

Linking Plasma Amyloid Beta and Neurofilament Light Chain to Intracortical Myelin Content in Cognitively Normal Older Adults

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Evidence suggests that lightly myelinated cortical regions are vulnerable to aging and Alzheimer's disease (AD). However, it remains unknown whether plasma markers of amyloid and neurodegeneration are related to deficits in intracortical myelin content, and whether this relationship, in turn, is associated with altered patterns of restingstate functional connectivity (rs-FC). To shed light into these questions, plasma levels of amyloid- β fragment 1–42 (A β_{1-42}) and neurofilament light chain (NfL) were measured using ultra-sensitive single-molecule array (Simoa) assays, and the intracortical myelin content was estimated with the ratio T1-weighted/T2-weighted (T1w/T2w) in 133 cognitively normal older adults. We assessed: (i) whether plasma $A\beta_{1-42}$ and/or NfL levels were associated with intracortical myelin content at different cortical depths and (ii) whether cortical regions showing myelin reductions also exhibited altered rs-FC patterns. Surface-based multiple regression analyses revealed that lower plasma $A\beta_{1-42}$ and higher plasma NfL were associated with lower myelin content in temporoparietal-occipital regions and the insular cortex, respectively. Whereas the association with $A\beta_{1-42}$ decreased with depth, the NfL-myelin relationship was most evident in the innermost layer. Older individuals with higher plasma NfL levels also exhibited altered rs-FC between the insula and medial orbitofrontal cortex. Together, these findings establish a link between plasma markers of amyloid/neurodegeneration and intracortical myelin content in cognitively normal older adults, and support the role of plasma NfL in boosting aberrant FC patterns of the insular cortex, a central brain hub highly vulnerable to aging and neurodegeneration.

Keywords: aging, Alzheimer's disease, intracortical myelin, functional connectivity, blood biomarkers, amyloidbeta, neurofilament light

INTRODUCTION

Aging is the major risk factor for Alzheimer's disease (AD), but the reasons why aging increases susceptibility to AD are poorly understood. One contributing factor may be the perturbation of myelin-related genes (Mathys et al., 2019) that eventually leads to widespread degeneration of myelin sheaths. This phenomenon results in slowing of conduction velocity along nerve fibers, modifying the timing of network oscillations, and ultimately affecting

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Fernandez-Alvarez M, Atienza M, Zallo F, Matute C, Capetillo-Zarate E and Cantero JL (2022) Linking Plasma Amyloid Beta and Neurofilament Light Chain to Intracortical Myelin Content in Cognitively Normal Older Adults. Front. Aging Neurosci. 14:896848. doi: 10.3389/fnagi.2022.896848 functional connections within neural circuits (Peters, 2002). The efficiency of remyelination also declines with age, likely due to the limited regenerative capacity of oligodendrocyte progenitor cells (Irvine and Blakemore, 2006). Consequently, lightly myelinated axons become more vulnerable to irreversible degeneration during aging, favoring cognitive decline (Wang et al., 2020) and the spread of AD pathology before the onset of symptoms (Mitew et al., 2010; Braak and Del Tredici, 2015; Couttas et al., 2016; Dean et al., 2017; Luo et al., 2019).

New ultrasensitive quantitative technologies allow the identification of proteins in blood at subfemtomolar concentrations (Rissin et al., 2010; Yang et al., 2017), opening new avenues for the development of blood biomarkers capable of detecting individuals at risk for cognitive decline and AD. Accumulated evidence suggests that lower levels of plasma amyloid- β fragment 1–42 (A β_{1-42}) are associated with accelerated aging and AD. Accordingly, low levels of plasma $A\beta_{1-42}$ at baseline have shown to increase the risk of cognitive decline (Seppälä et al., 2010), mild cognitive impairment (MCI) (Rembach et al., 2014) and AD (Rembach et al., 2014; Chouraki et al., 2015; Hilal et al., 2018; de Wolf et al., 2020). Moreover, higher plasma concentrations of neurofilament light chain (NfL), the main cytoskeletal structure of myelinated axons, have been associated with increased brain AB load in cognitively unimpaired older adults (Chatterjee et al., 2018; Benedet et al., 2020), and have shown to predict cortical thinning and subsequent cognitive impairment in the preclinical stage of both familial and sporadic AD (Preische et al., 2019; Lee et al., 2022). However, it remains largely unknown whether plasma markers of amyloid and neurodegeneration bear any relation to brain myelin content in cognitively normal older individuals, which may be relevant to establish surrogate markers of vulnerability to AD.

Although myelination is a prominent feature of the subcortical white matter (WM), the gray matter (GM) of the cerebral cortex also contains a substantial amount of myelinated axons unevenly distributed in layers (Nieuwenhuys, 2013). Thus, myelin density is higher in deeper than in superficial cortical layers and in sensorimotor than in association regions (Nave and Werner, 2014). This particular distribution of cortical myelin may partially account for the functional organization of the neocortex (Beul et al., 2017; Huntenburg et al., 2017). Moreover, evidence has shown that subtle changes in myelin have meaningful effects on neuronal network function (Waxman, 1980; Felts et al., 1997; Pajevic et al., 2014). Therefore, aging-related patterns of rs-FC may be driven by cortical regions showing myelin deficits, which, in turn, may be conditioned by plasma markers of amyloid and neurodegeneration.

Here, we specifically addressed these questions by investigating whether the T1w/T2w ratio, considered as a proxy of myelin content in the cortical GM (Glasser and Van Essen, 2011), is associated with plasma levels of $A\beta_{1-42}$ and NfL in cognitively normal older adults, and whether this relationship varies with cortical depth. Next, we assessed whether those cortical areas showing plasma measurements-related myelin deficits also exhibited alterations in their pattern of rs-FC, which in turn may account for the variability in cognitive functioning. Therefore, the present study sought to test three inter-related

hypotheses. As amyloid pathology has been previously associated, either direct or indirectly, with myelin damage (Desai et al., 2010; Mitew et al., 2010; Schmued et al., 2013; Dean et al., 2017; Luo et al., 2019; Chen et al., 2021), we first hypothesized that lower levels of plasma $A\beta_{1-42}$ will be associated with lower intracortical myelin content in cognitively normal older adults. The relationship between aging-related axonal damage and myelin breakdown is likely strengthened by dysfunctional oligodendrocytes, which supply energy for axonal metabolism (Mot et al., 2018). Accordingly, our second prediction was that higher levels of plasma NfL will be accompanied by lower myelin content, and that this relationship will become more evident in inner cortical layers, where oligodendrogenesis is most impaired (Orthmann-Murphy et al., 2020; Xu et al., 2021). Finally, and given the critical role of myelin in neuronal communication and fine-tuning of neuronal circuits (Fields, 2015; Monje, 2018), we postulated that cortical regions showing lower myelin content related to plasma levels of $A\beta_{1-42}$ and/or NfL will also show altered patterns of rs-FC.

MATERIALS AND METHODS

Participants

One hundred thirty-three cognitively normal older adults participated in the study (65 \pm 5.9 years; range: 54-76 years; 81 females). They were recruited from senior citizen's associations, health-screening programs, and hospital outpatient services. All of them underwent neurological and neuropsychological assessment to discard both the presence of dementia and objective cognitive impairment. Individuals with medical conditions that affect brain structure or function (e.g., cerebrovascular disease, epilepsy, head trauma, history of neurodevelopmental disease, alcohol abuse, hydrocephalus, and/or intracranial mass) were not included in the study. Participants met the following criteria: (i) normal global cognitive status in the Mini-Mental State Examination (scores \geq 26); (ii) normal cognitive performance in the neuropsychological tests relative to appropriate reference values for age and education level; (iii) global score of 0 (no dementia) in the Clinical Dementia Rating; (iv) functional independence as assessed by the Spanish version of the Interview for Deterioration in Daily Living Activities (Böhm et al., 1998); (v) scores ≤ 5 (no depression) in the short form of the Geriatric Depression Scale (Sheikh and Yesavage, 1986); and (vi) not be taking medications that affected cognition, sleep, renal or hepatic function. All participants gave informed consent to the experimental protocol approved by the Ethical Committee for Clinical Research of the Junta de Andalucía according to the principles outlined in the Declaration of Helsinki. Table 1 contains sample characteristics.

Neuropsychological Assessment

All participants received neuropsychological assessment. They were administered with the following tests: the Spanish version of the Memory Binding Test (MBT) (Gramunt et al., 2016); the short form of the Boston Naming Test (BNT); the semantic and phonological fluency tests based on the "Animal" and letter

Age	65 ± 5.9			
Sex (F/M)	81/52			
Education years	11.7 ± 4.5			
ApoE4 (yes/no)	31/102			
MMSE	28.6 ± 1.3			
Memory Binding Test				
Total free recall	16.2 ± 4.6			
Pairs in free recall	5.3 ± 2.6			
Total paired recall	27.1 ± 4.9			
Paired recall pairs	9.8 ± 4.1			
Total delayed free recall	15.1 ± 5.0			
Pairs in delayed free recall	5.8 ± 3.9			
Total delayed paired recall	28.9 ± 5.4			
Boston Naming Test	12.1 ± 2.1			
Phonological fluency	15.7 ± 4.4			
Semantic fluency	22.0 ± 17.0			
Trail Making Test-A	47.0 ± 21.5			
Trail Making Test-B	119.5 ± 67.7			
Tower of London	319.3 ± 113.2			
Plasma Aβ ₁₋₄₂ (pg/ml)	12.1 ± 5.1 (2.4 – 24)			
Plasma NfL (pg/ml)	12.9 ± 7.2 (2.3 – 39.6			

Results are expressed as mean \pm SD, unless otherwise stated. F/M, females/males; MMSE, Mini Mental State Examination; NfL: neurofilament light.

