# Appendixes Supporting information

# **Appendix 1 [Supporting information of Chapter 1](#page-4-0)**



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# **Appendix 1**

<span id="page-4-0"></span>Supporting information of Chapter 1

#### <span id="page-6-0"></span>**A1.1 Experimental section. General instrumentation**

**Elemental analyses** (C, H, N) were performed on a Leco CHNS-932 microanalyser.

**Thermogravimetric analysis** (TG/DTA) were performed on a TG-Q500 TA Instruments thermal analyser from room temperature to 800 °C under a synthetic air atmosphere (79 %  $N<sub>2</sub>/21$ % O<sub>2</sub>) at a heating rate of 10 °C min<sup>-1</sup>.

**FT-IR** spectra of the ligand and prepared coordination compound were collected in the region of 400–4000 cm−1 on a Nicolet 6700 FTIR (Fourier transform infrared) spectrophotometer (Thermo Phisher Scientific, TX, USA) KBr pellets.

**Photoluminescence measurements** at low temperature were done in an Edinburgh Instruments FLS920 spectrometer using a close cycle helium cryostat enclosed, in an applied vacuum (10<sup>-7</sup> bar). For steady state measurements in the UV-Vis range an IK3552R-G HeCd continuous laser (325 nm) was used as excitation source, whereas a Müller‐Elektronik‐Optik SVX1450 Xe lamp was employed to collect the excitation spectra. LDH-P-C-370 laser diode of PicoQuant was employed for recording the decay curves corresponding to the lifetimes of ns range.

**Chiroptical properties:** CD spectra were recorded at 25 °C on a Jasco J-815 or on an Olis DSM172 equipped with a Hamamatsu 150 W xenon arc lamp as light source. 1 cm pathlength cuvettes and the following parameters were used for data acquisition on Jasco J-815: data pitch 0.5 nm; digital integration time (D.I.T.) 1 sec; scanning speed 200 nm/min; bandwidth 1 nm; accumulations 4. HT voltage was controlled (HT ≤ 800) to give reliable ellipticity values over the investigated wavelength range. CD raw data were processed with Jasco Spectra Manager 2 (Jasco) and Origin 9.5 (OriginLab Corp.). In case of compounds Cd-L\_tyr (**1.8**) and Cd-D\_tyr (**1.9**) 1 mm length cuvette was employed.

**Circularly polarized luminescence** (CPL) measurements were performed on an Olis DSM172 spectrophotometer equipped with fixed wavelengths LED (300 nm) as light source. A Hamamatsu photon counting detector for CPL measurements. Different settings and data processing were selected and carried out with the aim of ruling out the presence of any artefacts due to anisotropies.

**Relaxation time measurements.** Proton relaxation times  $T_1$  and  $T_2$  were measured at 60 MHz in a Bruker Minispec mq60 TD-NMR spectrometer working at clinical MRI field (1.4 T). Mn\_gly (**1**) solutions of paramagnetic compound **1.1** was prepared at concentrations (1, 0.5, 0.1 mM relative to Mn<sup>2+</sup>) were analysed. Three different measurements of  $T_1$  and  $T_2$  were performed for each sample and each concentration. Relaxativity values *r*<sup>1</sup> and *r*<sup>2</sup> were obtained from the slopes of the curves  $1/T_1$  and  $1/T_2$  *vs*. the concentration of  $Mn^{2+}$  expressed in mM. MRI

scans were carried out in a preclinical 7-T magnet (Agilent, Palo Alto, CA, USA) interfaced to Avance III electronics, using a quadrature transmit-receive coil (Bruker, Ettlingen, Germany). *T<sup>1</sup>* values were estimated from images acquired using the rapid acquisition with relaxation enhancement (RARE) sequence with inversion recovery (IT = 50, 200, 400, 800, 1500, 3000, 5500, 8000, 12,000 ms, TE = 7.0 ms, echo train length 2, data matrix size 128  $\times$  64, field of view  $30 \times 15$  mm<sup>2</sup>, slice thickness = 3 mm, 1 scan).

The **X-ray powder diffraction** (XRPD) patterns were collected at 25 °C on a Phillips X'PERT powder diffractometer with Cu-Kα radiation ( $\lambda$  = 1.5418 Å) over the range 5 < 2θ < 50<sup>°</sup> with a step size of 0.02° and an acquisition time of 2.5 s per step. Indexation of the diffraction profiles were made by means of the FULLPROF program (pattern- matching analysis) based on the space group and the cell parameters found by single crystal X-ray diffraction.[1]

<span id="page-7-0"></span>**X-ray data collection** of suitable single crystals were done at 100(2) K on a Bruker D8 VENTURE area detector equipped with graphite monochromated Mo−Kα radiation  $(\lambda = 0.71073 \text{ Å})$  by applying the  $\omega$ -scan method. The data reduction was performed with the APEX270 software and corrected for absorption using SADABS.[2] Crystal structures were solved by direct methods using the SIR97 program[3] and refined by full-matrix least-squares on F2 including all reflections using anisotropic displacement parameters by means of the WINGX[4] crystallographic package. All hydrogen atoms were included as fixed contributions riding on attached atoms with isotropic thermal displacement parameters 1.2 times or 1.5 times those of their parent atoms for the organic ligands. Lattice solvent molecules could not be refined owing to their disordered disposition in the voids of the structures. so the electron density at the voids was subtracted from the reflection data by the SQUEEZE procedure as implemented in PLATON program[5] during the refinement.

# **A1.2 Chemical characterization**

# **A1.2.1 Elemental Analysis**





#### **A1.2.2 FT-IR spectroscopy**



**Figure A1.1.** Figure of the infrared spectra of glycine, N-cyanoethylglycine, and compound **1.1**.



**Figure A1.2.** Figure of the infrared spectra of L/D-.valine, N-cyanoethyl-L/D-valine and compounds **1.2** and **1.3**



**Figure A1.3.** Figure of the infrared spectra of L/D-phenylalanine, N-cyanoethyl-L/D-phenylalanine and compounds **1.4** and **1.5.**



Figure A1.4. Figure of the infrared spectra of L/D-.tyrosine, N-cyanoethyl-L/D-tyrosine and compounds **1.6-1.9.**

# **A1.3 Crystallographic data**

<span id="page-11-0"></span>

**Table A1.2.** Crystallographic data and structure refinement details of compounds.



$Mn$ gly (1.1)		Zn-L_val (1.2)				$Zn-D_val(1.3)$		
Mn1	O <sub>11</sub>	2.1644(13)	Zn1	N <sub>1</sub>	2.0450(17)	Zn1	N <sub>1</sub>	2.005(2)
Mn1	O <sub>1</sub> W	2.1709(13)	Zn1	N4 <sup>1</sup>	2.0067(17)	Zn1	N <sub>4</sub>	2.046(2)
Mn1	O <sub>2</sub> W	2.2373(14)	Zn1	N <sub>8</sub>	2.1258(16)	Zn1	N <sub>8</sub>	2.128(2)
Mn1	N <sub>8</sub>	2.3004(15)	Zn1	O11	2.1325(14)	Zn1	O11	2.1295(17)
Mn1	N <sub>1</sub>	2.2345(14)	Zn1	$O12^2$	2.0816(14)	Zn1	O11	2.1295(17)
Mn1	N2 <sup>1</sup>	2.2456(15)						
$11-x. -1-y. 1-z$			$11/2$ -x. 1-y. $1/2$ +z; $2-1/2+x$ . $1/2-y$ . 2-z					
		Zn-D_phen (1.5)			Zn-L_tyr (1.6)			Zn-D_tyr (1.7)
Zn1	O <sub>11</sub>	1.978(2)	Zn1	O <sub>12</sub>	2.162(2)	Zn1	O12 <sup>1</sup>	2.1555(19)
Zn1	N4	2.141(3)	Zn1	O11 <sup>1</sup>	1.964(2)	Zn1	O11	1.963(2)
Zn1	O <sub>12</sub>	2.117(3)	Zn1	N4 <sup>2</sup>	2.156(3)	Zn1	N4 <sup>2</sup>	2.150(3)
Zn1	N <sub>8</sub>	2.126(3)	Zn1	N <sub>8</sub>	2.112(2)	Zn1	N8 <sup>1</sup>	2.114(2)
Zn1	N <sub>1</sub>	2.014(3)	Zn1	N <sub>1</sub>	2.019(3)	Zn1	N1 <sup>1</sup>	2.017(3)
			Zn1	O <sub>12</sub>	2.162(2)	Zn1	O12 <sup>1</sup>	2.1555(19)
		$11/2+x$ . $1/2-y$ . -z; $21/2+x$ .-1/2-y.-z				$11/2+x.1/2-y.1-z$ ; $2+x.-1+y.+z$		

**Table A1.3.** Table of the selected bond lengths (Å) and angles (°) for compound **1.1**-**1.7**.





	$D-H\cdots A^b$	D-H	$H\cdots A$	D…A	D-HA
	O1W-H1WAO12 (i)	0.76(3)	1.92(3)	2.6800(19)	170(3)
	O1W-H1WBO3W	0.81(3)	1.88(3)	2.692(2)	178(3)
$Mn$ gly (1.1)	O2W-H2WBO12 (ii)	0.90(3)	1.76(3)	2.660(2)	174(3)
	O2W-H2WAN4 (iii)	0.78(3)	2.05(3)	2.827(2)	169(3)
	O3W-H3WAN3 (iv)	0.79(3)	2.08(3)	2.863(2)	168(3)
$Zn-L_{tyr}(1.6)$	O19-H19O1W (vi)	0.82	1.90	2.651(4)	150.8
Zn-D_tyr (1.7)	O19-H19O1W	0.84	1.82	2.640(3)	164.7

**Table A1.4.** Hydrogen bonding interactions (A. °)

<sup>a</sup>Symmetry codes: (i) -x+1. y-1/2. -z+1/2; (ii) x. -y-1/2. z+1/2; (iii) x+1. y. z; (iv) -x. -y-1. -z+1

(vi) 1/2-x.-y.1/2+z

**bD: donor. A: acceptor.** 

<span id="page-17-0"></span>

# **A1.4 Powder X-ray diffraction analysis**

**Figure A1.5.** Figure of the pattern matching analysis and experimental PXRD for complexes **1.1**-**1.6**.



**Figure A1.6.** Figure of the pattern matching analysis and experimental PXRD for complexes **1.7**-**1.9**.

# <span id="page-19-0"></span>**A1.5 Continuous Shape Measurements**



Table A1.5. Continuous Shape Measurements for the MN<sub>3</sub>O<sub>3</sub> coordination environment.

Structure $[MnN_3O_3]$	HP-6	PPY-6	OC-6	TPR-6	JPPY-6
$Mn$ gly $(1.1)$	33.464	25.371	1.030	12.467	29.058

**Table A1.6.** Continuous Shape Measurements for the MN3O<sup>2</sup> coordination environment**.**





<span id="page-20-0"></span>

# **A1.6 Thermal analysis**

Figure A1.7. Figure of TG/DTG analysis of compounds Mn\_gly, Zn\_val, Zn\_phen and Zn\_tyr.



# <span id="page-21-0"></span>**A1.7 Additional views of the structure**

Figure A1.8. Perspective view along *a. b* and *c* axis from left to right of compound 1.2<sub>Zn-L\_val.</sub>



Figure A1.9. Perspective view along *a. b* and *c* axis from left to right of compound 1.5<sub>Zn-D\_phen</sub>.



**Figure A1.10.** Perspective view along *a*. *b* and *c* axis from left to right of compound **1.6Zn-L\_tyr**.

<span id="page-22-0"></span>

# **A1.8 Photoluminescent properties**

**Figure A1.11.** Ambient temperature micro-photoluminescence images taken on polycrystalline sample of compound Zn-L\_tyr (**1.6**) at different excitation lines.

# <span id="page-23-0"></span>**A1.9 Relaxativity measurements**



**Table A1.7.** Raw data of measured relaxativity at 1.4 T. 0 mM concentration corresponds to milli-Q H<sub>2</sub>O relaxation time.

