, although methanol has been generally employed for this step.^[22] This is because the use of methanol as solvent for this disassembly procedure was found to cause undesired esterification. The reaction proceeded smoothly, and the isolation of the amino acid was successfully carried out by filtering off most of the chiral ligand precipitated after the reaction and partitioning between water and dichloromethane to sequester the residual ligand into organics. C18 flash column chromatography was utilized to remove the residual Ni²⁺ prior to the following Fmoc protection. It should be pointed out the C18 flash column method was used during this trace experiment. This is proved to be the most straightfor-

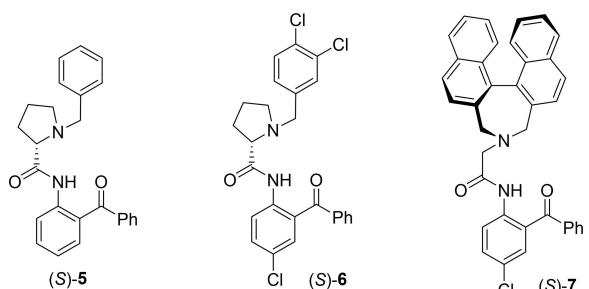
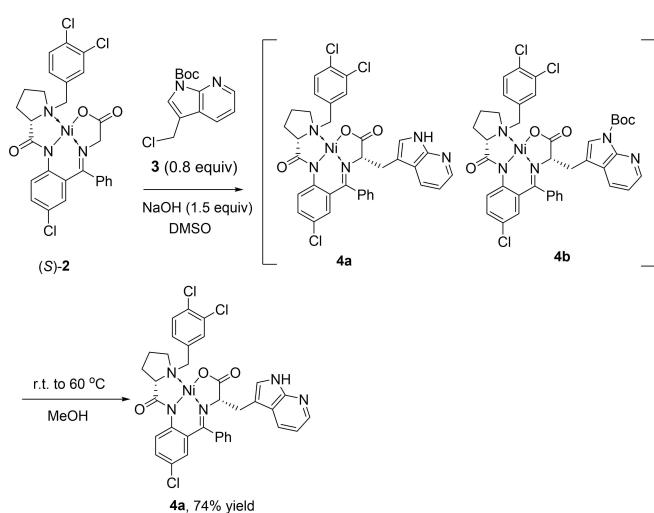
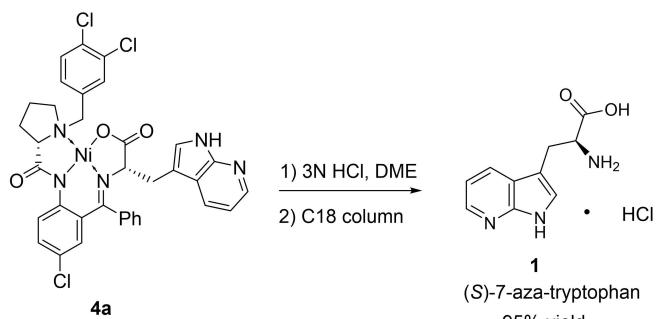


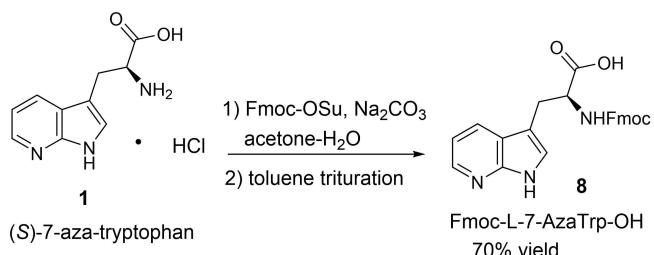
Figure 1. Chiral tridentate ligands **5–7**.



Scheme 2. Alkylation of chiral glycine-Ni(II) complex **2** with compound **3**.



Scheme 3. Disassembly of Ni-complex 4a.



Scheme 4. Fmoc protection of (S)-1.

ward for small scale, which worked well to provide the desired amino acid **1** in good yield and high purity. It should be emphasized that chiral ligand (*S*)-**6**, as well as Ni(II), can be recycled and reused for continuous production of this and other types of AAs, as illustrated in the recent publications.^[22,25]

The Fmoc protection was carried out employing general conditions using Fmoc-OSu in the presence of sodium carbonate (Scheme 4). The reaction proceeded smoothly and furnished the desired Fmoc-L-7-AzaTrp-OH (**8**) in 70% yield. It should be mentioned that the residual Ni²⁺ seems to complicate the Fmoc reaction and care must be taken to remove Ni²⁺ completely before this step. The crystalline product **8** was easy to isolate from the reaction. For the purification, the hydrolyzed impurity derived from unreacted Fmoc-OSu was effectively sequestered by the trituration with toluene. The synthesized compound **1** could be easily obtained with a high chemical purity as measured by ¹H-NMR and HPLC.