"P" naming tasks; the two forms of the Trail Making Test (TMT-A and TMT-B); and the Tower of London (TL). All neuropsychological scores were z transformed. In the case of TMT-A and TMT-B, we used the inverse z-values, while in the case of the MBT and TL we first computed a composite measure by summing the z-scores of the different scores. We then applied principal component analysis to obtain the Spearman's "g" factor as an index of global cognitive function. This analysis was done with R Statistical Software v3.0.1 (R Foundation for Statistical Computing, Vienna, Austria) using the prcomp function. We only retained the first component (eigenvalue 3.4), which explained 53.7% of variance in the data, due to the contribution of MBT (24.2), BNT (24.2), phonological fluency (24.2), semantic fluency (3.2), TMT-A (8.6), TMT-B (9.5), and TL (6.0). The standardized residuals were used to obtain the latent variable g (factor loading MBT: 0.92; BNT: 0.92; phonological fluency: 0.92; semantic fluency: 0.33; TMT-A: 0.55; TMT-B: 0.57; TL: 0.46).

Measurements of Plasma $A\beta_{1-42}$ and NfL

Fasting blood samples were taken at 9:00-10:00 AM in all participants to control for potential circadian effects. Briefly, venous blood samples were collected in 10 ml dipotassium ethylene diamine tetraacetic acid (EDTA) coated tubes (BD Diagnostics), and immediately centrifuged (1,989 g) at 4°C for 5 min. Supernatant plasma was aliquoted into polypropylene tubes containing 300 μ l of plasma mixed with 10 μ l of a protease inhibitor cocktail (cOmplete Ultra Tablets mini, Roche), and stored at -80°C until analysis. Plasma samples used in the present study were not previously thawed.

Plasma $A\beta_{1-42}$ and NfL levels were measured on the ultrasensitive single-molecule array (Simoa) HD-1 analyzer platform (Quanterix, MA, United States) following the manufacturer's instructions. The A β_{1-42} Simoa 2.0 assay (Cat. No. 101664) and the NF-light Simoa assay advantage kits (Cat. No. 103186) were purchased from Quanterix. These assays measure A β_{1-42} and NfL levels in human plasma with a detection limit of 0.044 and 0.038 pg/ml, respectively. Two quality control samples were run on each plate for each analyte. Plasma A β_{1-42} and NfL determinations were run in duplicates, and the average of the two measurements (pg/ml) was used for statistical analysis. Samples with coefficients of variation higher than 20% were excluded.

Magnetic Resonance Imaging Acquisition

Structural and functional magnetic resonance imaging (MRI) scans were performed on a 3T Philips Ingenia MRI scanner, equipped with a 32-channel radio-frequency (RF) receive head coil and body RF transmit coil (Philips, Best, Netherlands). The following MRI sequences were acquired in the same session: (i) 3D T1-weighted (T1w) magnetization prepared rapid gradient echo (MPRAGE) in the sagittal plane: repetition time (TR)/echo time (TE) = 2,600 ms/4.7 ms, flip angle (FA) = 9° , acquisition matrix = 384×384 , voxel resolution = 0.65 mm^3 isotropic, resulting in 282 slices without gap between adjacent slices; (ii) 3D T2w scan in the sagittal plane: TR/TE: 2,500 ms/251 ms, FA = 90° , acquisition matrix = $384 \text{ mm} \times 384 \text{ mm}$, voxel resolution $= 0.65 \text{ mm}^3$ isotropic, resulting in 282 slices without gap between adjacent slices; and (iii) T2w Fast Field Echo images using a blood-oxygen-level-dependent (BOLD) sensitive single-shot echo-planar imaging (EPI) sequence in the axial plane: TR/TE: 2,000 ms/30 ms, FA = 80° , acquisition matrix = $80 \text{ mm} \times 80 \text{ mm}$, voxel resolution = 3 mm^3 isotropic, resulting in 35 slices with 1 mm of gap between adjacent slices. To allow for optimal B1 shimming, a B1 calibration scan was applied before starting the EPI sequence. We acquired 250 EPI scans preceded by four dummy volumes to allow time for equilibrium in the spin excitation. Before starting the acquisition of the EPI sequence, participants were asked to remain still and keep their eyes closed without falling sleep. Pulse and respiratory rates were recorded using the scanner's built-in pulse oximeter placed on the left-hand index finger and a pneumatic respiratory belt strapped around the upper abdomen, respectively. Brain images were visually examined after each MRI sequence; they were repeated if artifacts were clearly identified. All participants underwent the same MRI protocol at the MRI facility of Pablo de Olavide University.

Structural MRI Preprocessing and Generation of Cortical Myelin Maps

T1w scans were preprocessed using Freesurfer v6.0.¹ The Freesurfer's pipeline included brain extraction, intensity normalization, automated tissue segmentation, generation of white matter (WM) and pial surfaces, correction of surface topology and inflation, co-registration, and projection of cortical surfaces to a sphere for establishing a surface-based coordinate system (Fischl and Dale, 2000). Pial surface misplacements and erroneous WM segmentation were manually corrected on a slice-by-slice basis by one experienced technician. The T2w image was

¹https://surfer.nmr.mgh.harvard.edu/

registered to the T1w image with *bbregister* employing a trilinear interpolation method. *bbregister* is a within-subject, cross-modal registration program that uses a boundary-based cost function constrained to 6 degrees of freedom (Greve and Fischl, 2009).

We have used the T1w/T2w ratio image to indirectly estimate the relative intracortical myelin content in individual cortical surfaces. Previous studies have shown that regional variations in the T1w/T2w ratio match the myelin content derived from histologically-defined cortical areas (Glasser and Van Essen, 2011) and they further correlate with cortical myelination patterns across the lifespan (Grydeland et al., 2019). This is partially due because myelin alters the signal intensity of T1w and T2w images in opposite directions and, consequently, the T1w/T2w ratio provides both enhanced tissue contrast and sensitivity to brain myelin content (Uddin et al., 2019). Furthermore, the T1w/T2w ratio cancels radiofrequency (RF) receive field (B1-) artifacts in the absence of head motion, and corrects reasonably well for RF transmit field (B1+) effects when the participants' head is precisely located at the isocenter of the magnetic field (Glasser et al., in press). In the present study, the latter requirements were strictly met to reduce transmit field biases and spurious results in cortical myelin maps.

The tissue fraction effect was corrected in individual T1w/T2w ratio images using PETsurfer (Greve et al., 2016), setting the point spread function estimate to zero. Next, three intermediate surfaces were generated within the cortical GM at fixed relative distances between the WM and pial surfaces (Polimeni et al., 2010). These surfaces were located at 90% (outer layer), 50% (medium layer) and 10% (inner layer) of the cortical thickness away from the WM surface. Finally, individual myelin maps obtained at different cortical depths were projected onto the average cortical surface, transformed to z-scores, and smoothed using non-linear spherical wavelet-based de-noising schemes (Bernal-Rusiel et al., 2008). All processing steps were visually checked for quality assurance. **Figure 1** illustrates the analysis pipeline for intracortical myelin mapping.

Functional MRI Preprocessing and Resting State-Functional Connectivity Analysis

Resting-state functional magnetic resonance imaging (rsfMRI) data were preprocessed using AFNI functions,² version AFNI_20.3.01. For each participant, high-frequency spikes were eliminated (*3dDespike*), time-locked cardiac and respiratory motion artifacts on cerebral BOLD signals were minimized using RETROICOR (Glover et al., 2000), time differences in slice-acquisition were corrected (*3dTshift*), EPI scans were aligned using rigid body motion correction and selecting the first volume as reference (*3dVolreg*), and aligned EPI scans were co-registered to their corresponding T1w volumes (*3dAllineate*; cost function: lpc + ZZ).

Dynamics were removed provided that more than 5% of voxels exhibited signal intensities that deviated from the median

absolute deviation of time series (*3dToutcount*); and/or when the Euclidean norm (*enorm*) threshold exceeded 0.3 mm in head motion. None of the participants showed more than 20% of artifactual dynamics after applying censoring. Simultaneous regression was further applied to minimize the impact of non-neuronal fluctuations on the rs-fMRI signal (*3dTproject*). Nuisance regressors included: (i) six head motion parameters (3 translational and 3 rotational) derived from the EPI scan alignment along with their first-order derivatives, (ii) time series of mean total WM/CSF signal intensity, and (iii) cardiac (measured by pulse oximeter) and respiratory fluctuations plus their derivatives to mitigate effects of extracerebral physiological artifacts on cerebral BOLD signals. No temporal band-pass filtering was applied.

Preprocessed rs-fMRI scans were projected onto the *fsaverage5* cortical surface space. Seeds for FC analyses were derived from cortical regions showing significant associations between plasma measurements (i.e., $A\beta_{1-42}$ and NfL levels) and intracortical myelin content. Surface-based rs-FC seed to whole cortex maps were obtained using the Fisher's z-transform of the corresponding Pearson's correlation coefficients.