**Table A1.8.** Raw data of measured relaxativity at 7 T. 0 mM concentration corresponds to milli-Q H2O relaxation time.

<b>Concentration (mM)</b>	$T_1$ (ms)	$T_1$ error	$T_2$ (ms)	$T_2$ error
	265	4	21	
0.5	447	8	32	
0.1	1298	25	107	
	2127	51	256	2
1 of $MnCl2$	203		13	

# <span id="page-24-0"></span>**A1.10 Chiroptical properties. Statistical analysis of the CPL data**



**Table A1.9.** Integrated areas of the CPL spectra and the corresponding statistic parameters

**Table A1.10.** F-test two sample for variances corresponding to Zn-L\_phen (**1.4**)

$Zn-L_p$ hen (1.4)						
F-Test two sample for Variances	Variable 1	Variable 2				
Mean	$-0.13$	6.27E-04				
Variance	0.00	2.58E-06				
Observations	10.00	10				
Degrees of freedom	9.00	9				
F	100.41					
$P(F \le f)$ one tail	0.00					
<b>F</b> Critical one tail	3.18					

$Zn-D_p$ hen (1.5)						
F-Test two sample for Variances	Variable 1	Variable 2				
Mean	0.05	6.27E-04				
Variance	0.00	2.58E-06				
Observations	10.00	10				
Degrees of freedom	9.00	9				
F	107.99					
$P(F \le f)$ one tail	0.00					
<b>F Critical one tail</b>	3.18					

**Table A1.11.** *F-test* two sample for variances corresponding to Zn-D\_phen (**1.5**)

**Table A1.12***. t-student* test for two sample assuming equal variances for Zn-L\_phen (**1.4**)

$Zn-L_p$ hen (1.4)					
t-Test: Two-Sample Assuming Equal Variances	Variable 1	Variable 2			
Mean	$-0.13$	6.27E-04			
Variance	0.00	2.58E-06			
Observations	10.00	10.00			
Pooled Variance	0.00				
Hypothesized Mean Difference	0.00				
Degrees of freedom	18.00				
t Stat	$-26.48$				
$P(T \le t)$ one-tail	0.00				
T Critical one-tail	1.73				
$P(T \le t)$ two-tail	0.00				
t Critical two-tail	2.10				

**Table A1.13.** *t-student* test for two sample assuming equal variances for Zn-D\_phen (**1.5**)

$Zn-D_p$ hen (1.5)						
Method A	Method B					
0.05	0.00063					
0.00	$2.6E-06$					
10.00	10					
0.00						



# **Appendix 2**

<span id="page-28-0"></span>Supporting information of Chapter 2

#### <span id="page-30-0"></span>**A2.1 Experimental section. General instrumentation**

**Elemental analyses** (C, H, N) were performed on a Leco CHNS-932 microanalyser. Infrared (IR) spectra (400-4000 cm-1 ) were recorded on a Nicolet FT-IR 6700 spectrometer in KBr pellets.

**Thermogravimetric analysis** (TG/DTA) were performed on a TG-Q500 TA Instruments thermal analyser from room temperature to 800 °C under a synthetic air atmosphere (79 %  $N<sub>2</sub>/21$ % O<sub>2</sub>) at a heating rate of 10 °C min<sup>-1</sup>.

**Scanning electron microscopy** (SEM) images were acquired using either a Hitachi S4100 field emission gun tungsten filament instrument working at 25 kV or a high-resolution Hitachi SU-70 working at 4 kV. Samples were prepared by deposition on aluminium sample holders followed by carbon coating using an Emitech K950X carbon evaporator. EDS (energy dispersive X-ray spectroscopy) data and SEM mapping images were recorded using the latter microscope working at 15 kV and using either a Bruker Quantax 400 or an Esprit 1.9 EDS microanalysis system.

**Magnetic susceptibility** measurements were performed on polycrystalline samples of the complexes with a Quantum Design SQUID MPMS-7T susceptometer at an applied magnetic field of 1000 G. The susceptibility data were corrected for diamagnetism estimated from Pascal's tables,[6] the temperature-independent paramagnetism and magnetisation of the sample holder. The ac measurements were performed on a physical property measurement system quantum design model 6000 magnetometer under a 3.5 G ac field and frequencies ranging from 60 to 10000 Hz.

**Photoluminescence Spectroscopy**. The emission and excitation spectra were recorded at ambient-temperature and 12 K using a Fluorolog®-3Horiba Scientific (Model FL3-2T) spectroscope, with a modular double grating excitation spectrometer (fitted with a 1200 grooves/mm grating blazed at 330 nm) and a TRIAX 320 single emission monochromator (fitted with a 1200 grooves/mm grating blazed at 500 nm, reciprocal linear density of 2.6 nm·mm<sup>-1</sup>), coupled to a R928 Hamamatsu photomultiplier, using the front face acquisition mode. The excitation source was a 450 W Xe arc lamp. The emission spectra were corrected for detection and optical spectral response of the spectrofluorimeter and the excitation spectra were corrected for the spectral distribution of the lamp intensity using a photodiode reference detector. Timeresolved measurements have been carried out using a 1934D3 phosphorimeter coupled to the Fluorolog®-3, and a Xe-Hg flash lamp (6 μs/pulse half width and 20-30 μs tail) was used as the excitation source. The low temperature measurements (12 K) were performed using a heliumclosed cycle cryostat with vacuum system measuring ca.  $5 \times 10^{-6}$  mbar and a Lakeshore 330 autotuning temperature controller with a resistance heater.

N<sub>2</sub> (77 and 273 K) and CO<sub>2</sub> (273 and 298 K) **physisorption** data were measured in a Quantachrome Autosorb‐iQ MP, Prior to measurements, all samples were outgassed under vacuum at 150 °C for 6 hours. To estimate  $CO<sub>2</sub>$  adsorption enthalpies ( $Q<sub>st</sub>$ ), the isotherms were fitted to the modified Clausius−Clapeyron equation.

**X-ray data collection** of suitable single crystals were done at 100(2) K on a Bruker D8 VENTURE area detector equipped with graphite monochromated Mo−Kα radiation (λ = 0.71073 Å) by applying the ω-scan method. The data reduction was performed with the APEX270 software and corrected for absorption using SADABS.[2] Crystal structures were solved by direct methods using the SIR97 program[3] and refined by full-matrix least-squares on F2 including all reflections using anisotropic displacement parameters by means of the WINGX[4] crystallographic package. All hydrogen atoms were included as fixed contributions riding on attached atoms with isotropic thermal displacement parameters 1.2 times or 1.5 times those of their parent atoms for the organic ligands. Lattice solvent molecules could not be refined owing to their disordered disposition in the voids of the structures, so the electron density at the voids was subtracted from the reflection data by the SQUEEZE procedure as implemented in PLATON program[5] during the refinement.

The **X-ray powder diffraction** (XRPD) patterns were collected at 25 °C on a Phillips X'PERT powder diffractometer with Cu-Kα radiation ( $\lambda$  = 1.5418 Å) over the range 5 < 2θ < 50<sup>°</sup> with a step size of 0.02° and an acquisition time of 2.5 s per step. Indexation of the diffraction profiles were made by means of the FULLPROF program (pattern- matching analysis) based on the space group and the cell parameters found by single crystal X-ray diffraction.[1]

**Variable-temperature powder X-ray diffraction** measurements were conducted on a Bruker D8 Advance diffractometer, using polycrystalline sample of compound 2.7<sub>Dy</sub> under ambient atmosphere with heating rate of  $5^{\circ}$ C·min<sup>-1</sup> and measuring a complete diffractogram every 20 °C up to 510 °C, and every 50 °C from 510 °C up to 710 °C.

**Catalytic studies**: All experiments involving moisture-sensitive compounds were performed under an inert atmosphere of  $N_2$  using standard techniques. Unless otherwise indicated, reagents and substrates were purchased from commercial sources and used as received. Solvents not required to be dry were purchased as technical grade and used as received. Conversion values relative to the limiting reagent were calculated from the <sup>1</sup>H NMR spectra of the reaction crudes. Isolated products were obtained after centrifugation (8000 rpm, 3 min) and washed with dichloromethane  $(2 \times 0.5 \text{ mL})$  in order to remove the catalyst or column chromatography in silica gel using hexane as eluent.

**NMR measurements:** NMR spectra were measured in a Bruker Avance III 300 spectrometer equipped with a direct double SmartProbe BBFO <sup>1</sup>H/BB(<sup>19</sup>F) probe. Chemical shifts are reported in parts per million (ppm) relative to residual solvent peak (CDCl<sub>3</sub>, <sup>1</sup>H: 7.26 ppm; <sup>13</sup>C: 77.16 ppm). Coupling constants are reported in Hertz. Multiplicity is reported with the usual abbreviations (s: singlet, bs: broad singlet, d: doublet, dd: doublet of doublets, ddd: doublet of doublet of doublets, t: triplet, td: triplet of doublets, q: quartet, dq: doublet of quartet, p: pentet, sex: sextet, hept: heptet, m: multiplet).

**Dynamic water vapour sorption** (DVS) measurements were performed on a DVS Resolution water vapour analyzer (Surface Measurement Systems™), at 25 °C and from 0 to 95 % RH, with steps of 10 % RH, using 200 SCCM N2 (N50, purity ≥ 99.999%) as carrier gas. Both the sorption and desorption curves were recorded by setting a stability criterion for the mass change (gravimetric precision of 0.1  $\mu$ g) as the minimum in the variation of the mass over the time variation (d*m*/d*t*) of 0.002 % min-1 , or a maximum stage time at each RH of 360 min (in the case when the dm/dt minimum was not attained). Prior to the measurements, the sample of 2.6<sub>Tb</sub> was pretreated *in situ* in at 75 °C for 2 h at 0% RH and then at 25 °C for 1 h at 0 % RH, to ensure dehumidification for an initial reference state. An equivalent treatment was used for samples **2.6Tb@PSF** and **2.6Tb@PMMA** but with the first isothermal dwell at 60 °C to prevent softening of the polymer matrices.

The electrical conductivity (*σ*) of pelletized sample **2.6Tb** and **composite membranes (2.6Tb@PSF** and **2.6Tb@PMMA)** were studied by impedance spectroscopy using an Agilent E4980A Precision LCR meter. Disc shaped sample 1 was obtained after pressing the powder in a uniaxial press at 10 MPa, and then isostatically at 200 MPa. Silver electrodes were applied on both sides of the pellet and membranes (with an area of approximately  $1x1$  cm<sup>2</sup>) by painting a commercial paste (Agar Scientific). Samples were placed on ceramic tubular sample holders (equipped with platinum wires for current collection) inside a climatic chamber (ACS DY110) in order to carry out the measurements under variable temperature (40-94 °C) and relative humidity (RH, 20-95%). The impedance spectra were collected between 20 Hz and 2 MHz with a test signal amplitude of 100 mV. The current collection was ensured by separate platinum wires for voltage and current. The spectra were analyzed with ZView (Version 2.6b, Scribner Associates) to assess the ohmic resistance  $(R)$ , which was then normalized to the samples geometry to calculate the conductivity using Equation A2.1:

$$
\sigma = L(RA)^{-1}
$$

#### −1 **Equation A2.1**

where  $L$  is the sample thickness and  $A$  is the surface area of the electrodes. The maximum relative error in the conductivity data is estimated to be of the order of 4% through the conventional chain rule of differentiation of Equation A2.1 and the uncertainties in the measured parameters  $(\Delta R = 0.1\%$  to 0.3% of *R*,  $\Delta L = \pm 0.002$  cm,  $\Delta A = \pm 0.005$  cm).