In summary, the first total synthesis of Fmoc-L-7-AzaTrp-OH via tridentate chiral auxiliary derived Ni(II)-complexes methodology was successfully accomplished, which could be operated under convenient conditions resulting in an overall 49% yield (three-step). In particular, excellent diastereoselectivity was obtained for the alkylation step (only one isomer).

Acknowledgements

We gratefully acknowledge the financial support from the National Natural Science Foundation of China (No. 21761132021) and IKERBASQUE, Basque Foundation for Science.

Conflict of Interest

The authors declare no conflict of interest.

Keywords: Aza-tryptophanes · Asymmetric synthesis · Chiral tridentate ligands · Ni catalysis · Tailor-Made Amino Acids™ · Schiff bases

- [1] a) H. B. Vickery, C. L. Schmidt, *Chem. Rev.* **1931**, *9*, 169–318; b) M. A. T. Blaskovich, *J. Med. Chem.* **2016**, *59*, 10807–10836; c) Asymmetric Synthesis and Application of α -Amino Acids, ACS Symposium Series #1009; V. A. Soloshonok, K. Izawa, Eds., Oxford University Press: **2009**; d) A. Stevenazzi, M. Marchini, G. Sandrone, B. Vergani, M. Lattanzio, *Bioorg. Med. Chem. Lett.* **2014**, *24*, 5349–5356.
- [2] a) B. Vig, J. Rautio, *Ther. Delivery* **2011**, *2*, 959–962; b) P. Conti, L. Tamborini, A. Pinto, A. Blondel, P. Minoprio, A. Mozzarelli, C. De Micheli, *Chem. Rev.* **2011**, *111*, 6919–6946.
- [3] a) J. Han, A. E. Sorochinsky, T. Ono, V. A. Soloshonok, *Curr. Org. Synth.* **2011**, *8*, 281–294; b) H. Mei, J. Han, K. D. Klika, K. Izawa, T. Sato, N. A. Meanwell, V. A. Soloshonok, *Eur. J. Med. Chem.* **2020**, *186*, 111826; c) V. A. Soloshonok, A. E. Sorochinsky, *Synthesis* **2010**, 2319–2344; d) H. Mei, J. Han, S. White, D. J. Graham, K. Izawa, T. Sato, S. Fustero, N. A. Meanwell, V. A. Soloshonok, *Chem. Eur. J.* **2020**, *26*, 11349; e) J. Han, A. M. Remete, L. S. Dobson, L. Kiss, K. Izawa, H. Moriawaki, V. A. Soloshonok, D. O'Hagan, *J. Fluorine Chem.* **2020**, *239*, 109639; f) Z. Yin, W. Hu, W. Zhang, H. Konno, H. Moriawaki, K. Izawa, J. Han, V. A. Soloshonok, *Amino Acids* **2020**, *52*, 1227; g) J. Liu, J. Han, K. Izawa, T. Sato, S. White, N. A. Meanwell, V. A. Soloshonok, *Eur. J. Med. Chem.* **2020**, *208*, 112736; h) H. Mei, A. M. Remete, Y. Zou, H. Moriawaki, S. Fustero, L. Kiss, V. A. Soloshonok, J. Han, *Chin. Chem. Lett.* **2020**, *31*, 2401; i) H. Mei, J. Han, S. Fustero, M. Medio-Simon, D. M. Sedgwick, C. Santi, R. Ruzziconi, V. A. Soloshonok, *Chem. Eur. J.* **2019**, *25*, 11797; j) J. Han, H. Konno, T. Sato, V. A. Soloshonok, K. Izawa, *Eur. J. Med. Chem.