Sample Size Estimation

To estimate the sample size, we performed power analysis with the G*Power software (v3.1.9.6).3 Only two studies have previously assessed the relationship between $A\beta_{1-42}$ and intracortical myelin content, one in the AD continuum using both CSF A β_{1-42} and amyloid PET (Luo et al., 2019) and other in cognitively normal older adults using amyloid PET (Yasuno et al., 2017). To our knowledge, research investigating associations between NfL levels and intracortical myelin content or addressing the potential moderating effect of plasma measurements (i.e., $A\beta_{1-42}$ or NfL) on the relationship between intracortical myelin content and cortical rs-FC patterns is lacking. Therefore, we computed an a priori (prospective) power analysis (fixed model, R^2 deviation from zero) to achieve statistical power of 80% given a 0.025 two-sided significance level and an overall Cohen's effect size (f²) ranging from 0.05 to 0.2. **Supplementary Figure 1** shows the results of this analysis for additive models evaluating the relationship between one of the plasma measurements (either $A\beta_{1-42}$ or NfL) and intracortical myelin content (3 predictors), and for interactive models assessing the moderating effect of one of the aforementioned plasma measurements on the relationship between intracortical myelin and rs-FC (5 predictors). This analysis revealed that 263 and 308 participants are required to detect an overall effect size of 0.05 for additive and interactive models, respectively. As the study sample consisted of 133 participants, interpretation of significances is conditional on the existence of overall effect sizes equal to or greater than 0.10 and 0.12 for main and interaction effects, respectively.

Statistical Analysis

We first applied the frequentist approach to determine whether age was related to plasma measurements and intracortical myelin

²https://afni.nimh.nih.gov/afni

³https://www.psychologie.hhu.de/arbeitsgruppen/allgemeine-psychologie-und-arbeitspsychologie/gpower.html





after adjusting for the effects of sex. Next, by applying the same approach, we performed vertex-wise multiple linear regression analyses to evaluate the relationship of plasma measurements (either $A\beta_{1-42}$ or NfL) with intracortical myelin content at different cortical depths (90, 50, and 10% away from the WM surface), including age and sex as covariates of no interest. To assess whether associations between plasma measurements and intracortical myelin content differed with cortical depth, we first computed the mean of the sum of all significant regions at the three cortical depths to extract the fitted values. These fitted values were entered into a mixed model with subjects as random effect to determine whether the strength of these

associations varied significantly as a function of cortical depth. The effect of the moderator factor (i.e., cortical depth) was evaluated through an ANOVA that compared the null model only including the intercept with the experimental model including cortical depth as a fixed effect. If differences reached significance, *post hoc* comparisons were performed with the R package emmeans and multiple comparisons were adjusted using the Bonferroni correction.

Using the frequentist approach, we further evaluated whether cortical regions showing significant associations with plasma measurements also exhibited altered patterns of rs-FC. For this, we applied a vertex-wise multiple regression model that included as regressor of interest the interaction between plasma measurements (either $A\beta_{1-42}$ or NfL) and the mean intracortical myelin of the region used as seed in the rs-FC analysis. These models were adjusted by age and sex.

Vertex-wise regression analyses for intracortical myelin and rs-FC were performed using the SurfStat package.⁴ Results were corrected for multiple comparisons using a hierarchical statistical model that first controls the family-wise error rate at the level of clusters by applying random field theory over smoothed statistical maps ($p_{vertex} < 0.001$, $p_{cluster} < 0.05$), and next controls the false discovery rate at the level of vertex within each cluster (p < 0.05) over unsmoothed statistical maps (Bernal-Rusiel et al., 2010). The anatomical location of significant results was based on vertices showing the maximum statistic within each significant cluster (Desikan et al., 2006).

After inferential evidence of a main or an interaction effect, we computed the standardized measure of effect size (i.e., Cohen's f^2) to evaluate local effect size within the context of a multivariate regression model (Cohen, 1988). To establish the precision of standardized effect sizes, we computed 95% confidence intervals (CI_{95%}) using the normal approximated interval with bootstrapped bias and standard error (N = 10,000 bootstrap samples) with the Matlab's *bootci* function.

Next, for each cortical vertex showing the maximum statistic within each significant cluster, we applied Bayesian linear regression analyses using JASP, version 0.12.2.5 The Bayesian approach allowed us to quantify the evidence for the alternative hypothesis and to overcome the problem of multiple comparisons resulting from performing different models for two hemispheres and three cortical depths. Bayesian linear regression analyses were based on non-informative priors such as the Jeffreys-Zellner-Siow prior with an r scale of 0.354 (Liang et al. (2008). We compared the strength of the Bayes factor for the model including all covariates of no interest (null model) with the model including the predictor of interest (experimental model) (BF₁₀). The classification scheme proposed by Lee and Wagenmakers (2013) was employed to interpret the BF₁₀. We only reported those results that reached significance according to the frequentist approach as long as the evidence in favor of the alternative hypothesis was at least moderate (BF₁₀ \geq 3).

Finally, we evaluated whether associations of intracortical myelin content with either plasma $A\beta_{1-42}/NfL$ levels or rs-FC accounted for the variability in global cognitive function. For this purpose, we first applied the Yeo-Johnson transformation to the cognitive index (i.e., the latent variable *g*) to reduce the detrimental effects of skewedness and heteroscedasticity in the different models (Yeo and Johnson, 2000). Next, we built four models to assess main and interaction effects. The first model only included the covariates of no interest; the second model the main effects; the third model the two-way interactions between the different predictors of interest; and the fourth model the three-way interaction. We first calculated F-tests between the different sequential models, and then compared the relative strength of the Bayes factor between the models.

⁴https://www.math.mcgill.ca/keith/surfstat/

RESULTS

Associations of Age With Intracortical Myelin and Plasma Levels of $A\beta_{1-42}$ and NfL

Age was positively related to plasma $A\beta_{1-42}$ (F_{1,130} = 9.8, p = 0.002) and NfL levels (F_{1,130} = 28.8, $p < 10^{-6}$). On the contrary, age was negatively associated with intracortical myelin content in postcentral regions of the left hemisphere (F_{1,130} = 21.2, $p < 10^{-5}$) and superior parietal lobe of the right hemisphere (F_{1,130} = 21.3, $p < 10^{-5}$). These results are illustrated in **Supplementary Figure 2**.

Relationship Between Intracortical Myelin and Plasma Levels of $A\beta_{1-42}$ and NfL

Figure 2 displays intracortical myelin maps projected onto the average cortical surface obtained at different depths (i.e., outer, medium and inner layers) before z-transformation. As expected, T1w/T2w ratio values increased with cortical depth (Nieuwenhuys, 2013; Nave and Werner, 2014). Our T1w/T2w ratio maps differ from those reported in Glasser and Van Essen (2011), especially in some regions of the occipital cortex, whose intracortical myelin content was associated with plasma levels of $A\beta_{1-42}$ (see Figure 3). These differences may be due to the distinct approaches used to obtain the intracortical myelin maps in the two studies. Thus, we used absolute values comprising the entire range of T1w/T2w ratio values instead of a percentile scaling between 3 and 96%, as in Glasser and Van Essen (2011). Neither we adjusted the color palette to identify the transitions between adjacent areas, since we were not interested in delineating spatial gradients or myeloarchitectural features of cortical regions (Glasser and Van Essen, 2011).

Plasma levels of $A\beta_{1-42}$ were positively associated with T1w/T2w ratio values in temporo-parietal-occipital regions of both hemispheres at the three cortical depths, as illustrated in **Figure 3** and reported in **Table 2**. The evidence in favor of the alternative hypothesis was extreme (BF₁₀ > 100) for myelin content of inner and outer layers, and moderate in the medium layer (BF₁₀ was reported in **Table 2**).

Regarding the association between $A\beta_{1-42}$ and intracortical myelin, mixed model ANOVAs revealed a significant main effect of cortical depth for the right hemisphere ($\chi^2 = 346.6$, $p < 10^{-15}$) that was not extended to the left hemisphere ($\chi^2 = 5.7$, p = 0.06). *Post hoc* analyses showed that the strength of positive associations in the right hemisphere decreased with cortical depth, being stronger for the outer than for the other two layers (p < 0.0001), and stronger for the medium than for the innermost layer (p < 0.0001). **Figures 4A,B** illustrate these results.

The regression analysis also revealed a negative association between plasma NfL levels and T1w/T2w ratio values of the right insular cortex (**Figure 5** and **Table 2**). This result was restricted to the inner layer, where the Bayesian linear regression analysis showed a BF₁₀ of 3.4, indicating that the alternative hypothesis is

⁵https://jasp-stats.org/



FIGURE 2 | Intracortical myelin maps projected onto the average cortical surface obtained at different depths (outer, medium, and inner layer) before z-transformation. Note that, in general, T1w/T2w ratio values increase with cortical depth. Left (L) and right (R).





TABLE 2 | Significant associations of plasma Ab₁₋₄₂ and NfL with T1w/T2w ratio intensity values.