# <span id="page-33-0"></span>**A2.2 Chemical characterization**

**2.9** C51H64N9O29Er<sup>5</sup> 2103.41

**2.10** C51H64N9O29Tm<sup>5</sup> 2111.78

**2.11** C51H64N9O29Yb<sup>5</sup> 2132.31

**2.12** C51H64N9O29Y<sup>5</sup> 1711.64

**2.19** C57H74N11O29Eu<sup>5</sup> 2137

#### **A2.2.1 Elemental analysis**



5.99; O: 22.06; Er:

C: 29.01; H: 3.05; N: 5.97; O: 21.97; Tm:

C: 28.73; H: 3.03; N: 5.91; O: 21.76; Yb:

C: 35.79; H: 3.77; N: 7.36; O: 27.11; Y:

C: 32.04; H: 3.49; N: 7.21; O: 21.71; Eu: 6.00; O: 22.08; Er:

C: 29.03; H: 3.06; N: 5.97; O: 21.97; Tm:

C: 28.74; H: 3.06; N: 5.93; O: 21.76; Yb:

C: 35.82; H: 3.74; N: 7.41; O: 27.16; Y:

C: 31.98; H: 3.51; N: 7.19; O: 21.68; Eu:

39.82;

40.02;

40.56;

26.02

35.54

39.76;

40.00;

40.58;

25.97

35.55

**Table A2.1.** Elemental analysis of compounds **2.2Nd**-**2.11Yb**

#### **A2.2.2 Determination of the metal content by ICP-AES**



**Table A2.2.** ICP-AES results of compounds **2.16-2.18.**

#### **A2.2.3 FT-IR spectroscopy**

FTIR spectra of compound **2.1Co** exhibit broad and intense band around 3439 cm-1 that corresponds to the O–H bond vibration of the of 3-amino-4-hydroxybenzoate free ligand, The bands between 3331 cm<sup>-1</sup> and 2928–2857 cm<sup>-1</sup> can be attributed to aromatic ring's C–H bond vibrations of the ligand. The intense vibrations in the 1668–1430 cm<sup>-1</sup> region are referred to both the asymmetric stretching vibrations of the carboxylate groups and the aromatic C–C and C–N bonds. The symmetric stretching vibrations of the carboxylate groups appear in the lower range of 1391–1297 cm<sup>-1</sup>. The remaining bands that are found at lower frequency can be attributed to the distortions originated in the aromatic ring and the carboxylate groups of the ligands. The vibration bands of the M–O and M–N bonds are observed below 670 cm-1 .

FTIR spectra of compound **2.7Dy** display a narrow peak at around 3625 cm-1 , attributed to the N-H stretching vibration of amine group, which is practically hided below the intense broad band around 3412 cm<sup>-1</sup> attributed to O-H bond vibration of the of 3-amino-4-hydroxybenzoate free ligand.

At lower frequency, a set of intense bands are visible between  $3207 \text{ cm}^{-1}$  and  $2921-$ 2830 cm<sup>-1</sup> which corresponds to aromatic ring's C-H bond vibrations of the ligand The intense vibrations in the 1661–1433 cm<sup>-1</sup> region are referred to both the asymmetric stretching vibrations of the carboxylate groups and the aromatic C–C and C–N bonds. The symmetric stretching vibrations of the carboxylate groups appear in the lower range of  $1381-1281$  cm<sup>-1</sup>. The remaining bands that are found at lower frequency can be attributed to the distortions originated in the aromatic ring and the carboxylate groups of the ligands. The vibration bands of the M-O and M-N bonds are observed below 646 cm<sup>-1</sup>.



**Figure A2.1.** Infrared spectra of the ligand and compound 2.7<sub>Dy</sub>.
## **A2.3 Crystallographic data**



**Table A2.3.** Crystallographic data and structure refinement details of compounds **2.1** and **2.7**.

<b>Complex</b>		2.1 <sub>co</sub>			2.7 <sub>Py</sub>			2.12 <sub>Y</sub>			2.19 <sub>Eu</sub>
Atom1-2		Length/Å	Atom1-2		Length/Å		Atom1-2	Length/Å	Atom1-2		Length/Å
Co1	O <sub>2</sub>	1.8610(157)	Dy1	Dy1 <sup>1</sup>	3.4984(12)	Y1	O <sub>3</sub>	2.536(5)	Eu1	Eu1 <sup>1</sup>	3.5922(4)
	O <sub>3</sub>	1.9145(173)	Dy1	$Dy2^2$	3.9242(6)	Y1	N <sub>1</sub>	2.517(6)	Eu1	O <sub>1</sub>	2.5265(17)
	O <sub>1</sub>	1.9367(242)	Dy1	Dy <sub>2</sub>	3.9242(6)	Y1	O <sub>1</sub> H	2.349(4)	Eu1	O1 <sup>2</sup>	2.5266(17)
	O <sub>1</sub>	2.0802(241)	Dy1	O1H <sup>3</sup>	2.366(5)	Y2	O <sub>1</sub>	2.425(5)	Eu1	O1 <sup>3</sup>	2.5265(17)
	N <sub>1</sub>	2.4091(169)	Dy1	O <sub>1</sub> H	2.366(5)	Y2	O <sub>2</sub>	2.396(5)	Eu1	N <sub>1</sub>	2.546(2)
	O <sub>2</sub>	3.1768(204)	Dy1	O1H <sup>2</sup>	2.366(5)	Y2	O <sub>3</sub>	2.349(5)	Eu1	N1 <sup>3</sup>	2.546(2)
	Co <sub>1</sub>	3.304(14)	Dy1	O <sub>3</sub>	2.513(6)	Y2	O <sub>1</sub> D	2.305(7)	Eu1	N1 <sup>2</sup>	2.546(2)
	O <sub>3</sub>	3.3833(160)	Dy1	O3 <sup>2</sup>	2.513(6)	Y2	O <sub>1</sub> H	2.348(6)	Eu1	Eu2 <sup>2</sup>	3.94656(18)
	N <sub>1</sub>	3.5469(246)	Dy1	O3 <sup>3</sup>	2.513(6)				Eu1	Eu <sub>2</sub>	3.94655(18)
			Dy1	N <sub>1</sub>	2.509(7)				Eu1	O4 <sup>3</sup>	2.4145(16)
			Dy1	N1 <sup>3</sup>	2.510(7)				Eu1	O <sub>4</sub>	2.4145(16)
			Dy1	N1 <sup>2</sup>	2.510(7)				Eu1	O4 <sup>2</sup>	2.4485(18)
			Dy <sub>2</sub>	O1 <sup>4</sup>	2.394(6)				Eu <sub>2</sub>	O2 <sup>4</sup>	2.4485(18)
			Dy <sub>2</sub>	O1 <sup>5</sup>	2.394(6)				Eu <sub>2</sub>	O2 <sup>5</sup>	2.4675(18)
			Dy <sub>2</sub>	O <sub>1</sub> H	2.337(7)				Eu <sub>2</sub>	O3 <sup>5</sup>	2.4675(18)
			Dy <sub>2</sub>	O2 <sup>4</sup>	2.417(6)				Eu <sub>2</sub>	O3 <sup>4</sup>	2.349(2)
			Dy <sub>2</sub>	O2 <sup>5</sup>	2.417(6)				Eu <sub>2</sub>	O <sub>4</sub>	2.385(3)
			Dy <sub>2</sub>	O <sub>3</sub>	2.371(5)				Eu <sub>2</sub>	O <sub>5</sub>	2.3837(16)
			Dy <sub>2</sub>	O3 <sup>1</sup>	2.371(5)				Eu <sub>2</sub>	O <sub>1</sub>	2.4485(18)
			Dy <sub>2</sub>	O <sub>4</sub>	2.327(11)						
					$1+x, +y, 3/2-z$ ; $21-y, 1+x$ - $y, +z$ ; $3+y-x, 1-x, +z$ ; $4+y, 1-z$ $x+y,1/2+z$ ; $5+y,1-x+y,1-z$						$1+x, +y, 3/2-z$ ; $21-y, 1+x$ - $y, +z$ ; $3+y-x, 1-x, +z$ ; $4+y, 1-z$ $x+y, 1-z$ ; $5+y, 1-x+y, 1/2+z$

**Table A2.4.** Table of the selected bond lengths (Å) for compound **2.1Co**, **2.7Dy**, **2.12<sup>Y</sup>** and **2.19Eu**









$D-H\cdots A^a$		$D-H$ $H\cdots A$ $D\cdots A$		D-H <sub>.</sub> A			
N1-H1AO1 <sup>1</sup> 0.91 2.10 2.951 (10) 155.8							
<sup>a</sup> D: donor. A: acceptor. $11-x, 1-y, 1-z$							

**Table A2.5.** Hydrogen bonding interactions (Å. <sup>o</sup>) of compound 2.7<sub>Dy</sub>

## **A2.4 Powder X-ray diffraction analysis**



**Figure A2.2.** Figure of the pattern matching analysis and experimental PXRD for complex **2.1Co.**



Figure A2.3. Experimental PXRD of [Co<sub>x</sub>Zn<sub>1-x</sub>L]<sub>n</sub> heterometallic samples.





**Figure A2.4.** Figure of the pattern matching analysis and experimental PXRD for complexes **2.2- 2.11.**



Figure A2.5. Figure of the theoretical simulated-PXRD for complexes 2.7<sub>Dy</sub> and 2.12<sub>Y</sub>.



**Figure A2.6.** PXRD for complexes **2.12-2.15.**

When the quaternary compounds (containing yttrium. europium and terbium ions and the ligand) are characterized, it is observed that, depending on the  $Y^{3+}$  to  $Ln^{3+}$  doping proportion. PXRD patterns present diffraction maxima corresponding to both pure compounds **2.12** and **2.2**- **2.11.** This fact, *a priori*, is indicative of a crystal phase segregation, although it makes no much sense given the isostructural nature of the compounds. Nevertheless, SEM mapping experiments have shown that even if mixture of two type of crystals could happen, the three elements are randomly distributed along a single crystal (Figure A2.17 and Figure A2.18).



**Figure A2.7.** PXRD for complexes **2.4, 2.12, 2.12, 2.16-2.18.**

## **A2.5 Continuous Shape Measurements**

CShMs for the coordination environment of compound **2.1** and **2.7**. The lowest SHAPE values for each ion are shown highlighted in bold, indicating best fits.

SP-4	D <sub>4h</sub>	Square			
$T-4$	Td	Tetrahedron			
$SS-4$	$C_{2v}$	Seesaw			
vTBPY-4	$\mathrm{C}_{3\mathrm{v}}$	Vacant trigonal bipyramid			
<b>Complex</b>	$SP-4$	T-4	$SS-4$	<b>vTBPY-</b>	
2.1 <sub>co</sub>	30.170	1.878	4.528	2.270	

Table A2.6. Table of the continuous Shape Measurements for the CoO<sub>4</sub> coordination environment.

**Table A2.7.** Table of the continuous Shape Measurements for the CoN<sub>2</sub>O<sub>4</sub> coordination environment.

$HP-6$ PPY-6	$D_{6h}$ $C_{5v}$		Hexagon Pentagonal pyramid				
OC-6	Oh		Octahedron				
TPR-6	$D_{3h}$		Trigonal prism				
JPPY-6	$C_{5v}$		Johnson pentagonal pyramid J2				
<b>Complex</b>	$HP-6$	PPY-6	OC-6	TPR-6	JPPY-6		
2.1 <sub>co</sub>	34.045	12.509	12.883	7.490	15.585		

Table A2.8. Table of the continuous Shape Measurements for the LnN<sub>3</sub>O<sub>6</sub> coordination environment.



$HH-9$ MFF-9	$C_{2v}$ $\mathsf{C}_\mathsf{s}$	Hula-hoop <b>Muffin</b>			
<b>Complex</b>	<b>JCSAPR-9</b>	<b>CSAPR-9</b>	<b>JTCTPR-9</b>	TCTPR-9	MFF-9
$2.7Py$ (Dy1)	2.280	1.243	2.073	0.890	1.630
$2.12_Y (Y1)$	2.372	1.493	1.855	0.829	2.180
$2.19Eu$ (Eu1)	2.315	1.361	1.904	0.666	2.050

**Table A2.9.** Table of the continuous Shape Measurements for the LnO<sub>8</sub> coordination environment





#### **A2.6 Thermal analysis**

Thermogravimetric analyses have been performed over polycrystalline sample in compound **2.1, 2.7, 2.12** and **2.19** in order to check the stability of the product.