* **2021**, *220*, 113448; k) H. Mei, J. Han, S. White, D. J. Graham, K. Izawa, T. Sato, S. Fustero, N. A. Meanwell, V. A. Soloshonok, *Chem. Eur. J.* **2020**, *26*, 11349–11390; l) W. Qiu, X. Gu, V. A. Soloshonok, M. D. Carducci, V. J. Hruby, *Tetrahedron Lett.* **2001**, *42*, 145–148; m) M. Shevchuk, Q. Wang, R. Pajkert, J. Xu, H. Mei, G. V. Röschenthaler, J. Han, *Adv. Synth. Catal.* **2021**, doi.org/10.1002/adsc.202001464.
- [4] a) V. De Filippis, S. De Boni, E. De Dea, D. Dalzoppo, C. Grandi, A. Fontana, *Protein Sci.* **2004**, *13*, 1489–1502; b) W. C. Chao, T. H. Chiang, P. D. Chaudhari, L. J. Lin, J. F. Lu, B. C. Hong, J. S. Wang, T. C. Lin, J. Y. Shen, P. T. Chou, *Bioorg. Chem.* **2018**, *81*, 504–511; c) L. D. Newton, S. I. Pascu, R. M. Tyrrell, I. M. Eggleston, *Org. Biomol. Chem.* **2019**, *17*, 467–471; d) M. G. Hoesl, M. Larregola, H. Cui, N. Budisa, *J. Pept. Sci.* **2010**, *16*, 589–595.
- [5] T. B. Beghyn, J. Charton, F. Leroux, G. Laconde, A. Bourin, P. Cos, L. Maes, B. Deprez, *J. Med. Chem.* **2011**, *54*, 3222–3240.
- [6] L. Gazzard, B. Appleton, K. Chapman, H. Chen, K. Clark, J. Drobnick, S. Goodacre, J. Halladay, J. Lyssikatos, S. Schmidt, S. Sideris, C. Wiesmann, K. Williams, P. Wu, I. Yen, S. Malek, *Bioorg. Med. Chem. Lett.* **2014**, *24*, 5704–5709.
- [7] a) M. Katragadda, J. D. Lambris, *Protein Expre. Purif.* **2006**, *47*, 289–295; b) J. Broos, E. Gabellieri, E. Beimans-Odehinkel, G. B. Strambini, *Protein Sci.* **2003**, *12*, 1991–2000.
- [8] a) F. Crestey, V. Collot, S. Stiebing, S. Rault, *Tetrahedron* **2006**, *62*, 7772–7775; b) F. Crestey, V. Collot, S. Stiebing, J. F. Lohier, J. S. O. Santos, S. Rault, *Tetrahedron Lett.* **2007**, *48*, 2457–2460; c) M. Jeanty, F. Suzenet, G. Guilliaumet, *J. Org. Chem.* **2008**, *73*, 7390–7393.
- [9] R. Sanchez-Obregon, A. G. Fallis, *Can. J. Chem.* **1992**, *70*, 1531–1536.
- [10] L. Lecointe, V. Rolland-Fulcrand, M. L. Rou mestant, Ph. Viallefont, J. Martinez, *Tetrahedron: Asymmetry* **1998**, *9*, 173–1758.
- [11] S. Nakano, Y. Minamino, F. Hasebe, S. Ito, *ACS Catal.* **2019**, *9*, 10152–10158.
- [12] M. Winn, D. Francis, J. Micklefield, *Angew. Chem. Int. Ed.* **2018**, *57*, 6830–6833; *Angew. Chem.* **2018**, *130*, 6946–6949.
- [13] a) G. V. Röschenthaler, V. P. Kukhar, I. B. Kulik, M. Y. Belik, A. E. Sorochinsky, E. B. Rusanov, V. A. Soloshonok, *Tetrahedron Lett.* **2012**, *53*, 539–542; b) K. V. Turcheniuk, K. O. Poliashko, V. P. Kukhar, A. B. Rozhenko, V. A. Soloshonok, A. E. Sorochinsky, *Chem. Commun.* **2012**, *48*, 11519–11521.