Vertex location with maximum statistic	MNI	R ²	F _{5,127}	f ² _{local}	CI _{95%}	BF ₁₀
Positive associations with $A\beta_{1-42}$ (inner layer,)					
L fusiform ($p_{cluster} = 10^{-6}$)	-40 -73 -14	0.18	27.3	0.50	0.47-0.54	220 ^E
L lateral occipital ($p_{cluster} = 10^{-5}$)	-30 -90 -15	0.19	34.7	0.53	0.50-0.56	1234 ^E
L inferior temporal ($p_{cluster} = 0.02$)	-42 -51 -11	0.12	16.4	0.28	0.25-0.32	87 ^{VS}
R lateral occipital ($p_{cluster} = 10^{-6}$)	19 –85 –8	0.11	18.3	0.35	0.32-0.38	2436 ^E
R precuneus (p _{cluster} = 0.0004)	13 –54 16	0.13	19.0	0.38	0.35-0.42	6145 ^E
R lingual ($p_{cluster} = 0.003$)	7 –60 2	0.13	19.4	0.35	0.32-0.38	2047 ^E
Positive associations with $A\beta_{1-42}$ (medium lag	ver)					
L fusiform ($p_{cluster} = 0.0003$)	-30 -72 -16	0.14	13.2	0.31	0.28-0.34	3.9 ^M
R lingual ($p_{cluster} = 10^{-5}$)	4 –62 7	0.14	16.7	0.36	0.32-0.39	4.2 ^M
Positive associations with $A\beta_{1-42}$ (outer layer)					
L fusiform ($p_{cluster} = 10^{-5}$)	-40 -71 -18	0.19	13.2	0.34	0.31-0.37	1817 ^E
L superior parietal ($p_{cluster} = 0.0002$)	-23 -84 24	0.14	18.8	0.37	0.34-0.40	2643 ^E
R pericalcarine ($p_{cluster} = 10^{-5}$)	5 –78 12	0.12	5.5	0.34	0.31-0.37	7472 ^E
R lateral occipital ($p_{cluster} = 10^{-5}$)	20 - 89 - 8	0.15	21.3	0.36	0.33-0.39	3547 ^E
Negative associations with NfL (inner layer)						
R insula (p _{cluster} = 0.04)	41 0 4	0.16	25.4	0.46	0.42-0.49	3.4 ^M

MNI coordinates are in MNI152 space. ℓ^2 : measure of local effect size. $Cl_{95\%}$: 95% confidence interval. BF_{10} : Bayes factor yielded by the Bayesian linear regression analysis. The superscript of the BF_{10} indicates the qualitative interpretation of the evidence for the alternative hypothesis: ^M moderate; ^{VS} very strong; ^E extreme. L: left; R: right.



FIGURE 4 Boxplots showing individual fitted mean intracortical myelin values for regions showing a significant association with plasma measurements at different cortical depths. Effect of cortical depth for the $A\beta_{1-42}$ associations with intracortical myelin in the left **(A)** and right hemisphere **(B)**. Note that positive associations between plasma $A\beta_{1-42}$ levels and intracortical myelin content (T1w/T2w ratio values) increased from inner to outer cortical layers only in the right hemisphere. **(C)** Effect of cortical depth for the NfL associations with intracortical myelin of the right insula. Note that the association between plasma NfL levels and intracortical myelin content (T1w/T2w ratio values) was more negative in the inner than in the medium and outer layers.

about three times more likely than the null, which is classified as moderate. The mixed model ANOVA addressing the effect of cortical depth on the strength of this association revealed a significant main effect in the right insular cortex ($\chi^2 = 354.9$, $p < 10^{-15}$). The association of NfL with intracortical myelin was more negative for the innermost layer than for the other two layers (p < 0.0001). **Figure 4C** illustrates these results.

Effect of the Relationship Between Plasma Measurements and Myelin Content on Rs-FC

We next performed rs-FC analysis using as seeds those regions derived from significant associations between

plasma measurements (A β_{1-42} and NfL) and myelin content (regions displayed in **Figures 3**, **5**). The interaction between plasma A β_{1-42} and intracortical myelin did not predict rs-FC for none of the seeds evaluated. Conversely, the multiple regression analysis performed on the rs-FC map with the right insula as seed revealed a significant two-way interaction in the left medial orbitofrontal cortex ($R_{max}^2 = 0.13$, $t_{max} = 4.36$, $p_{cluster} = 0.006$, rho = -2.22, $CI_{95\%}$ [-2.81 to -1.63]). **Figure 6A** shows the spatial location of this result. *Post hoc* analyses shown in **Figure 6B** revealed a negative relationship between FC and myelin in individuals with higher plasma levels of NfL (i.e., the group with + 1SD) and a positive relationship in those with lower NfL levels (i.e., the group with -1SD). The Bayesian analysis yielded a BF₁₀



content (T1w/T2w ratio values) on the cortical surface obtained at 10% of the cortical thickness away from the WM surface (inner layer). (A) Projected size effects on the inflated cortical surface indicate negative associations of plasma NfL levels with intracortical myelin content, adjusted by age and sex. The color bar indicates the range of overall size effects (Cohen's f²). Right (R). (B) The scatter plot shows the partial correlation between plasma NfL levels and the mean myelin content of the significant cluster for the inner cortical layer located in the right insula, adjusted by age and sex.

of 304, and therefore the evidence in favor of the alternative hypothesis is considered as extreme.

Associations of Plasma Measurements, Intracortical Myelin and Resting State-Functional Connectivity With Cognition

The ANOVAs performed to compare the sequential models revealed no significant main or interaction effects on cognition. BF_{10} ranged between 0.33 and 1. Accordingly, no evidence in favor of the null hypothesis was observed in either case.

DISCUSSION

We showed that plasma levels of $A\beta 1-42$ and NfL were associated with intracortical myelin content, suggesting that they are able to track variations in myelin content in the aging neocortex. Interestingly, associations with plasma NfL were most evident in inner cortical layers, where fibers are more densely myelinated (Nave and Werner, 2014) and oligodendrogenesis is



FIGURE 6 | Moderating effect of plasma NfL levels on the relationship between the mean myelin content and the rs-FC pattern of the right insula. (A) Projected size effects on the inflated cortical surface show the moderating role of plasma NfL levels in the association between the myelin content and rs-FC pattern of the right insula. The color bar indicates the overall range of size effects (Cohen's f²). Left (L). (B) Scatter plot for the association of the mean myelin content of the right insula with rs-FC between the right insula and left medial orbitofrontal cortex (mOFC) at 1 SD below and above the mean of plasma NfL levels, adjusted by age and sex.

most impaired (Orthmann-Murphy et al., 2020; Xu et al., 2021). Furthermore, we found that higher concentrations of plasma NfL were associated with aberrant FC patterns of the insular cortex, a central brain hub highly vulnerable to neurodegeneration involved in affective and cognitive processes (Cauda et al., 2011). Considering that myelin undergoes significant alterations during aging and that aging is the major risk factor for AD, these results may be helpful in detecting myelin dysfunctions in aging and hypothetically serve to identify vulnerability to develop AD.

Plasma A_{β1-42} Levels Are Positively Associated With Intracortical Myelin Content

Previous studies have shown that low plasma $A\beta_{1-42}$ concentrations appear in MCI/AD patients carrying the Apolipoprotein E ϵ 4 allele (APOE4) (Kleinschmidt et al., 2016) and in cognitively normal individuals showing abnormal CSF-amyloid status and positive amyloid PET scans (Verberk et al., 2018). Moreover, plasma $A\beta_{1-42}$ levels have been

negatively associated with WM hyperintensities and positively with global cognition, memory performance, hippocampal volume and cortical thickness of the temporal lobe (Llado-Saz et al., 2015; Poljak et al., 2016; Hilal et al., 2018), as well as with a steeper rate of subsequent cognitive decline (Rembach et al., 2014; Poljak et al., 2016, Verberk et al., 2020) and increased risk of AD (Chouraki et al., 2015; Hilal et al., 2018; de Wolf et al., 2020).

Our results revealed a positive relationship between plasma $A\beta_{1-42}$ levels and intracortical myelin in cognitively normal older adults, linking plasma amyloid markers to cortical myelin integrity in normal aging. The association between cerebral amyloid aggregation and myelin deficits has been mainly supported by research in mouse models of AD (Desai et al., 2010; Mitew et al., 2010; Schmued et al., 2013; Chen et al., 2021). These studies showed that myelin pathology extends across regions with AB deposits (Mitew et al., 2010; Schmued et al., 2013), suggesting that the absence of oligodendrocytes in the plaque core may contribute to the destabilization of neuronal networks surrounding A β deposits (Palop and Mucke, 2010). This interpretation has been reinforced by research showing in vivo associations between myelin alterations and amyloid pathology in preclinical AD (Dean et al., 2017; Luo et al., 2019). In particular, lower CSF A β_{1-42} levels were found to be associated with lower myelin water fraction (MWF) in the WM of late myelinating brain regions (Dean et al., 2017), which are affected by AD lesions from the earliest stage (Braak and Del Tredici, 2015). Further evidence indicates that preclinical AD patients exhibit the lowest T1w/T2w ratio in the inferior parietal lobe, which continues to decrease with disease progression (Luo et al., 2019). Our results complement these findings, providing evidence that positive associations of intracortical myelin with plasmatic $A\beta_{1-42}$ are restricted to posterior cortical regions in normal aging. Together, these results suggest that myelin breakdown in aging and preclinical AD occurs in the reverse direction in which myelination develops, from cortical association regions to subcortical WM projection tracts, and from posterior to anterior brain regions (Bartzokis, 2004).