The TG curves for **2.1** and **2.7** has been collected before and after solvent exchange with MeOH. Solvent exchange procedure has been accomplished as an approach to replace lattice-solvent molecules (dimethylformamide and water) to ease material activation to posteriorly analyse its adsorptive-capacity. Powder X-ray diffraction confirmed that compounds **2.1** and **2.7**  remains stable after solvent exchange with MeOH as it can be seen in Figure A2.8 and Figure A2.9, down.



**Figure A2.8.** Figure of TG/DTG analysis of compound **2.1** (left -up, as synthesised, right -up, after solvent exchange with MeOH) and figure of the experimental PXRD for complex **2.1** before and after solvent exchange with MeOH (down).

More concretely, TG curve of Co\_MOF (**2.1**) shows two main steps of weight loses. The first step concerns to the progressive loss of the solvent lattice molecules (DMF and H<sub>2</sub>O) which are released from room temperature up to 300ºC. Then, an abrupt descent can be seen that corresponds to the collapse of the crystal structure. From the shape of the TG curve, it seems that solvent molecules stabilise the structure and their removal promote crystal structure decomposition. However, TG curve of Co\_MOF (**2.1**) after solvent exchange with methanol (this procedure has been carried out suspending three times the material in MeOH for an hour) shows two well defined steps. The first step comprises the loss of lattice solvent molecules, which occurs at 100 ºC, and agrees with an efficient DMF to MeOH exchange. Subsequently, from 110 ºC to 300 °C the TG curve of the compound describes a plateau, where the empty skeleton of the MOF is gotten. Finally, at 300 ºC ligand decomposition occurs and involves the collapse of the crystal structure, evolving to Co<sub>3</sub>O<sub>4</sub> that is obtained at 800 °C as the final residue.

The TG curves has been collected for compound 2.7<sub>Dy</sub> before and after solvent exchange with MeOH. This procedure has been carried out suspending the material in MeOH for 16 h. The thermal behaviour of the bulk [Dy<sub>5</sub>L<sub>6</sub>(OH)<sub>3</sub>(DMF)<sub>3</sub>] $\cdot$ 5H<sub>2</sub>O, compound 2.7<sub>Dy</sub>, as synthesised materials were also investigated between ambient temperature and ca. 800 ◦C in order to study its thermal stability. Due to the isotypical nature of the compounds the following paragraph discussion will be solely focused on compound **2.7Dy**. Three main regions are appreciable in the TG curve. The first weight loss, between ambient temperature and up to 300 ºC concerns to the progressive loss of solvent molecules, firstly, lattice, water molecules and then coordinated DMF molecules. Above this temperature there is an abrupt descent that corresponds to the collapse of the crystal structure. From the shape of the TG curve, it seems that solvent molecules stabilise the structure and their removal promote crystal structure decomposition. In the final step as a consequence of the decomposition of the organic content metal oxide is obtained.



Figure A2.9. Figure of TG/DTG analysis of compound 2.7<sub>Dy</sub> (up-left, as synthesised). after solvent exchange with MeOH during 16 h (up-right) and figure of the experimental PXRD for complex **2.7Dy** before and after solvent exchange with MeOH (down).

After solvent exchange with MeOH, TG curve performed in 2.7<sub>Dy</sub>\_MeOH shows a plateau at around 100 ºC suggesting that solvent molecules (DMF and water) have been properly replaced with MeOH which would evaporate up to the indicated temperature. Around 100-200 ºC it seems that the MOF desolvatated skeleton is obtained. Finally, as another evidence for the proper solvent exchange, final-residue  $Dy_2O_3$  percentage has increased by 6 %. It is expected since DMF and water molecules are heavier and have higher boiling points than MeOH.

Thermogravimetric analyses have been performed over polycrystalline sample in compounds **2.12<sup>Y</sup>** show two main steps of weight loses. The first steps concerns to the release of the water lattice molecules which are released from room temperature up to 275 ºC. Then, it starts the loss of coordinated DMF molecules, which is somewhat overlapped with the third step corresponding to the decomposition of the ligands. The latter involves the collapse of the crystal structure, evolving to  $Y_2O_3$  that is obtained at 800 °C as the final residue. On its part, thermogravimetric analyses have been performed over polycrystalline sample in compounds **2.19** exhibit two main steps of weight loses. The first steps concern to the release of the solvent molecules which are released from room temperature up to 275 ºC. The second step refers to the



decomposition of the ligands which involves the collapse of the crystal structure, evolving, to Eu2O<sup>3</sup> obtained at 800 ºC as the final residue.

**Figure A2.10.** Figure of TG analysis of compounds **2.12<sup>Y</sup>** (left) and **2.19Eu** (right).

 $T (°C)$ 

### **A2.7 Thermal evolution**

Thermal evolution of **2.1Co** shows that the compound maintains its crystallinity up to 300 ºC. These results come in line with thermogravimetric analysis, where it could be seen that above this temperature structure collapses and evolves into the metallic residue  $Co_3O_4$  at around 800 °C.



**Figure A2.11.** Thermal evolution of compound **2.1Co**.

Thermal evolution of compound 2.7<sub>Dy</sub> shows that the compound maintains its crystallinity up to 230 °C. These results come in line with thermogravimetric analysis, where it could be seen that above this temperature structure collapses and evolves into the metallic residue  $Dy_2O_3$  at around 800 ºC.



Figure A2.12. Thermal evolution of compound 2.7<sub>Dy</sub>.

## **A2.8 Additional views of the structure**



**Compound 2.1**

**Figure A2.13.** View along *a, b* and *c* axis (from left to right) of complex **2.1**Co.



**Figure A2.14.** View of pentametallic nodous showing Dy1 and Dy2 coordination polyhedral.



**Compound 2.7**

**Figure A2.15.** View along *a, b* and *c* axis (from left to right) of complex the topological representation along *a*. *b* and *c* axis (down).



# **A2.9 Scanning electron Microscopy**









Figure A2.16. SEM-EDS spectrum performed in single crystal of  $[Co_{0.9}Zn_{0.1}L]_n$ .





**Figure A2.17.** SEM images of compound **2.17Y-Tb-Eu10%** where two types of crystal are distinguished.



representative of the crystal type 1 of compound **2.17Y-Tb-Eu10%**.





 $2 \mu m$ 



**Figure A2.19.** SEM image, EDS spectrum and elemental quantitative data representative of the crystal type 2 of compound **2.17Y-Tb-Eu10%.**



**Figure A2.20.** Cross section EDS mapping of **2.6Tb@PMMA** (up) and **2.6Tb @PSF** (buttom) membranes.



**A2.10** *Ac* **magnetic susceptibility measurements**

**Figure A2.21.** Temperature dependence of in-phase (blue) and out of phase (red) components of the *ac* susceptibility in a *dc* applied field of 1000 Oe for **2.1Co**.



**Figure A2.22.** Plot of  $ln(\chi_M''/\chi_M')$  versus 1/T at 10000 Hz for compound  $2.1<sub>Co</sub>$  under an applied field of 1000 Oe. The solid lines represent the linear fit with  $\ln(\chi_M''/\chi_M') = \ln(2\pi v \tau_0) + E_a/k_B T$ .

**Table A2.10.** NEVPT2- calculated on the fully optimized structures D, E/D, g-tensor and energyseparation between KD1-KD2 and KD1-KD3.

<b>Parameters</b>	TPR	Τd
	$-58.7$ ( $-48.6$ , $46.7$ )	$-31.4(25.2, -28.1)$
$D$ (D <sub>KD1-2</sub> , D <sub>KD1-3</sub> )/ cm <sup>-1</sup> F/D	O 21	0 18
$g_{xx}$ , $g_{yy}$ , $g_{zz}$	2.08, 2.45, 2.93	2.16, 2.28, 2.6
$\Delta E(1-2)$ , $\Delta E(1-3)$ / cm <sup>-1</sup>	2367.2, 5325.4	3432.7.4453.7



Figure A2.23. Magnetic axes of the Dy<sup>III</sup> ions calculated with the Magellan software for compound **2.7**.[7]



**Figure A2.24.** Temperature dependence of in-phase (red) and out-of-phase (blue) components of the *ac* susceptibility in a zero applied dc field for **2.7Dy**.



**Figure A2.25.** Temperature dependence of in-phase (red) and out-of-phase (blue) components of the ac susceptibility in a zero applied dc field for **2.9Er**.



**Figure A2.26.** Temperature dependence of in-phase components of the *ac* susceptibility in a *dc* applied field of 1000 Oe for **2.11Yb**.



**Figure A2.27.** Cole-Cole plots in a *dc* applied field of 1000 Oe for **2.11Yb**.



Figure A2.28. Variable-temperature frequency dependence of the χ<sub>M</sub>" signal under 1000 Oe applied field for **2.11Yb**. Solid lines represent the best fitting of the experimental data to the Debye model.



**Figure A2.29.** Temperature dependence of in-phase (blue) and out of phase (red) components of the *ac* susceptibility in a *dc* applied field of 1000 Oe for **2.13Y-Dy**.



**Figure A2.30.** Plot of  $\ln(\chi_M''/\chi_M')$  versus 1/T at 10000 Hz for compound 2.13<sub>Y-Dy</sub> under an applied field of 1000 Oe. The solid lines represent the linear fit with  $\ln(\chi_M''/\chi_M') = \ln(2\pi v \tau_0) + E_a/k_B T$ .



**Figure A2.31.** Temperature dependence of in-phase components of the *ac* susceptibility in a *dc* applied field of 1000 Oe for **2.14Y-Er**.



**Figure A2.32.** Cole-Cole plots in a *dc* applied field of 1000 Oe for **2.14Y-Er**.



**Figure A2.33.** Variable-temperature frequency dependence of the *χM"* signal under 1000 Oe applied field for 2.14<sub>Y-Er</sub>. Solid lines represent the best fitting of the experimental data to the Debye model.



**Figure A2.34.** Temperature dependence of in-phase components of the *ac* susceptibility in a *dc* applied field of 1000 Oe for **2.15Y-Yb**.



**Figure A2.35.** Cole-Cole plots in a *dc* applied field of 1000 Oe for **2.15Y-Yb**.



Figure A2.36. Variable-temperature frequency dependence of the x<sub>M</sub>" signal under 1000 Oe applied field for **2.15Y-Yb**. Solid lines represent the best fitting of the experimental data to the Debye model.

#### **A2.11 Diffuse reflectance measurements**

Absorption spectrum of 3-amino-4-hydroxybenzoic acid ligand, pure Co<sup>II</sup> and Zn<sup>II</sup> homometallic compounds and  $[Co<sub>0.05</sub>Zn<sub>0.95</sub>L]<sub>n</sub>$  heterometallic compound show absorption bands in the range of 220–800 nm. Three main regions can be differentiated, the first one from 220–300 nm attributed to the ligand 3-amino-4-hydroxybenzoic acid ligand π–π\* transitions, the shoulder at around 310 nm (clearly observed for both homometallic and heterometallic compounds) corresponds to metal-to-ligand charge transfer (MLCT) transitions. Moreover, the bands with a broad and structured shape covering the 400–700 nm region, are attributed to spin-allowed d-d transitions found in heterometallic [Co<sub>0.05</sub>Zn<sub>0.95</sub>L]<sub>n</sub> and homometallic Co<sup>II</sup> compounds.



**Figure A2.37.** Diffuse reflectance of 3-amino-4-hydroxybenzoic acid ligand, homometallic Co\_MOF (**2.1**), and Zn compounds and heterometallic [Co0.05Zn0.95L]<sup>n</sup> heterometallic sample.



#### **A2.12 Photoluminescence measurements**

**Figure A2.38.** Figure caption of the experimental room temperature photoluminescence excitation and emission spectra under  $\lambda_{em}$  = 391 nm and  $\lambda_{ex}$  = 330 nm, respectively for compounds 1 and isostructural Zn<sup>II</sup> homometallic counterpart.

The normalized excitation and emission spectra of compound 2.1 and isostructural [ZnL]<sub>n</sub> compounds are sown in Figure A2.38.