- [14] a) P. Bravo, A. Farina, V. P. Kukhar, A. L. Markovsky, S. V. Meille, V. A. Soloshonok, A. E. Sorochinsky, F. Viani, M. Zanda, C. Zappala, *J. Org. Chem.* **1997**, *62*, 3424–3425; b) V. A. Soloshonok, T. Hayashi, *Tetrahedron: Asymmetry* **1994**, *5*, 1091–1094; c) V. A. Soloshonok, A. G. Kirilenko, V. P. Kukhar, G. Resnati, *Tetrahedron Lett.* **1993**, *34*, 3621–3624.
- [15] a) S. Zhou, J. Wang, X. Chen, J. L. Aceña, V. A. Soloshonok, H. Liu, *Angew. Chem. Int. Ed.* **2014**, *53*, 7883–7886; *Angew. Chem.* **2014**, *126*, 8017–8020; b) R. Takeda, A. Kawamura, A. Kawashima, T. Sato, H. Moriwaki, K. Izawa, K. Akaji, S. Wang, H. Liu, J. L. Aceña, V. A. Soloshonok, *Angew. Chem. Int. Ed.* **2014**, *53*, 12214–12217; *Angew. Chem.* **2014**, *126*, 12410–12413; c) Y. Nian, J. Wang, S. Zhou, S. Wang, H. Moriwaki, A. Kawashima, V. A. Soloshonok, H. Liu, *Angew. Chem. Int. Ed.* **2015**, *54*, 12918–12922; d) N. Wang, J. Xu, H. Mei, H. Moriwaki, K. Izawa, V. A. Soloshonok, J. Han, *Chin. J. Org. Chem.* **2021**, doi: 10.6023/cjoc202102043.
- [16] a) J. Han, T. T. Romoff, H. Moriwaki, H. Konno, V. A. Soloshonok, *ACS Omega* **2019**, *4*, 18942–18947; b) T. T. Romoff, A. B. Palmer, N. Mansour, C. J. Creighton, T. Miwa, Y. Ejima, H. Moriwaki, V. A. Soloshonok, *Org. Process Res. Dev.* **2017**, *21*, 732–739.
- [17] a) T. K. Ellis, V. M. Hochla, V. A. Soloshonok, *J. Org. Chem.* **2003**, *68*, 4973–4976; b) V. A. Soloshonok, X. Tang, V. J. Hruby, L. V. Meervelt, *Org. Lett.* **2001**, *3*, 341–343.
- [18] V. A. Soloshonok, V. P. Kukhar, S. V. Galushko, N. Y. Svistunova, D. V. Avilov, N. A. Kuzmina, N. I. Raevski, Y. T. Struchkov, A. P. Pisarevsky, Y. N. Belokon, *J. Chem. Soc. Perkin Trans. 1* **1993**, *1993*, 3143–3155.
- [19] A. Kawamura, H. Moriwaki, G. V. Röschenthaler, K. Kawada, J. L. Aceña, V. A. Soloshonok, *J. Fluorine Chem.* **2015**, *171*, 67–72.
- [20] a) V. A. Soloshonok, D. V. Avilov, V. P. Kukhar, L. V. Meervelt, N. Mischenko, *Tetrahedron Lett.* **1997**, *38*, 4903–4904; b) V. A. Soloshonok, C. Cai, V. J. Hruby, *Tetrahedron Lett.* **2000**, *41*, 135–139; c) T. Yamada, T. Okada, K. Sakaguchi, Y. Ohfune, H. Ueki, V. A. Soloshonok, *Org. Lett.* **2006**, *8*, 5625–5628.
- [21] a) Y. Zou, J. Han, A. S. Saghyan, A. F. Mkrtchyan, H. Konno, H. Moriwaki, K. Izawa, V. A. Soloshonok, *Molecules* **2020**, *25*, 2739; b) Y. Wang, X. Song, J. Wang, H. Moriwaki, V. A. Soloshonok, H. Liu, *Amino Acids* **2017**, *49*, 1487–1520.
- [22] a) Y. Shigeno, J. Han, V. A. Soloshonok, H. Moriwaki, W. Fujiwara, H. Konno, *Chirality* **2021**, *33*, 115–123; b) B. Fu, R. Takeda, Y. Zou, H. Konno, H. Moriwaki, H. Abe, J. Han, K. Izawa, V. A. Soloshonok, *Chirality* **2020**, *32*, 1354–1360; c) T. T. Romoff, B. G. Ignacio, N. Mansour, A. B. Palmer, C. J. Creighton, H. Abe, H. Moriwaki, J. Han, H. Konno, V. A. Soloshonok, *Org. Process Res. Dev.* **2020**, *24*, 294–300; d) Y. Tokairin, Y. Shigeno, J. Han, G. V. Röschenthaler, H. Konno, H. Moriwaki, V. A. Soloshonok, *ChemistryOpen* **2020**, *9*, 93–96; e) H. Mei, J. Han, R. Takeda, T. Sakamoto, T. Miwa, Y. Minamitsuji, H. Moriwaki, H. Abe, V. A. Soloshonok, *ACS Omega* **2019**, *4*, 11844–11851; f) Z. Yin, H. Moriwaki, H. Abe, T. Miwa, J. Han, V. A. Soloshonok, *ChemistryOpen* **2019**, *8*, 701.
- [23] Y. Nian, J. Wang, H. Moriwaki, V. A. Soloshonok, H. Liu, *Dalton Trans.* **2017**, *46*, 4191–4198.
- [24] a) V. A. Soloshonok, H. Ueki, *J. Am. Chem. Soc.* **2007**, *129*, 2426–2427; b) V. A. Soloshonok, T. Ono, H. Ueki, N. Vanthuyne, T. S. Balaban, J. Bürck, H. Fliegl, W. Klopper, J. V. Naubron, T. T. Tam, A. F. Drake, C. Roussel, *J. Am. Chem. Soc.* **2010**, *132*, 10477–10483.
- [25] a) H. Mei, Z. Yin, T. Miwa, H. Moriwaki, H. Abe, J. Han, V. A. Soloshonok, *Symmetry* **2019**, *11*, 578; b) H. Mei, T. Hiramatsu, R. Takeda, H. Moriwaki, H. Abe, J. Han, V. A. Soloshonok, *Org. Process Res. Dev.* **2019**, *23*, 629–634.

Manuscript received: April 19, 2021

Revised manuscript received: May 6, 2021

Accepted manuscript online: May 10, 2021