An alternative interpretation is that the load of WM lesions, instead of A β deposition in plaques, could drive the relationship between plasma $A\beta_{1-42}$ and intracortical myelin content. In line with this hypothesis, it has been reported that associations between CSF $A\beta_{1-42}$ levels and cortical atrophy are noticeable in dementia patients but absent in cognitively normal older adults, whereas correlations between CSF $A\beta_{1-42}$ and WM lesions are present in normal elderly individuals but lacked in dementia patients (Skoog et al., 2018). Further, if associations between plasma $A\beta_{1-42}$ and intracortical myelin content were a reflection of the cerebral amyloid burden, this relationship should be stronger in deeper cortical layers, where the myelin content is higher (Hughes et al., 2018). This prediction is supported by evidence indicating that A β staining in 5×FAD transgenic mice is more intense in deeper than in superficial cortical layers. As a result, T1 and T2 values were more drastically reduced in deeper cortical regions in transgenic mice compared to wild-type mice (Spencer et al., 2013). Moreover, analysis of the laminar distribution of transmitter receptors

in the human neocortex has shown that deeper layers of temporo-occipital regions concentrate a high density of kainate receptors (Palomero-Gallagher and Zilles, 2019). Given that oligodendrocytes contribute to the control of extracellular glutamate levels (Matute et al., 2001) and oligodendrogenesis is mostly impaired in the deeper cortical layers (Orthmann-Murphy et al., 2020; Xu et al., 2021), alterations in glutamate homeostasis may result in overactivation of kainate receptors and subsequent excitotoxic oligodendroglial death, especially in the innermost layers. However, our results showed that associations between plasma $A\beta_{1-42}$ and intracortical were stronger in the outer cortical layer (at least in the right hemisphere), suggesting that plasma $A\beta_{1-42}$ -myelin associations are not likely driven by cerebral amyloid burden.

It is also worth noting that T1 relaxation rate correlates with iron concentration in the brain, showing much higher iron relaxivity values in the cortex than in other brain structures (Ogg and Steen, 1998). Accordingly, high-field MRI data from postmortem brain samples has revealed a layered organization of iron content in the cortical mantle suggestive of each region's myeloarchitecture (Fukunaga et al., 2010). Therefore, we cannot rule out that associations between plasma measurements and intracortical myelin content are partially due to iron accumulation. Future experiments should use more sophisticated *in vivo* approaches to separate iron and myelin content in the human cortex (Shin et al., 2021) in order to reliably determine their specific effects on aging and aging-related neurodegenerative conditions.

Plasma NfL Levels Are Negatively Associated With Intracortical Myelin Content and Abnormal Resting State-Functional Connectivity

Neurofilament light chain is one of the scaffolding proteins of the axonal cytoskeleton with important roles in axonal and dendritic branching (Yuan et al., 2017). NfL is particularly abundant in myelinated axons and is released to the periphery upon damage of the central nervous system (Gafson et al., 2020). Although plasma NfL is not a specific AD biomarker, it has been positively associated with higher risk of cognitive decline in cognitively normal older adults, MCI and AD patients (Mattsson et al., 2017; Aschenbrenner et al., 2020; Lee et al., 2022) and higher CSF NfL concentration in the early presymptomatic stages of familial AD (Preische et al., 2019) and in all stages of sporadic AD (Mattsson et al., 2017). At baseline, it has been positively related to neocortical AB and tau in cognitively unimpaired and impaired older adults, respectively (Chatterjee et al., 2018; Benedet et al., 2020) and negatively to hippocampal atrophy in all stages of AD (Mattsson et al., 2017; Hu et al., 2019) and to the mean cortical thickness of the whole-brain and specific brain regions in MCI and cognitively normal older adults (Lee et al., 2022). Over time, decreasing plasma NfL levels have also been related to progressive enlargement of the lateral ventricles and decreasing brain metabolism, hippocampal volume and cortical thickness especially in MCI and AD patients (Mattsson et al., 2017; Benedet et al., 2020). Consequently, plasma NfL is considered a promising

and cost-effective biomarker of axonal injury in AD and a variety of neurological conditions (Khalil et al., 2018).

High blood NfL levels may further reflect myelin breakdown, presumably due to dysfunctional oligodendrocytes, which provide trophic support to axons (Mot et al., 2018). Multiple lines of evidence indirectly support this hypothesis in humans. For instance, recent experiments have revealed associations between high serum NfL levels and MRI markers of myelin damage in multiple sclerosis (Yik et al., 2022), pediatric acquired demyelinating syndrome (Simone et al., 2021), and X-linked adrenoleukodystrophy (Weinhofer et al., 2021). Our results add further support to this hypothesis, showing a negative relationship between plasma NfL levels and intracortical myelin content of the right insular cortex, a lightly myelinated cortical region that has shown increased vulnerability in aging (Hu et al., 2014), preclinical (Cantero et al., 2020) and prodromal AD (Cantero et al., 2017).

We also showed that plasma NfL levels modulated patterns of FC between the insula and medial orbitofrontal cortex, two lightly myelinated cortical areas that appeared to be vulnerable to detrimental effects of aging (Bartzokis, 2011; Vidal-Piñeiro et al., 2016). Recent evidence has linked rs-FC patterns of insular cortex and orbitofrontal cortex to apathy (Jang et al., 2021), a multi-domain syndrome associated with poor outcomes and incident dementia in normal older adults (van Dalen et al., 2018), suggesting that impaired FC of these two regions may have unfavorable implications for aging. Tract tracing studies in primates have revealed that the anterior insula has prominent connections to the orbitofrontal cortex (Flynn et al., 1999), forming a functional unit that serves in the integration of complex autonomic, cognitive and emotional processes (Morel et al., 2013), all of them manifestly affected by aging (Salthouse, 2012). We speculate that as long as plasma NfL levels are elevated, reduced myelin content of the insular cortex may affect synchronized timing of neuronal impulses leading to abnormal functional connections with the medial orbitofrontal cortex.

Study Limitations

Results of the present study were cross-sectionally obtained, which has strengths and weaknesses that should be acknowledged. While cross-sectional studies are valuable for establishing preliminary evidence in planning advanced studies, they are not helpful for drawing predictive conclusions. As long as the study sample is representative of the overall population, cross-sectional studies are adequate for measuring the prevalence of health outcomes, understand determinants of health, and describe features of a population (Wang and Cheng, 2020). Therefore, our results cannot establish a true cause and effect relationship between T1w/T2w ratio values and plasma levels of $A\beta_{1-42}$ and NfL, nor can they determine a temporal relationship between outcomes and risk factors. However, they could be used to generate predictive hypothesis for longitudinal studies in which T1w/T2w ratio values are used as a predictive or dependent variable.

Linking our results to AD requires caution. Brain-specific proteins are present in much lower concentrations in blood than in CSF because the blood-brain barrier prevents free passage of molecules between the CNS and blood compartments (Zetterberg and Burnham, 2019). More importantly, A β species are expressed in non-brain tissues and bind to a variety of blood proteins (Marcello et al., 2009), which may not reflect brain A β turnover/metabolism and consequently reduce the potential for monitoring A β pathology in the blood. Moreover, the plasma A $\beta_{1-42}/A\beta_{1-40}$ ratio has shown better concordance with cerebral amyloid load than plasma A β_{1-42} alone (Janelidze et al., 2016; Schindler et al., 2019), suggesting that the A $\beta_{1-42}/A\beta_{1-40}$ ratio has an added value as a pre-screening biomarker in AD.

CONCLUSION

The present study showed that lower plasma $A\beta_{1-42}$ and higher plasma NfL levels were associated with lower intracortical myelin content in temporo-parietal-occipital regions and the insula, respectively. Notably, the latter association was most evident in inner cortical layers, where axons are more heavily myelinated (Nave and Werner, 2014) and oligodendrogenesis is most impaired (Orthmann-Murphy et al., 2020; Xu et al., 2021). Plasma NfL levels further moderated the relationship between intracortical myelin content and rs-FC, likely revealing a complex inter-relationship between axonal damage, myelin breakdown and FC in aging. As myelin undergoes significant alterations in aging, the most important risk factor for AD, these results may be helpful in detecting aging-related myelin dysfunctions and potentially serve to identify vulnerability to develop AD.

DATA AVAILABILITY STATEMENT

The raw data supporting the conclusions of this article will be made available by the authors on reasonable request.

ETHICS STATEMENT

The studies involving human participants were reviewed and approved by Ethical Committee for Clinical Research of the Junta de Andalucía. The patients/participants provided their written informed consent to participate in this study.

AUTHOR CONTRIBUTIONS

JLC conceived the study and wrote the manuscript. MF-A, MA, and JLC contributed to data acquisition, data analysis, and preparation of figures and tables. FZ, CM, and EC-Z performed the plasma $A\beta_{1-42}$ and NfL determinations. All authors read and approved the final version of the manuscript.