The excitation spectra have been measured for both homometallic compounds by monitoring the emission maxima, at 391 nm. As observed in Figure A2.37, the excitation spectra show a band covering the 275–350 nm range in which four peaks (sited at ca. 288, 310, 322 and 334 nm) are distinguished.

Ambient temperature emission spectra monitored at the excitation maxima, at 330 nm, show a main band with the maxima at 362 nm and 391 nm and a tail. In case of homometallic  $Zn<sup>II</sup>$ compound a more prominent shoulder can be appreciable peaking at 447 nm.

Normalization of the spectra has been carried out in order to compare the position of the maxima in both structures more than to compare the relative intensity, which in case of  $Co<sup>II</sup>$  was, as expected, relatively weaker.

Compound	$\tau_1/ms$	$\tau_2$ / ms	$<\tau$ > / ms
$2.6fb$ (377)	$0.08 \pm 0.01$	$0.36 \pm 0.01$	0.29
$2.6_{\text{Tb}}@$ PMMA at 294 K (315)	$0.10 \pm 0.01$	$0.40 \pm 0.01$	0.35
$2.6_{\text{fb}}@$ PMMA at 12 K (315)	$0.09 \pm 0.01$	$0.38 \pm 0.01$	0.33

**Table A2.11.** Comparison of lifetime values of compound **2.6Tb** in bulk and PMMA membrane at ambient temperature (294 K) and low temperature (12 K).



**Figure A2.39.** Ambient temperature micro-photoluminescence images taken on single-crystal of compound 2.6<sub>Tb</sub> at different excitation lines.

#### **A2.13 Adsorption properties**



**Figure A2.40.** Isotherm of compound 2.7<sub>Dy</sub> in cm<sup>3</sup>/g. The conversion to mmol/g has done taking into account that 1 mmol of any gas at stp (standard temperature and pressure conditions. according to the IUPAC. at 273 K and 1 bar pression conditions) occupies 22.414 cm<sup>3</sup> volume.



**Figure A2.41.** Isosteric heats of adsorption (Q<sub>st</sub>) of CO<sub>2</sub> per Dy<sub>5</sub> cluster for compound 2.7<sub>Dy</sub> (left) and the deconvolution of the curve (right).


Figure A2.42. Isosteric heat of adsorption of CO<sub>2</sub> loading (mmol/g) per Dy<sub>5</sub> cluster.



Figure A2.43. High pressure CO<sub>2</sub> adsorption-desorption regeneration cycles for compound 2.7 at 273 K**.**



**Figure A2.44.** Derivative Geometric Pore Volume (pore size distribution) and cumulative volume in anhydrous compound 2.7<sub>Dy</sub>.

		Area & Vol			Pore diameter		
Compound	S <sub>area</sub> $(m^2 \cdot q^{-1})$	$\bm{{\mathsf{V}}}_\mathsf{pore}$ $(cm3·q-1)$	<b>Porosity</b> (%)	Limiting 'A	<b>Maximun</b> A	<b>Dimension</b> ality	
2.7 <sub>Dv</sub> anhydrous	713.2	0.319	50.9	4.89	6.35	3 D	

**Table A2.12.** Calculations performed by Monte Carlo procedure.[8,9]

### **A2.14 Catalytic activity. Characterization Data of Products.**

#### *Cyanosilylated carbonyl compounds catalysed by 2.12<sup>Y</sup>*

**2-Phenyl-2-((trimethylsilyl)oxy)acetonitrile (3a).** This product has been previously reported.[10] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 7.50–7.35 (m, 5H, ArH), 5.50 (s, 1H, *CH*CN), 0.23 (s, 9H, TMS) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 136.2 (C*ipso*), 129.3(ArCH), 128.9 (ArCH), 126.3 (ArCH), 119.1 (CN), 63.6 (CH), -0.29 (TMS) ppm.

**2-(***p***-Tolyl)-2-((trimethylsilyl)oxy)acetonitrile (3b).** This product has been previously reported.[11] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 7.37 (d, *J* = 7.9 Hz, 2H, ArH), 7.22 (d, *J* = 7.9 Hz, 2H, ArH), 5.47 (s, 1H, *CH*CN), 2.38 (s, 3H, CH3), 0.23 (s, 9H, TMS) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 139.3 (C*ipso*), 133.3 (C*ispo*), 129.5 (ArCH), 126.3 (ArCH), 119.2 (CN), 63.5 (CH), 21.1 (CH3), -0.29 (TMS) ppm.

**2-(4-Methoxyphenyl)-2-((trimethylsilyl)oxy)acetonitrile (3c).** This product has been previously reported.[12] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 7.38 (d, *J* = 8.6 Hz, 2H, ArH), 6.92 (d, *J* = 8.6 Hz, 2H, ArH), 5.43 (s, 1H, *CH*CN), 3.82 (s, 3H, OMe), -0.21 (s, 9H, TMS) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 160.3 (C*ipso*), 128.3 (C*ipso*), 127.9 (ArCH), 119.3 (CN), 114.2 (ArCH), 63.3 (CH), 55.3 (OCH3), -0.24 (TMS) ppm.

**2-(4-(Dimethylamino)phenyl)-2-((trimethylsilyl)oxy)acetonitrile (3d).** <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 7.31 (d, *J* = 8.7 Hz, 2H, ArH), 6.71 (d, *J* = 8.7 Hz, 2H, ArH), 5.39 (s, 1H, *CH*CN), 2.98 (s, 6H, N*Me2*), 0.19 (s, 9H, TMS) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 151.1 (C*ipso*), 127.8 (ArCH), 123.6 (C*ipso*), 119.6 (CN), 112.1 (ArCH), 63.7 (CH), 40.3 (CH3), -0.16 (TMS) ppm. IR (ATR): ν 2959 (CH3), 2230 (C≡N), 1621 (C=N), 1531 (C=C), 1371 (C-N), 1257 (C-O), 1061 (C-O), 835 cm<sup>-1</sup>. Elemental Analysis calc. for C<sub>13</sub>H<sub>20</sub>N<sub>2</sub>OSi: C 62.86, N, 11.28, H 8.12; found: C 63.23, N 11.65, H 7.67.

**2-(3-Fluorophenyl)-2-((trimethylsilyl)oxy)acetonitrile (3e).** This product has been previously reported.[13] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 7.45–7.35 (m, 1H, ArH), 7.25–7.15 (m, 2H, ArH), 7.10–7.05 (m, 1H, ArH), 5.50 (s, 1H, *CH*CN), 0.25 (s, 9H, TMS) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 162.8 (d, C*ipso*, <sup>1</sup>*J*C-F = 247.7 Hz,), 138.6 (d, C*ipso*, <sup>3</sup>*J*C-F = 7.8 Hz,), 130.5 (d, ArCH,  ${}^{3}$ *J*c-F = 8.1 Hz,), 121.7 (d, ArCH,  ${}^{4}$ *J*c-F = 3.0 Hz,), 118.7 (CN), 116.3 (d, ArCH,  ${}^{2}$ *J*c-F = 21.2 Hz,), 113.3 (d, ArCH, <sup>2</sup>*J*C-F = 23.4 Hz,), 62.8 (d, CH, <sup>4</sup>*J*C-F = 2.1 Hz,), -0.4 (TMS) ppm. <sup>19</sup>F-NMR (282.4 MHz, CDCl3): δ -111.4 ppm.

**2-(4-Chlorophenyl)-2-((trimethylsilyl)oxy)acetonitrile (3f)**. This product has been previously reported.[14] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 7.45–7.40 (m, 5H, ArH), 5.49 (s, 1H, *CH*CN), 0.26 (s, 9H, TMS) ppm. 13C NMR (75.48 MHz, CDCl3): δ 135.3 (C*ipso*), 134.8 (C*ipso*), 129.1 (ArCH), 127.7 (ArCH), 118.8 (CN), 63.0 (CH), -0.30 (TMS) ppm.

**2-(4-Nitrophenyl)-2-((trimethylsilyl)oxy)acetonitrile (3g)**. This product has been previously reported.[14] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 8.29 (d, *J* = 8.8 Hz, 2H, ArH), 7.67 (d, *J*  = 8.8 Hz, 2H, ArH), 5.59 (s, 1H, *CH*CN), 0.29 (s, 9H, TMS) ppm. 13C NMR (75.48 MHz, CDCl3): δ 148.4 (C*ipso*), 142.8 (C*ispo*), 127.0 (ArCH), 124.1 (ArCH), 118.1 (CN), 62.6 (CH), -0.40 (TMS) ppm.

**2-(Pyridin-2-yl)-2-((trimethylsilyl)oxy)acetonitrile (3h).** This product has been previously reported.[15] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 8.60–8.55 (m, 1H, ArH), 7.79 (dt, *J* = 7.7, 1.7 Hz, 1H, ArH), 7.59 (d, *J* = 7.7 Hz, 1H, ArH), 7.35–7.25 (m, 1H, ArH), 5.58 (s, 1H, *CH*CN), 0.26 (s, 9H, TMS) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 155.4 (C*ipso*), 149.3 (ArCH), 137.5(ArCH), 124.0 (ArCH), 120.5 (ArCH), 118.6 (CN), 65.1 (CH), -0.37 (TMS) ppm.

**2-(Quinolin-2-yl)-2-((trimethylsilyl)oxy)acetonitrile (3i).** <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 8.27 (d, *J* = 8.5 Hz, 1H, ArH), 8.09 (d, *J* = 8.5 Hz, 1H, ArH), 7.85 (d, *J* = 8.1 Hz, 1H, ArH), 7.80– 7.70 (m, 2H, ArH), 7.60–7.55 (m, 1H, ArH), 5.75 (s, 1H, *CH*CN), 0.27 (s, TMS) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 155.4 (C*ipso*), 147.2 (C*ipso*), 137.9 (ArCH), 130.1 (ArCH), 129.3 (ArCH), 128.0 (C*ipso*), 127.6 (ArCH), 127.3 (ArCH), 118.6 (CN), 117.8 (ArCH), 65.9 (CH), -0.31 (TMS) ppm. IR (ATR): ν 3058 (Csp<sup>2</sup> -H), 2959 (CH3), 2170 (C≡N), 1593 (C=N), 1504 (C=C), 1254 (C-O), 1046 (C-O), 838, 774 cm<sup>-1</sup>. Elemental Analysis calc. for C<sub>14</sub>H<sub>16</sub>N<sub>2</sub>OSi: C 65.59, N, 10.93, H 6.29; found: C 64.85, N 10.66, H 5.98.

**2-(Furan-2-yl)-2-((trimethylsilyl)oxy)acetonitrile (3j).** This product has been previously reported.[16] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 7.45 (d, *J* = 1.7 Hz, 1H, CH), 6.54 (d, *J* = 3.3 Hz, 1H, CH), 6.40 (dd, *J* = 3.3, 1.7 Hz, 1H, CH), 5.54 (s, 1H, *CH*CN), 0.19 (s, 9H, TMS) ppm. <sup>13</sup>CNMR (75.48 MHz, CDCl3): δ 148.2 (C), 143.8 (FurCH), 117.1 (CN), 110.8 (FurCH), 109.7 (FurCH), 57.4 (CH), -0.42 (TMS) ppm.

**2-(Thiophen-2-yl)-2-((trimethylsilyl)oxy)acetonitrile (3k).** This product has been previously reported.[16] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 7.37 (d, *J* = 5.3 Hz, 1H, CH), 7.20–7.15 (m, 1H, CH), 7.05–7.00 (m, 1H, CH), 5.73 (s, 1H, *CH*CN), 0.23 (s, 9H, TMS) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 139.5 (C), 127.2 (TiophCH), 126.9 (TiophCH), 126.3 (TiophCH), 118.3 (CN), 59.5 (CH), -0.31 (TMS) ppm.

**2-((Trimethylsilyl)oxy)butanenitrile (3l).** This product has been previously reported.[17] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 4.34 (t, J = 6.3 Hz, 1H, CH), 1.85–1.75 (m, 2H, CH2), 1.04 (t, J  $=$  7.4 Hz, 3H, CH<sub>3</sub>), 0.21 (s, 9H, CH<sub>3</sub> x 3) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl<sub>3</sub>): δ 119.9 (CN), 62.7 (CH), 29.6 (CH2), 8.9 (CH3), 0.4 (TMS) ppm.