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REFERENCES

- Aschenbrenner, A. J., Gordon, B. A., Fagan, A. M., Schindler, S. E., Balota, D. A., Morris, J. C., et al. (2020). Neurofilament light predicts decline in attention but not episodic memory in preclinical Alzheimer's disease. J. Alzheimers Dis. 74, 1119–1129. doi: 10.3233/JAD-200018
- Bartzokis, G. (2004). Age-related myelin breakdown: a developmental model of cognitive decline and Alzheimer's disease. *Neurobiol. Aging* 25, 5–18. doi: 10. 1016/j.neurobiolaging.2003.03.001
- Bartzokis, G. (2011). Alzheimer's disease as homeostatic responses to agerelated myelin breakdown. *Neurobiol. Aging* 32, 1341–1371. doi: 10.1016/j. neurobiolaging.2009.08.007
- Benedet, A. L., Leuzy, A., Pascoal, T. A., Ashton, N. J., Mathotaarachchi, S., Savard, M., et al. (2020). Stage-specific links between plasma neurofilament light and imaging biomarkers of Alzheimer's disease. *Brain* 143, 3793–3804. doi: 10.1093/brain/awaa342
- Bernal-Rusiel, J. L., Atienza, M., and Cantero, J. L. (2008). Detection of focal changes in human cortical thickness: spherical wavelets versus Gaussian smoothing. *Neuroimage* 41, 1278–1292. doi: 10.1016/j.neuroimage.2008.03.022
- Bernal-Rusiel, J. L., Atienza, M., and Cantero, J. L. (2010). Determining the optimal level of smoothing in cortical thickness analysis: a hierarchical approach based on sequential statistical thresholding. *Neuroimage* 52, 158–171. doi: 10.1016/j. neuroimage.2010.03.074
- Beul, S. F., Barbas, H., and Hilgetag, C. C. (2017). A predictive structural model of the primate connectome. *Sci. Rep.* 7:43176. doi: 10.1038/srep43176
- Böhm, P., Peña-Casanova, J., Aguilar, M., Hernández, G., Sol, J. M., and Blesa, R. (1998). Clinical validity and utility of the interview for deterioration of daily living in dementia for Spanish-speaking communities NORMACODEM Group. *Int. Psychogeriatr.* 10, 261–270. doi: 10.1017/s1041610298005377
- Braak, H., and Del Tredici, K. (2015). The preclinical phase of the pathological process underlying sporadic Alzheimer's disease. *Brain* 138, 2814–2833. doi: 10.1093/brain/awv236
- Cantero, J. L., Atienza, M., Lage, C., Zaborszky, L., Vilaplana, E., Lopez-Garcia, S., et al. (2020). Atrophy of basal forebrain initiates with tau pathology in individuals at risk for Alzheimer's disease. *Cereb. Cortex* 30, 2083–2098. doi: 10.1093/cercor/bhz224
- Cantero, J. L., Zaborszky, L., and Atienza, M. (2017). Volume loss of the nucleus basalis of meynert is associated with atrophy of innervated regions in mild cognitive impairment. *Cereb. Cortex* 27, 3881–3889. doi: 10.1093/cercor/ bhw195
- Cauda, F., D'Agata, F., Sacco, K., Duca, S., Geminiani, G., and Vercelli, A. (2011). Functional connectivity of the insula in the resting brain. *Neuroimage* 55, 8–23. doi: 10.1016/j.neuroimage.2010.11.049
- Chatterjee, P., Goozee, K., Sohrabi, H. R., Shen, K., Shah, T., Asih, P. R., et al. (2018). Association of plasma neurofilament light chain with neocortical amyloid-β load and cognitive performance in cognitively normal elderly participants. J. Alzheimers Dis. 63, 479–487. doi: 10.3233/JAD-180025
- Chen, J. F., Liu, K., Hu, B., Li, R. R., Xin, W., Chen, H., et al. (2021). Enhancing myelin renewal reverses cognitive dysfunction in a murine model of Alzheimer's disease. *Neuron* 109, 2292–2307. doi: 10.1016/j.neuron.2021.05.012
- Chouraki, V., Beiser, A., Younkin, L., Preis, S. R., Weinstein, G., Hansson, O., et al. (2015). Plasma amyloid-β and risk of Alzheimer's disease in the Framingham Heart Study. *Alzheimers Dement*. 11, 249–257. doi: 10.1016/j.jalz.2014.07.001
- Cohen, J. E. (1988). *Statistical Power Analysis for the Behavioral Sciences*. Hillsdale, NJ: Lawrence Erlbaum Associates, Inc.
- Couttas, T. A., Kain, N., Suchowerska, A. K., Quek, L. E., Turner, N., Fath, T., et al. (2016). Loss of ceramide synthase 2 activity, necessary for myelin biosynthesis, precedes tau pathology in the cortical pathogenesis of Alzheimer's disease. *Neurobiol. Aging* 43, 89–100. doi: 10.1016/j.neurobiolaging.2016.03.027

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fnagi. 2022.896848/full#supplementary-material

- de Wolf, F., Ghanbari, M., Licher, S., McRae-McKee, K., Gras, L., Weverling, G. J., et al. (2020). Plasma tau, neurofilament light chain and amyloid-β levels and risk of dementia; a population-based cohort study. *Brain* 143, 1220–1232. doi: 10.1093/brain/awaa054
- Dean, D. C. III, Hurley, S. A., Kecskemeti, S. R., O'Grady, J. P., Canda, C., Davenport-Sis, N. J., et al. (2017). Association of amyloid pathology with myelin alteration in preclinical Alzheimer disease. *JAMA Neurol.* 74, 41–49. doi: 10.1001/jamaneurol.2016.3232
- Desai, M. K., Mastrangelo, M. A., Ryan, D. A., Sudol, K. L., Narrow, W. C., and Bowers, W. J. (2010). Early oligodendrocyte/myelin pathology in Alzheimer's disease mice constitutes a novel therapeutic target. *Am. J. Pathol.* 177, 1422– 1435. doi: 10.2353/ajpath.2010.100087
- Desikan, R. S., Ségonne, F., Fischl, B., Quinn, B. T., Dickerson, B. C., Blacker, D., et al. (2006). An automated labeling system for subdividing the human cerebral cortex on MRI scans into gyral based regions of interest. *Neuroimage* 31, 968–980. doi: 10.1016/j.neuroimage.2006.01.021
- Felts, P. A., Baker, T. A., and Smith, K. J. (1997). Conduction in segmentally demyelinated mammalian central axons. J. Neurosci. 17, 7267–7277. doi: 10. 1523/JNEUROSCI.17-19-07267.1997
- Fields, R. D. (2015). A new mechanism of nervous system plasticity: activitydependent myelination. Nat. Rev. Neurosci. 16, 756–767. doi: 10.1038/nrn4023
- Fischl, B., and Dale, A. M. (2000). Measuring the thickness of the human cerebral cortex from magnetic resonance images. *Proc. Natl. Acad. Sci. U. S. A.* 97, 11050–11055. doi: 10.1073/pnas.200033797
- Flynn, F. G., Benson, F., and Ardila, A. (1999). Anatomy of the insula Functional and clinical correlates. *Aphasiology* 13, 55–78. doi: 10.1080/026870399402325
- Fukunaga, M., Li, T. Q., van Gelderen, P., de Zwart, J. A., Shmueli, K., Yao, B., et al. (2010). Layer-specific variation of iron content in cerebral cortex as a source of MRI contrast. *Proc. Natl. Acad. Sci. U. S. A.* 107, 3834–3839. doi: 10.1073/pnas.0911177107
- Gafson, A. R., Barthélemy, N. R., Bomont, P., Carare, R. O., Durham, H. D., Julien, J. P., et al. (2020). Neurofilaments: neurobiological foundations for biomarker applications. *Brain* 143, 1975–1998. doi: 10.1093/brain/awaa098
- Glasser, M. F., Coalson, T. S., Harms, M. P., Xu, J., Baum, G. L., Autio, J. A., et al. (in press). Empirical transmit field bias correction of T1w/T2w myelin maps. *bioRxiv* [Preprint]. doi: 10.1101/2021.08.08.455570
- Glasser, M. F., and Van Essen, D. C. (2011). Mapping human cortical areas in vivo based on myelin content as revealed by T1- and T2-weighted MRI. J. Neurosci. 31, 11597–11616. doi: 10.1523/JNEUROSCI.2180-11. 2011
- Glover, G. H., Li, T. Q., and Ress, D. (2000). Image-based method for retrospective correction of physiological motion effects in fMRI: RETROICOR. *Magn. Reson. Med.* 44, 162–167. doi: 10.1002/1522-2594(200007)44:1<162::aid-mrm23<3.0. co;2-e
- Gramunt, N., Sánchez-Benavides, G., Buschke, H., Diéguez-Vide, F., Peña-Casanova, J., Masramon, X., et al. (2016). The memory binding test: development of two alternate forms into Spanish and Catalan. J. Alzheimers Dis. 52, 283–293. doi: 10.3233/JAD-151175
- Greve, D. N., and Fischl, B. (2009). Accurate and robust brain image alignment using boundary-based registration. *Neuroimage* 48, 63–72. doi: 10.1016/j. neuroimage.2009.06.060
- Greve, D. N., Salat, D. H., Bowen, S. L., Izquierdo-Garcia, D., Schultz, A. P., Catana, C., et al. (2016). Different partial volume correction methods lead to different conclusions: an 18 F-FDG-PET study of aging. *Neuroimage* 132, 334–343. doi: 10.1016/j.neuroimage.2016.02.042
- Grydeland, H., Vértes, P. E., Váša, F., Romero-Garcia, R., Whitaker, K., Alexander-Bloch, A. F., et al. (2019). Waves of maturation and senescence in microstructural MRI markers of human cortical myelination over the lifespan. *Cereb. Cortex* 29, 1369–1381. doi: 10.1093/cercor/bhy330