**(***E***)-4-Phenyl-2-((trimethylsilyl)oxy)but-3-enenitrile (3m).** This product has been previously reported.[18] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 7.45–7.30 (m, 5H, ArH), 6.82 (d, *J* = 15.8 Hz, 1H, CH), 6.20 (dd, *J* = 15.8, 6.0 Hz, 1H, CH), 5.13 (d, *J* = 6.0 Hz, 1H, CH), 0.26 (s, 9H, TMS) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 135.0 (C*ipso*). 133.9 (CH), 128.76 (ArCH), 128.71 (ArCH), 126.96 (ArCH), 123.5 (CH), 118.4 (CN), 62.2 (CH), 0.15 (TMS) ppm.

**2-Phenyl-2-((trimethylsilyl)oxy)propanenitrile (3n).** This product has been previously reported.[19] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 7.60–7.50 (m, 2H, ArH), 7.45–7.30 (m, 3H, ArH), 1.86 (s, 3H, CH3), 0.18 (s, 9H, TMS) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 142.0 (C*ipso*), 128.68 (ArCH), 128.66 (ArCH), 124.6 (ArCH), 121.6 (CN), 71.6 (C), 33.5 (CH3), 1.03 (TMS) ppm.

**2-(4-Isobutylphenyl)-2-((trimethylsilyl)oxy)propanenitrile (3o).** This product has been previously reported.[20] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 7.44 (d, *J* = 8.2 Hz, 2H, ArH), 7.16 (d, *J* = 8.2 Hz, 2H, ArH), 2.49 (d, *J* = 7.2 Hz, 2H, CH2), 1.90–1.80 (m, 4H, CH, CH3), 0.90 (d, *J* = 6.6 Hz, 6H, 2 x CH3), 0.16 (s, 9H, TMS) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 142.3 (C*ipso*), 139.2 (C*ipso*), 129.3 (ArCH), 124.5 (ArCH), 121.8 (CN), 71.5 (C), 44.9 (CH2), 33.4 (CH3), 30.1 (CH3), 22.3 (CH3), 1.04 (TMS) ppm.

**2,2-Diphenyl-2-((trimethylsilyl)oxy)acetonitrile (3p).** This product has been previously reported.[21] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 7.55–7.50 (m, 4H, ArH), 7.40–7.35 (m, 6H, ArH), 0.15 (s, 9H, TMS) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 141.9 (C*ipso*), 128.6 (ArCH), 128.5 (ArCH), 125.9 (ArCH), 120.7 (CN), 76.3 (C), 0.9 (TMS) ppm.

**2-Methyl-2-((trimethylsilyl)oxy)butanenitrile (3q).** This product has been previously reported.[22] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 1.85–1.65 (m, 2H, CH2), 1.55 (s, 3H, CH3), 1.04 (t, *J* = 7.4 Hz, 3H, *CH3*CH2), 0.23 (s, 9H, TMS) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 121.9 (CN), 70.2 (C), 36.4 (CH<sub>2</sub>), 28.4 (CH<sub>3</sub>), 8.6 (CH<sub>3</sub>), 1.21 (TMS) ppm.

**1-((Trimethylsilyl)oxy)cyclohexane-1-carbonitrile (3r).** This product has been previously reported.[23] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 2.10–2.00 (m, 2H, CH2), 1.75–1.70 (m, 2H, CH<sub>2</sub>), 1.70–1.45 (m, 6H, CH<sub>2</sub> x 3), 1.30–1.20 (m, 2H, CH<sub>2</sub>), 0.23 (s, 9H, TMS) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 121.9 (CN), 70.6 (C), 39.3 (CH2), 24.5 (CH2), 22.6 (CH2), 1.38 (TMS) ppm.

**1-((Trimethylsilyl)oxy)-1,2,3,4-tetrahydronaphthalene-1-carbonitrile (3s).** This product has been previously reported.[23] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 7.70–7.60 (m, 1H, ArH), 7.30– 7.20 (m, 2H, ArH), 7.15–7.05 (m, 1H, ArH), 2.85–2.80 (m, 2H), 2.40–2.30 (m, 1H), 2.25–2.15 (m, 1H), 2.10–1.95 (m, 2H), 0.21 (s, 9H, TMS) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 136.1 (C*ipso*), 135.7 (C*ipso*), 129.3 (ArCH), 129.1 (ArCH), 128.0 (ArCH), 126.6 (ArCH), 122.1 (CN), 69.6 (C), 37.7  $(CH<sub>2</sub>), 28.3 (CH<sub>2</sub>), 18.7 (CH<sub>2</sub>), 1.3 (TMS)$  ppm.

**2-((Trimethylsilyl)oxy)adamantane-2-carbonitrile (3t).** This product has been previously reported.[24,25] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 2.20–2.00 (m, 6H), 1.95–1.80 (m, 3H), 1.80– 1.65 (m, 3H), 1.60–1.50 (m, 2H), 0.25 (s, 9H, TMS) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 122.1

(CN), 74.7 (C), 38.1 (CH), 37.2 (CH2), 34.7 (CH2), 30.9 (CH2), 26.3 (CH), 26.1 (CH), 1.2 (TMS) ppm.

**2,2-Dicyclohexyl-2-((trimethylsilyl)oxy)acetonitrile (3u).** <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 1.90–1.75 (m, 8H), 1.70–1.60 (m, 4H), 1.30–1.05 (m, 10H, CH2), 0.23 (s, 9H, TMS) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 120.5 (CN), 80.6 (C), 43.6 (CH), 28.3 (CH2), 26.4 (CH2), 26.2 (CH2), 26.1 (CH2), 1.9 (TMS) ppm. IR (ATR): ν 2931 (CH2), 2858 (CH2), 1451, 1249, 1123, 1023, 842, 766 cm-1 . Elemental Analysis calc. for C17H31NOSi: C 69.56, N 4.77, H 10.65; found: C 69.62, N 4.68, H 10.21.

#### *Cyanosilylated carbonyl compounds catalysed by 2.19Eu*

**2-Phenyl-2-((trimethylsilyl)oxy)acetonitrile (4a).** This product has been previously reported.[10] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 7.50–7.35 (m, 5H, ArH), 5.50 (s, 1H, *CH*CN), 0.23 (s, 9H, TMS) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 136.2 (C*ipso*), 129.3(ArCH), 128.9 (ArCH), 126.3 (ArCH), 119.1 (CN), 63.6 (CH), -0.29 (TMS) ppm.

**2-(4-Methoxyphenyl)-2-((trimethylsilyl)oxy)acetonitrile (4b).** This product has been previously reported.[12] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 7.38 (d, *J* = 8.6 Hz, 2H, ArH), 6.92 (d, *J* = 8.6 Hz, 2H, ArH), 5.43 (s, 1H, *CH*CN), 3.82 (s, 3H, OMe), -0.21 (s, 9H, TMS) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 160.3 (C*ipso*), 128.3 (C*ipso*), 127.9 (ArCH), 119.3 (CN), 114.2 (ArCH), 63.3 (CH), 55.3 (OCH3), -0.24 (TMS) ppm.

**2-(4-Chlorophenyl)-2-((trimethylsilyl)oxy)acetonitrile (4c).** This product has been previously reported.[14] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 7.45–7.40 (m, 5H, ArH), 5.49 (s, 1H, *CH*CN), 0.26 (s, 9H, TMS) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 135.3 (C*ipso*), 134.8 (C*ipso*), 129.1 (ArCH), 127.7 (ArCH), 118.8 (CN), 63.0 (CH), -0.30 (TMS) ppm.

**2-Phenyl-2-((trimethylsilyl)oxy)propanenitrile (4d).** This product has been previously reported.[19] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 7.60–7.50 (m, 2H, ArH), 7.45–7.30 (m, 3H, ArH), 1.86 (s, 3H, CH3), 0.18 (s, 9H, TMS) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 142.0 (C*ipso*), 128.68 (ArCH), 128.66 (ArCH), 124.6 (ArCH), 121.6 (CN), 71.6 (C), 33.5 (CH3), 1.03 (TMS) ppm.

**2-(4-Methoxyphenyl)-2-((trimethylsilyl)oxy)propanenitrile (4e).** This product has been previously reported.[22] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 7.46 (d, *J* = 8.7 Hz, 2H, ArH), 6.91 (d, *J*  $= 8.7$  Hz, 2H, ArH), 3.82 (s, 3H, OMe), 1.85 (s, 3H, CH<sub>3</sub>), 0.16 (s, 9H, TMS) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 159.7 (C*ipso*), 133.9 (C*ipso*), 126.0 (ArCH), 121.7 (CN), 113.8 (ArCH), 71.2 (C), 55.2 (OCH3), 33.3 (CH3), 1.00 (TMS) ppm.

**2-(4-Isobutylphenyl)-2-((trimethylsilyl)oxy)propanenitrile (4f).** This product has been previously reported.[20] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 7.44 (d, *J* = 8.2 Hz, 2H, ArH), 7.16 (d, *J* = 8.2 Hz, 2H, ArH), 2.49 (d, *J* = 7.2 Hz, 2H, CH2), 1.90–1.80 (m, 4H, CH, CH3), 0.90 (d, *J* = 6.6 Hz, 6H, 2 x CH3), 0.16 (s, 9H, TMS) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 142.3 (C*ipso*), 139.2 (C*ipso*), 129.3 (ArCH), 124.5 (ArCH), 121.8 (CN), 71.5 (C), 44.9 (CH2), 33.4 (CH3), 30.1 (CH3), 22.3 (CH3), 1.04 (TMS) ppm.

**2-(4-Chlorophenyl)-2-((trimethylsilyl)oxy)propanenitrile (4g).** This product has been previously reported.[22] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 7.48 (d, *J* = 8.5 Hz, 2H, ArH), 7.37 (d, *J*  $= 8.5$  Hz, 2H, ArH), 1.83 (s, 3H, CH<sub>3</sub>), 0.19 (s, 9H, TMS) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl<sub>3</sub>): δ 140.6 (C*ipso*), 134.5 (C*ipso*), 128.8 (ArCH), 126.0 (ArCH), 121.2 (CN), 71.0 (C), 33.4 (CH3), 1.00 (TMS) ppm.

**2-(3-Chlorophenyl)-2-((trimethylsilyl)oxy)propanenitrile (4h).** This product has been previously reported.[26] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 7.55 (s, 1H, ArH), 7.50–7.40 (m, 1H, ArH), 7.40–7.30 (m, 2H, ArH), 1.87 (s, 3H, CH3), 0.23 (s, 9H, TMS) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 144.1 (C*ipso*), 134.7 (C*ipso*), 130.0 (ArCH), 128.8 (ArCH), 124.9 (ArCH), 122.58 (ArCH), 121.1 (CN), 71.0 (C), 33.5 (CH3), 1.03 (TMS) ppm.

**2-(2-Chlorophenyl)-2-((trimethylsilyl)oxy)propanenitrile (4i).** This product has been previously reported.[27] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 7.75–7.70 (m, 1H, ArH), 7.45–7.40 (m, 1H, ArH), 7.35–7.30 (m, 2H, ArH), 2.00 (s, 3H, CH3), 0.29 (s, 9H, TMS) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 138.0 (C*ipso*), 131.5 (ArCH), 131.2 (C*ipso*), 129.9 (ArCH), 127.0 (ArCH), 126.9 (ArCH), 120.4 (CN), 70.2 (C), 29.7 (CH3), 1.14 (TMS) ppm.

**2-(2,4-Difluorophenyl)-2-((trimethylsilyl)oxy)propanenitrile** (**4j**). This product has been previously reported.[28] <sup>1</sup>H NMR (500.13 MHz, CDCl3): δ 7.56 (td, *J* = 8.8, 6.4 Hz, 1H, ArH), 6.95– 6.90 (m, 1H, ArH), 6.86 (ddd, *J* = 11.1, 8.8, 2.5 Hz, 1H, ArH), 1.92 (s, 3H, CH3), 0.27 (s, 9H, TMS) ppm. <sup>13</sup>C NMR (125.77 MHz, CDCl3): δ 163.2 (dd, *J* = 250.8, 12.0 Hz, C*ipso*-F), 159.4 (dd, *J* = 252.5, 12.0 Hz, C*ipso*-F), 127.8 (dd, *J* = 9.7, 4.3 Hz, ArCH), 125.0 (dd, *J* = 11.2, 3.9 Hz, C*ipso*-F), 120.4 (CN), 111.2 (d, *J* = 21.2 Hz, ArCH), 104.9 (t, *J* = 25.6 Hz, ArCH), 68.0 (d, *J* = 2.0 Hz, C), 30.8 (d, *J* = 2.9 Hz, CH3), 1.08 (TMS) ppm.

**2-(Pyridin-2-yl)-2-((trimethylsilyl)oxy)propanenitrile (4k)**. This product has been previously reported.[29] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 8.62 (d, *J* = 4.7 Hz, 1H, ArH), 7.77 (t, *J* = 7.8 Hz, 1H, ArH), 7.60 (d, J = 7.8 Hz, 1H, ArH), 7.30–7.25 (m, 1H, ArH), 1.93 (s, 3H, CH3), 0.26 (s, 9H, TMS) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 160.0 (C*ipso*), 149.0 (ArCH), 137.2 (ArCH), 123.4 (ArCH), 121.3 (CN), 118.9 (ArCH), 72.9 (C), 31.2 (CH3), 1.06 (TMS) ppm.

**3,3,3-Trifluoro-2-(4-fluorophenyl)-2-((trimethylsilyl)oxy)propanenitrile** (**4l**). <sup>1</sup>H NMR (500.13 MHz, CDCl3): δ 7.65 (dd, *J* = 8.0, 5.2 Hz, 2H ArH), 7.25–7.20 (m, 2H, ArH), 0.26 (s, 9H, TMS) ppm. <sup>13</sup>C NMR (125.77 MHz, CDCl3): δ 164.0 (d, *J* = 251.2Hz, C*ipso*-F), 129.0 (d, *J* = 9.0 Hz, *C*H-CH-C-F), 128.5 (d, *J* = 3.3 Hz, C*ipso*), 121.5 (q, *J* = 285.5 Hz, CF3), 115.9 (d, *J* = 22.2 Hz, *C*H-C-F), 115.6 (CN), 74.5 (q, *J* = 34.4 Hz, C-CF3), 0.60 (TMS) ppm. <sup>19</sup>F NMR (470.54 MHz, CDCl3): δ -80.2 (CF3), -109.8 (C*ipsoF*) ppm. IR (ATR): ν 2966 (CF3), 1606, 1509, 1415 (C=C), 1259 (C-O), 1185, 1144, 846, 817, 756 cm<sup>-1</sup>. Elemental Analysis calc. for C<sub>12</sub>H<sub>13</sub>F<sub>4</sub>NOSi: C 49.5, N 4.8, H 4.5, found: C 49.1, N 4.7, H 4.4.

**2,2-Diphenyl-2-((trimethylsilyl)oxy)acetonitrile (4m).** This product has been previously reported.[21] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 7.55–7.50 (m, 4H, ArH), 7.40–7.35 (m, 6H, ArH), 0.15 (s, 9H, TMS) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 141.9 (C*ipso*), 128.6 (ArCH), 128.5 (ArCH), 125.9 (ArCH), 120.7 (CN), 76.3 (C), 0.9 (TMS) ppm.

**2-Phenyl-2-(***o***-tolyl)-2-((trimethylsilyl)oxy)acetonitrile** (**4n**). <sup>1</sup>H NMR (500.13 MHz, CDCl3): δ 7.95–7.90 (m, 1H, ArH), 7.45–7.40 (m, 2H, ArH), 7.40–7.35 (m, 3H, ArH), 7.35–7.30 (m, 2H, ArH), 7.15–7.10 (m, 1H, ArH), 2.00 (s, 3H, CH3), 0.11 (s, 9H, TMS) ppm. <sup>13</sup>C NMR (125.77 MHz, CDCl3): δ 140.9 (C*ipso*), 138.3 (C*ipso*), 136.3 (C*ipso*), 132.5 (ArCH), 129.1 (ArCH), 128.7 (ArCH), 128.5 (ArCH), 126.9 (ArCH), 126.4 (ArCH), 125.7 (ArCH), 120.1 (CN), 76.4 (C), 20.5 (CH3), 0.84 (TMS) ppm. **IR (ATR):** ν 3063, 3028 (=C-H), 2959 (CH3), 1601, 1489 (C=C), 1266, 1253 (C-O), 1070, 1055, 1031, 872, 840 cm-1 . **Elemental Analysis calc. fo**r C18H21NOSi: C 73.2, N 4.7, H 7.2, found: C 69.35, N 4.56, H 6.48.

**(***E***)-2,4-Diphenyl-2-((trimethylsilyl)oxy)but-3-enenitrile** (4**o**). This product has been previously reported.[30] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 7.60 (d, *J* = 7.4 Hz, 2H, ArH), 7.45–7.25 (m, 8H, ArH), 7.03 (d, *J* = 15.9 Hz, 1H, CH), 6.21 (d, *J* = 15.9 Hz, 1H, CH), 0.27 (s, 9H, TMS) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 140.3 (C*ipso*), 135.1 (C*ipso*), 130.9, 128.9, 128.8, 128.71, 128.67, 127.0, 125.4, 119.6 (CN), 75.0 (C), 1.27 (TMS) ppm.

**2,2-Dicyclohexyl-2-((trimethylsilyl)oxy)acetonitrile (4p).** This product has been previously reported.[31] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 1.90–1.75 (m, 8H), 1.70–1.60 (m, 4H), 1.30–1.05 (m, 10H, CH2), 0.23 (s, 9H, TMS) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 120.5 (CN), 80.6 (C), 43.6 (CH), 28.3 (CH<sub>2</sub>), 26.4 (CH<sub>2</sub>), 26.2 (CH<sub>2</sub>), 26.1 (CH<sub>2</sub>), 1.9 (TMS) ppm. IR (ATR): ν 2931 (CH2), 2858 (CH2), 1451, 1249, 1123, 1023, 842, 766 cm-1 .

**2-Methyl-2-((trimethylsilyl)oxy)butanenitrile (4q).** This product has been previously reported.[22] <sup>1</sup>H NMR (300.13 MHz, CDCl<sub>3</sub>): δ 1.85–1.65 (m, 2H, CH<sub>2</sub>), 1.55 (s, 3H, CH<sub>3</sub>), 1.04 (t, *J* = 7.4 Hz, 3H, *CH3*CH2), 0.23 (s, 9H, TMS) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 121.9 (CN), 70.2 (C), 36.4 (CH2), 28.4 (CH3), 8.6 (CH3), 1.21 (TMS) ppm.

**1-((Trimethylsilyl)oxy)cyclohexane-1-carbonitrile (4r).** This product has been previously reported.[22] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 2.10–2.00 (m, 2H, CH2), 1.75–1.70 (m, 2H, CH<sub>2</sub>), 1.70–1.45 (m, 6H, CH<sub>2</sub> x 3), 1.30–1.20 (m, 2H, CH<sub>2</sub>), 0.23 (s, 9H, TMS) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl<sub>3</sub>): δ 121.9 (CN), 70.6 (C), 39.3 (CH<sub>2</sub>), 24.5 (CH<sub>2</sub>), 22.6 (CH<sub>2</sub>), 1.38 (TMS) ppm.

**1-((Trimethylsilyl)oxy)-1,2,3,4-tetrahydronaphthalene-1-carbonitrile (4s).** This product has been previously reported.[23] <sup>1</sup>H NMR (300.13 MHz, CDCl<sub>3</sub>): δ 7.70–7.60 (m, 1H, ArH), 7.30– 7.20 (m, 2H, ArH), 7.15–7.05 (m, 1H, ArH), 2.85–2.80 (m, 2H), 2.40–2.30 (m, 1H), 2.25–2.15 (m, 1H), 2.10–1.95 (m, 2H), 0.21 (s, 9H, TMS) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 136.1 (C*ipso*), 135.7 (C*ipso*), 129.3 (ArCH), 129.1 (ArCH), 128.0 (ArCH), 126.6 (ArCH), 122.1 (CN), 69.6 (C), 37.7 (CH2), 28.3 (CH2), 18.7 (CH2), 1.3 (TMS) ppm.

**2-((Trimethylsilyl)oxy)adamantane-2-carbonitrile (4t).** This product has been previously reported.[24,25] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 2.20–2.00 (m, 6H), 1.95–1.80 (m, 3H), 1.80– 1.65 (m, 3H), 1.60–1.50 (m, 2H), 0.25 (s, 9H, TMS) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 122.1 (CN), 74.7 (C), 38.1 (CH), 37.2 (CH2), 34.7 (CH2), 30.9 (CH2), 26.3 (CH), 26.1 (CH), 1.2 (TMS) ppm.



### **A2.15 Transformation into pellets and membranes**

**Figure A2.45.** Schematic representation of how pellets preparations have been performed with the home-made extrusion apparatus and how the coating of the pellets was done (up) and picture of an uncoated pellet of compound **2.6Tb** (down).



**Figure A2.46.** Schematic representation of how membrane preparations has been performed (up) and picture compound **2.6Tb** immobilized in polymethyl methacrylate (PMMA) and polysulphone (PSF) membranes from left to right, respectively (down).

#### **A2.15.1 Moisture stability**

PXRD results exhibit that compound 2.6<sub>Tb</sub> is stable and keeps its structure after being transformed into pellets or membranes and being exposed for 72 h at 98 % RH. Contrary to what happens when material is directly put in contact with water, that evolves into another crystalline phase, long exposure to humidity do not provoke material transformation and remains stable. Due to the amorphous nature of the polymer, only the main intense peaks of compound 2.6<sub>Tb</sub> stands over the amorphous background in polysulphone based membrane's PXRD.



**Figure A2.47.** Powder X-ray diffractograms of the studied materials, as-synthesized and after processing into pellets and membranes.

#### **A2.15.2 Temperature stability**

Once carrying out humidity tests, pellets were subsequently treated to temperature cycles. For that purpose, four heating and cooling down cycles were performed and materials were characterised by PXRD analysis. Therefore, uncoated pellets and polysulphone with coated were first tested against 98 % relative humidity (RH) for 72 h and later tested their thermal stability. For this purpose, temperature cycles of heating to 125 ºC and cooling down to room temperature were carried out. After each cycle, a photograph of both pellets (coated and uncoated) was taken in order to check their integrity. Subsequently, PXRD analysis was performed after the second and forth cycle in coated and uncoated pellets.



**Figure A2.48.** Powder X-ray diffractograms of the studied materials of compound **2.6Tb**, assynthesized and after processing into pellets.

# **Appendix 3**

Supporting information of Chapter 3

### **A3.1 Experimental section. General instrumentation**

**Elemental analyses** (C, H, N) were performed on a Leco CHNS-932 microanalyser. **Infrared** (FT-IR) spectra (400-4000 cm-1 ) were recorded on a Nicolet FT-IR 6700 spectrometer in KBr pellets.

**Thermogravimetric analysis** (TG/DTA) were performed on a TG-Q500 TA Instruments thermal analyser from room temperature to 800 °C under a synthetic air atmosphere (79 %  $N<sub>2</sub>/21$ % O<sub>2</sub>) at a heating rate of 10 °C min<sup>-1</sup>.

**X-ray data collection** of suitable single crystals were done at 100(2) K on a Bruker D8 VENTURE area detector equipped with graphite monochromated Mo−Kα radiation  $(\lambda = 0.71073 \text{ Å})$  by applying the  $\omega$ -scan method. The data reduction was performed with the APEX270 software and corrected for absorption using SADABS.[2] Crystal structures were solved by direct methods using the SIR97 program[3] and refined by full-matrix least-squares on F2 including all reflections using anisotropic displacement parameters by means of the WINGX[4] crystallographic package. Lattice solvent molecules could not be refined owing to their disordered disposition in the voids of the structures, so the electron density at the voids was subtracted from the reflection data by the SQUEEZE procedure as implemented in PLATON program[5] during the refinement.