- Hilal, S., Woltersm, F. J., Verbeek, M. M., Vanderstichele, H., Ikram, M. K., Stoops, E., et al. (2018). Plasma amyloid-β levels, cerebral atrophy and risk of dementia: a population-based study. *Alzheimers Res. Ther.* 10:63. doi: 10.1186/s13195-018-0395-6
- Hu, H., Chen, K. L., Ou, Y. N., Cao, X. P., Chen, S. D., Cui, M., et al. (2019). Neurofilament light chain plasma concentration predicts neurodegeneration and clinical progression in nondemented elderly adults. *Aging* 11, 6904–6914. doi: 10.18632/aging.102220
- Hu, S., Chao, H. H., Zhang, S., Ide, J. S., and Li, C. S. (2014). Changes in cerebral morphometry and amplitude of low-frequency fluctuations of BOLD signals during healthy aging: correlation with inhibitory control. *Brain Struct. Funct.* 219, 983–994. doi: 10.1007/s00429-013-0548-0
- Hughes, E. G., Orthmann-Murphy, J. L., Langseth, A. J., and Bergles, D. E. (2018). Myelin remodeling through experience-dependent oligodendrogenesis in the adult somatosensory cortex. *Nat. Neurosci.* 21, 696–706. doi: 10.1038/s41593-018-0121-5
- Huntenburg, J. M., Bazin, P. L., Goulas, A., Tardif, C. L., Villringer, A., and Margulies, D. S. (2017). A systematic relationship between functional connectivity and intracortical myelin in the human cerebral cortex. *Cereb. Cortex* 27, 981–997. doi: 10.1093/cercor/bhx030
- Irvine, K. A., and Blakemore, W. F. (2006). Age increases axon loss associated with primary demyelination in cuprizone-induced demyelination in C57BL/6 mice. J. Neuroimmunol. 175, 69–76. doi: 10.1016/j.jneuroim.2006.03.002
- Janelidze, S., Zetterberg, H., Mattsson, N., Palmqvist, S., Vanderstichele, H., Lindberg, O., et al. (2016). CSF Aβ42/Aβ40 and Aβ42/Aβ38 ratios: better diagnostic markers of Alzheimer disease. *Ann. Clin. Transl. Neurol.* 3, 154–165. doi: 10.1002/acn3.274
- Jang, J. Y., Han, S. D., Yew, B., Blanken, A. E., Dutt, S., Li, Y., et al. (2021). Restingstate functional connectivity signatures of apathy in community-living older adults. *Front. Aging Neurosci.* 13:691710. doi: 10.3389/fnagi.2021.691710
- Khalil, M., Teunissen, C. E., Otto, M., Piehl, F., Sormani, M. P., Gattringer, T., et al. (2018). Neurofilaments as biomarkers in neurological disorders. *Nat. Rev. Neurol.* 14, 577–589. doi: 10.1038/s41582-018-0058-z
- Kleinschmidt, M., Schoenfeld, R., Göttlich, C., Bittner, D., Metzner, J. E., Leplow, B., et al. (2016). Characterizing aging, mild cognitive impairment, and dementia with blood-based biomarkers and neuropsychology. J. Alzheimers Dis. 50, 111–126. doi: 10.3233/JAD-143189
- Lee, E. H., Kwon, H. S., Koh, S. H., Choi, S. H., Jin, J. H., Jeong, J. H., et al. (2022). Serum neurofilament light chain level as a predictor of cognitive stage transition. *Alzheimers Res. Ther.* 14:6. doi: 10.1186/s13195-021-00953-x
- Lee, M. D., and Wagenmakers, E. J. (2013). *Bayesian Cognitive Modeling: A Practical Course*. Cambridge: Cambridge University Press.
- Liang, F., Paulo, R., Molina, G., Clyde, M. A., and Berger, J. O. (2008). Mixtures of g priors for Bayesian variable selection. J. Am. Stat. Assoc. 103, 410–423. doi: 10.1198/016214507000001337
- Llado-Saz, S., Atienza, M., and Cantero, J. L. (2015). Increased levels of plasma amyloid-beta are related to cortical thinning and cognitive decline in cognitively normal elderly subjects. *Neurobiol. Aging* 36, 2791–2797. doi: 10.1016/j. neurobiolaging.2015.06.023
- Luo, X., Li, K., Zeng, Q., Huang, P., Jiaerken, Y., Wang, S., et al. (2019). Application of T1-/T2-weighted ratio mapping to elucidate intracortical demyelination process in the Alzheimer's disease continuum. *Front. Neurosci.* 13:904. doi: 10.3389/fnins.2019.00904
- Marcello, A., Wirthsm, O., Schneider-Axmann, T., Degerman-Gunnarsson, M., Lannfelt, L., and Bayer, T. A. (2009). Circulating immune complexes of Abeta and IgM in plasma of patients with Alzheimer's disease. *J. Neural Transm.* 116, 913–920. doi: 10.1007/s00702-009-0224-y
- Mathys, H., Davila-Velderrain, J., Peng, Z., Gao, F., Mohammadi, S., Young, J. Z., et al. (2019). Single-cell transcriptomic analysis of Alzheimer's disease. *Nature* 570, 332–337. doi: 10.1038/s41586-019-1195-2
- Mattsson, N., Andreasson, U., Zetterberg, H., Blennow, K., and Alzheimer's Disease Neuroimaging Initiative. (2017). Association of plasma neurofilament light with neurodegeneration in patients with Alzheimer Disease. JAMA Neurol. 74, 557–566. doi: 10.1001/jamaneurol.2016.6117
- Matute, C., Alberdi, E., Domercq, M., Pérez-Cerdá, F., Pérez-Samartín, A., and Sánchez-Gómez, M. V. (2001). The link between excitotoxic oligodendroglial death and demyelinating diseases. *Trends Neurosci.* 24, 224–230. doi: 10.1016/ s0166-2236(00)01746-x

- Mitew, S., Kirkcaldie, M. T., Halliday, G. M., Shepherd, C. E., Vickers, J. C., and Dickson, T. C. (2010). Focal demyelination in Alzheimer's disease and transgenic mouse models. *Acta Neuropathol.* 119, 567–577. doi: 10.1007/ s00401-010-0657-2
- Monje, M. (2018). Myelin plasticity and nervous system function. Annu. Rev. Neurosci. 41, 61–76. doi: 10.1146/annurev-neuro-080317-061853
- Morel, A., Gallay, M. N., Baechler, A., Wyss, M., and Gallay, D. S. (2013). The human insula: architectonic organization and postmortem MRI registration. *Neuroscience* 236, 117–135. doi: 10.1016/j.neuroscience.2012.12.076
- Mot, A. I., Depp, C., and Nave, K. A. (2018). An emerging role of dysfunctional axon-oligodendrocyte coupling in neurodegenerative diseases. *Dialogues Clin. Neurosci.* 20, 283–292. doi: 10.31887/DCNS.2018.20.4/knave
- Nave, K. A., and Werner, H. B. (2014). Myelination of the nervous system: mechanisms and functions. *Annu. Rev. Cell Dev. Biol.* 30, 503–533. doi: 10.1146/ annurev-cellbio-100913-013101
- Nieuwenhuys, R. (2013). The myeloarchitectonic studies on the human cerebral cortex of the Vogt-Vogt school, and their significance for the interpretation of functional neuroimaging data. *Brain Struct. Funct.* 218, 303–352. doi: 10.1007/ s00429-012-0460-z
- Ogg, R. J., and Steen, R. G. (1998). Age-related changes in brain T1 are correlated with iron concentration. *Magn. Reson. Med.* 40, 749–753. doi: 10.1002/mrm. 1910400516
- Orthmann-Murphy, J., Call, C. L., Molina-Castro, G. C., Hsieh, Y. C., Rasband, M. N., Calabresi, P. A., et al. (2020). Remyelination alters the pattern of myelin in the cerebral cortex. *Elife* 9:e56621. doi: 10.7554/eLife.56621
- Pajevic, S., Basser, P. J., and Fields, R. D. (2014). Role of myelin plasticity in oscillations and synchrony of neuronal activity. *Neuroscience* 276, 135–147. doi: 10.1016/j.neuroscience.2013.11.007
- Palomero-Gallagher, N., and Zilles, K. (2019). Cortical layers: cyto-, myelo-, receptor- and synaptic architecture in human cortical areas. *Neuroimage* 197, 716–741. doi: 10.1016/j.neuroimage.2017.08.035
- Palop, J. J., and Mucke, L. (2010). Amyloid-beta-induced neuronal dysfunction in Alzheimer's disease: from synapses toward neural networks. *Nat. Neurosci.* 13, 812–818. doi: 10.1038/nn.2583
- Peters, A. (2002). The effects of normal aging on myelin and nerve fibers: a review. *J. Neurocytol.* 31, 581–593. doi: 10.1023/a:1025731309829
- Polimeni, J. R., Fischl, B., Greve, D. N., and Wald, L. L. (2010). Laminar analysis of 7T BOLD using an imposed spatial activation pattern in human V1. *Neuroimage* 52, 1334–1346. doi: 10.1016/j.neuroimage.2010.05.005
- Poljak, A., Crawford, J. D., Smythe, G. A., Brodaty, H., Slavin, M. J., Kochan, N. A., et al. (2016). The relationship between plasma Aβ levels, cognitive function and brain volumetrics: sydney Memory and Ageing Study. *Curr. Alzheimer Res.* 13, 243–255. doi: 10.2174/1567205013666151218150202
- Preische, O., Schultz, S. A., Apel, A., Kuhle, J., Kaeser, S. A., Barro, C., et al. (2019). Serum neurofilament dynamics predicts neurodegeneration and clinical progression in presymptomatic Alzheimer's disease. *Nat. Med.* 25, 277–283. doi: 10.1038/s41591-018-0304-3
- Rembach, A., Watt, A. D., Wilson, W. J., Villemagne, V. L., Burnham, S. C., Ellis, K. A., et al. (2014). Plasma amyloid-β levels are significantly associated with a transition toward Alzheimer's disease as measured by cognitive decline and change in neocortical amyloid burden. J. Alzheimers Dis. 40, 95–104. doi: 10.3233/JAD-131802
- Rissin, D. M., Kan, C. W., Campbell, T. G., Howes, S. C., Fournier, D. R., Song, L., et al. (2010). Single-molecule enzyme-linked immunosorbent assay detects serum proteins at subfemtomolar concentrations. *Nat. Biotechnol.* 28, 595–599. doi: 10.1038/nbt.1641
- Salthouse, T. (2012). Consequences of age-related cognitive declines. *Annu. Rev. Psychol.* 63, 201–226. doi: 10.1146/annurev-psych-120710-100328
- Schindler, S. E., Bollinger, J. G., Ovod, V., Mawuenyega, K. G., Li, Y., Gordon, B. A., et al. (2019). High-precision plasma β -amyloid 42/40 predicts current and future brain amyloidosis. *Neurology* 93, e1647–e1659. doi: 10.1212/WNL. 000000000008081
- Schmued, L. C., Raymick, J., Paule, M. G., Dumas, M., and Sarkar, S. (2013). Characterization of myelin pathology in the hippocampal complex of a transgenic mouse model of Alzheimer's disease. *Curr. Alzheimer Res.* 10, 30–37. doi: 10.2174/1567205011310010005
- Seppälä, T. T., Herukka, S. K., Hänninen, T., Tervo, S., Hallikainen, M., Soininen, H., et al. (2010). Plasma Abeta42 and Abeta40 as markers of cognitive change

in follow-up: a prospective, longitudinal, population-based cohort study. *J. Neurol. Neurosurg. Psychiatry* 81, 1123–1127. doi: 10.1136/jnnp.2010.20 5757

- Sheikh, J. L., and Yesavage, J. A. (1986). Geriatric Depression Scale (GDS): recent evidence and development of a shorter version. *Clin. Gerontol.* 5, 165–173. doi: 10.1300/J018v05n01_09
- Shin, H. G., Lee, J., Yun, Y. H., Yoo, S. H., Jang, J., Oh, S. H., et al. (2021). χseparation: magnetic susceptibility source separation toward iron and myelin mapping in the brain. *Neuroimage* 240:118371. doi: 10.1016/j.neuroimage.2021. 118371
- Simone, M., Palazzo, C., Mastrapasqua, M., Bollo, L., Pompamea, F., Gabellone, A., et al. (2021). Serum neurofilament light chain levels and myelin oligodendrocyte glycoprotein antibodies in pediatric acquired demyelinating syndromes. *Front. Neurol.* 12:754518. doi: 10.3389/fneur.2021.754518
- Skoog, I., Kern, S., Zetterberg, H., Östling, S., Börjesson-Hanson, A., Guo, X., et al. (2018). Low cerebrospinal fluid Aβ42 and Aβ40 are related to white matter lesions in cognitively normal elderly. J. Alzheimers Dis. 62, 1877–1886. doi: 10.3233/JAD-170950
- Spencer, N. G., Bridges, L. R., Elderfield, K., Amir, K., Austen, B., and Howe, F. A. (2013). Quantitative evaluation of MRI and histological characteristics of the 5xFAD Alzheimer mouse brain. *Neuroimage* 76, 108–115. doi: 10.1016/j. neuroimage.2013.02.071
- Uddin, M. N., Figley, T. D., Solar, K. G., Shatil, A. S., and Figley, C. R. (2019). Comparisons between multi-component myelin water fraction, T1w/T2w ratio, and diffusion tensor imaging measures in healthy human brain structures. *Sci. Rep.* 9:2500. doi: 10.1038/s41598-019-39199-x
- van Dalen, J. W., Van Wanrooij, L. L., van Charante, E. P. M., Richard, E., and van Gool, W. A. (2018). Apathy is associated with incident dementia in community-dwelling older people. *Neurology* 90, e82–e89. doi: 10.1212/WNL. 000000000004767
- Verberk, I. M. W., Hendriksen, H. M. A., van Harten, A. C., Wesselman, L. M. P., Verfaillie, S. C. J., van den Bosch, K. A., et al. (2020). Plasma amyloid is associated with the rate of cognitive decline in cognitively normal elderly: the SCIENCe project. *Neurobiol. Aging* 89, 99–107. doi: 10.1016/j.neurobiolaging. 2020.01.007
- Verberk, I. M. W., Slot, R. E., Verfaillie, S. C. J., Heijst, H., Prins, N. D., van Berckel, B. N. M., et al. (2018). Plasma amyloid as prescreener for the earliest Alzheimer pathological changes. *Ann. Neurol.* 84, 648–658. doi: 10.1002/ana.2 5334
- Vidal-Piñeiro, D., Walhovd, K. B., Storsve, A. B., Grydeland, H., Rohani, D. A., and Fjell, A. M. (2016). Accelerated longitudinal gray/white matter contrast decline in aging in lightly myelinated cortical regions. *Hum. Brain Mapp.* 37, 3669–3684. doi: 10.1002/hbm.23267
- Vogt, O. (1903). Zur anatomischen Gliederung des Cortex cerebri. J. Psychol. Neurol. 2, 160–180.
- Wang, F., Ren, S. Y., Chen, J. F., Liu, K., Li, R. X., Li, Z. F., et al. (2020). Myelin degeneration and diminished myelin renewal contribute to age-related deficits in memory. *Nat. Neurosci.* 23, 481–486. doi: 10.1038/s41593-020-0 588-8

- Wang, X., and Cheng, Z. (2020). Cross-sectional studies: strengths, weaknesses, and recommendations. *Chest* 158, S65–S71. doi: 10.1016/j.chest.2020.03.012
- Waxman, S. G. (1980). Determinants of conduction velocity in myelinated nerve fibers. *Muscle Nerve* 3, 141–150. doi: 10.1002/mus.880030207
- Weinhofer, I., Rommer, P., Zierfuss, B., Altmann, P., Foiani, M., Heslegrave, A., et al. (2021). Neurofilament light chain as a potential biomarker for monitoring neurodegeneration in X-linked adrenoleukodystrophy. *Nat. Commun.* 12:1816. doi: 10.1038/s41467-021-22114-2
- Xu, Y. K. T., Call, C. L., Sulam, J., and Bergles, D. E. (2021). Automated in vivo tracking of cortical oligodendrocytes. Front. Cell. Neurosci. 15:667595. doi: 10.3389/fncel.2021.667595
- Yang, S. Y., Chiu, M. J., Chen, T. F., and Horng, H. E. (2017). Detection of plasma biomarkers using immunomagnetic reduction: a promising method for the early diagnosis of Alzheimer's disease. *Neurol. Ther.* 6, 37–56. doi: 10.1007/ s40120-017-0075-7
- Yasuno, F., Kazui, H., Morita, N., Kajimoto, K., Ihara, M., Taguchi, A., et al. (2017). Use of T1-weighted/T2-weighted magnetic resonance ratio to elucidate changes due to amyloid β accumulation in cognitively normal subjects. *Neuroimage Clin.* 13, 209–214. doi: 10.1016/j.nicl.2016.11.029
- Yeo, I. K., and Johnson, R. A. (2000). A new family of power transformations to improve normality or symmetry. *Biometrika* 87, 954–959. doi: 10.1093/biomet/ 87.4.954
- Yik, J. T., Becquart, P., Gill, J., Petkau, J., Traboulsee, A., Carruthers, R., et al. (2022). Serum neurofilament light chain correlates with myelin and axonal magnetic resonance imaging markers in multiple sclerosis. *Mult. Scler.* 57:103366. doi: 10.1016/j.msard.2021.103366
- Yuan, A., Rao, M. V., Veeranna, and Nixon, R. A. (2017). Neurofilaments and neurofilament proteins in health and disease. *Cold Spring Harb. Perspect. Biol.* 9:a018309. doi: 10.1101/cshperspect.a018309
- Zetterberg, H., and Burnham, S. C. (2019). Blood-based molecular biomarkers for Alzheimer's disease. *Mol. Brain* 12:26. doi: 10.1186/s13041-019-0448-1

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