**X-ray powder diffraction** (XRPD) patterns were collected at 25 °C on a Phillips X'PERT powder diffractometer with Cu-Kα radiation ( $λ = 1.5418$  Å) over the range  $5^{\circ} < 2θ < 50^{\circ}$  with a step size of 0.02<sup>o</sup> and an acquisition time of 2.5 s per step. Indexation of the diffraction profiles were made by means of the FULLPROF program (pattern- matching analysis) based on the space group and the cell parameters found by single crystal X-ray diffraction.

**Variable-temperature powder X-ray diffraction** measurements were conducted on a Bruker D8 Advance diffractometer, using polycrystalline sample of compound 2.7<sub>Dy</sub> under ambient atmosphere with heating rate of  $5^{\circ}$ C·min<sup>-1</sup> and measuring a complete diffractogram every 20 °C up to 510 °C, and every 50 °C from 510 °C up to 710 °C.

**Scanning electron microscopy** (SEM) images were acquired using either a Hitachi S4100 field emission gun tungsten filament instrument working at 25 kV or a high-resolution Hitachi SU-70 working at 4 kV. Samples were prepared by deposition on aluminium sample holders followed by carbon coating using an Emitech K950X carbon evaporator. EDS (energy dispersive X-ray spectroscopy) data and SEM mapping images were recorded using the latter microscope working at 15 kV and using either a Bruker Quantax 400 or an Esprit 1.9 EDS microanalysis system.

The **water vapour adsorption** isotherms were collected in a Dynamic Vapor Sorption apparatus from Surface Measurements Systems, using  $N_2$  as the carrier gas (Air Liquide Alphagaz, less than 3 ppm H2O, total flow of 200 sccm). Dry aliquots (16 mg) were loaded in a steel pan and suspended in the measuring chamber. The experiment started with a 2 h pretreatment at 125 °C, to completely dry the sample, followed by the isotherm at 25 °C with increasing and decreasing RH steps from 0 to 98 %. Each humidity step was kept until the rate of change of mass per fixed time (dm/dt) was lower than 0.002 %, for a period of at least 10 min.

### **A3.2 Chemical characterization**

#### **A3.2.1 Elemental Analysis**

Compound	Formula	<b>Molecular</b> weiaht	Calc.	Found.
3.1	$C_{20}H_{26}N_{2}O_{13}Cu_{3}$	692.5	C: 34.66; H: 3.78; N: C: 34.75; H: 3.74; N: 4.04; O: 30.01; Cu: 4.03; O: 30.00; Cu: 27.51	27.62

**Table A3.1.** Elemental analysis of compound **3.1**

#### **A3.2.2 FT-IR spectroscopy**

When FTIR spectra of is examined from left to right, from higher to smaller frequency, compound **3.1** exhibit an intense broad band around 3447 cm-1 assigned to O–H bond vibration of the of 3,4-dihydroxybenzoate free ligand. At lower frequency, a set of sharp and of medium intensity bands that corresponds to aromatic ring's C–H bond vibrations of the ligand are visible between 3073 cm-1 and 2989–2864 cm-1 . The following intense vibrations located in 1659–1426 cm-1 region are attributed to both the asymmetric stretching vibrations of the carboxylate groups and the aromatic C–C bonds. Moving to lower range of 1389–1252 cm-1 , symmetric stretching vibrations of the carboxylate groups appear in the spectrum. The remaining bands that are found at lower frequency can be attributed to the distortions originated in the aromatic ring and the carboxylate groups of the ligands.



**Figure A3.1.** Infrared spectra of the ligand and compound **3.1.**

# **A3.3 Crystallographic data**



**Table A3.2.** Crystallographic data and structure refinement details of compound **3.1**.

<b>Compound 3.1</b>					
<b>Atom</b>	Atom	Length/Å			
Cu1	Cu11	3.0121(8)			
Cu1	Cu <sup>2</sup>	3.0461(4)			
Cu1	O8	2.296(3)			
Cu1	O1	1.946(2)			
Cu1	O21	1.952(2)			
Cu1	O2	1.943(2)			
Cu1	O43	1.924(3)			
Cu <sub>2</sub>	O1 <sup>4</sup>	1.960(2)			
Cu2	O1 <sup>5</sup>	1.960(2)			
Cu2	O36	1.926(3)			
Cu2	O3	1.926(3)			

**Table A3.3.** Table of the selected bond lengths (Å) and angles (°) for compound **3.1.**

<sup>1</sup>1-x,+y,1/2-z; <sup>2</sup>3/2-x,-1/2+y,1/2-z; <sup>3</sup> -1/2+x,3/2-y,-1/2+z; <sup>4</sup>1/2+x,3/2-y,1/2+z; <sup>5</sup>3/2-x,1/2+y,1/2-z; <sup>6</sup>2-x,2-y,1-z

Atom 1	Atom 2	Atom 3	Angle 2,1,3 [∘]
Cu1 <sup>1</sup>	Cu1	Cu <sup>22</sup>	141.692(17)
Cu1	O2	Cu1 <sup>1</sup>	101.28(11)
O1	Cu1	O8	89.55(12)
O1	Cu1	O2 <sup>1</sup>	161.21(10)
O2 <sup>1</sup>	Cu1	Cu1 <sup>1</sup>	39.25(7)
O21	Cu1	O8	100.21(12)
O <sub>2</sub>	Cu1	O <sub>8</sub>	96.04(12)
O <sub>2</sub>	Cu1	O <sub>1</sub>	84.47(10)
O <sub>2</sub>	Cu1	O2 <sup>1</sup>	78.60(11)
O4 <sup>3</sup>	Cu1	Cu1 <sup>1</sup>	136.74(8)
O4 <sup>3</sup>	Cu1	Cu <sup>22</sup>	80.35(8)
O4 <sup>3</sup>	Cu1	O <sub>8</sub>	94.65(13)
O4 <sup>3</sup>	Cu1	O <sub>1</sub>	95.96(11)
O4 <sup>3</sup>	Cu1	O <sub>2</sub>	169.30(12)
$O4^3$	Cu1	O2 <sup>1</sup>	99.18(10)
Cu1 <sup>4</sup>	Cu2	Cu1 <sup>5</sup>	180.0
O15	Cu2	O1 <sup>4</sup>	180.00(13)
O3	Cu2	O1 <sup>4</sup>	91.30(11)
O <sub>3</sub>	Cu <sub>2</sub>	O15	88.69(11)
O36	Cu2	O1 <sup>5</sup>	91.30(11)
O36	Cu <sub>2</sub>	O1 <sup>4</sup>	88.70(11)
O36	Cu <sub>2</sub>	O <sub>3</sub>	180.0

<sup>1</sup>1-x,+y,1/2-z; <sup>2</sup>3/2-x,-1/2+y,1/2-z; <sup>3</sup> -1/2+x,3/2-y,-1/2+z; <sup>4</sup>3/2-x,1/2+y,1/2-z; <sup>5</sup>1/2+x,3/2-y,1/2+z; <sup>6</sup>2-x,2-y,1-z



### **A3.4 Powder X-ray diffraction analysis**

**Figure A3.2.** Figure of the pattern matching analysis and experimental PXRD for complex **3.1**.

Compound **3.1** stability was examined in various solvents. For that purpose, 100 mg of MOF were suspended in a 4 mL of solvent and left agitating for 16 h. Afterwards, material was filtered off and left drying before carrying on powder X-ray diffraction (PXRD) analysis.

According to the PXRD, compound **3.1** is stable in EtOH after 16 h keeping its structure, contrary to what it happens in MeOH and H2O. In the latter case, compound **3.1** completely evolves into a different crystalline product. However, in the former case, in MeOH, PXRD reveals that even if compound **3.1** most important peaks are present in the diffractogram, several new peaks appear. The position of those new peaks come in accordance with the evolved crystalline product obtained after soaking compound **3.1** for 16 h in water.



**Figure A3.3.**Experimental PXRD for complexes **3.1** after being soaked for 16 h in several solvents.

# **A3.5 Continuous Shape Measurements**

**Table A3.4.** Table of the continuous Shape Measurements for the CuO<sub>5</sub> coordination environment.





Table A3.5. Table of the continuous Shape Measurements for the CuO<sub>6</sub> coordination environment.





### **A3.6 Thermal analysis**

Thermogravimetric analyses performed over polycrystalline sample in compound **3.1** enabled checking the stability of the product. TG curves has been collected for compound **3.1** for the as synthesised compound **3.1** and after solvent exchange with EtOH. In stability section, we saw that material remains stable after being soaked in EtOH and being this latter solvent more volatile than the solvents used in the synthesis of the material, we tried to accomplish solvent exchange procedure as an approach to replace solvent molecules (dimethylformamide and water molecules) to ease material activation to posteriorly analyse its adsorptive-capacity. For this purpose, as for stability tests, compound **3.1** was suspended in EtOH for 16 h, left solvent evaporated and carried on thermal analysis. Powder X-ray diffraction confirmed that**3. 1** remains stable after solvent exchange with MeOH as it can be seen in Figure A3.5, right.

The thermal behaviour of the as synthesised [Cu3L2(DMF)2]·3H2O, compound **3.1**, exhibits two main regions. The first weight of loss was progressive and went from ambient temperature to 200 °C and corresponded to the loss of solvent molecules, firstly, lattice water molecules and then coordinated DMF molecules. Above this temperature, there is an abrupt loss corresponding to the complete collapse of the crystal structure. From the shape of the TG curve, it seems that solvent molecules stabilise the structure and their removal promote crystal structure decomposition. In the final step, as a consequence of the decomposition of the organic content metal oxide is obtained.



**Figure A3.4.** Figure of TG/DTG analysis of as synthesised compound **3.1**.

TG curve performed after solvent exchange with EtOH exhibits different pattern in comparison to the previous curve obtained with as synthesised compound **3.1** material. In this case, after solvent exchange procedure, the weight of loss occurs in three main regions. The first region comprises from the room temperature to 150 ºC and after an initial loss of weight occurring from 30 to 70 ºC a plateau is obtained around 80 ºC which goes up to 150 ºC. Then, from 150 ºC to 200 ºC a second loss of weight is visible in the spectra before the last abrupt loss corresponding to compound **3.1** degradation which occurs at 250 ºC. Above this temperature, CuO is obtained as final residue. The plateau obtained in the at around 80 ºC and the increase of final-residue percentage (in around 8 %) can be indicative of solvent molecules being fully or at least partly replaced by EtOH, being this solvent-exchange strategy a possible option to promote material activation at lower temperatures.



**Figure A3.5**. Figure of TG analysis of compound **3.1** performed after solvent exchange with EtOH during 16 h (up) and experimental PXRD for complex **3.1** before and after solvent exchange with EtOH (bottom).

## **A3.7 Thermal evolution**

Thermal evolution of as synthesised compound **3.1** shows that the material keeps its initial phase crystallinity up to 110 ºC. Above this temperature, new peaks appear in the diffractogram indicating the transformation into a different crystalline phase; this second phase, maintains its crystallinity up to 210  $\circ$ C and it must be noted that is very similar to the phase obtained when material was soaked in H2O. Finally, compound **3.1** collapses and evolves into metallic residue CuO, obtained around 800 ºC.



**Figure A3.6.** Thermal evolution of as synthesised compound **3.1.**



**Figure A3.7.** Similarity between the PXRD transformed phase of compound **3.1** obtained from thermal evolution at 190 ºC and the phase obtained soaking compound **3.1** in H2O during 16 h.



# **A3.8 Additional views of the structure**

**Figure A3.8.** View along *a*, *b* and *c* crystallographic axis, the corresponding topological representation and view of "ABBA..." SBU.



# **A3.9 Scanning Electron Microscopy**

**Figure A3.9.** SEM images of compound **3.1**.

# **Appendix 4**

Supporting information of Chapter 4

### **A4.1 Experimental section. General instrumentation**

**Elemental analyses** (C, H, N) were performed on a Leco CHNS-932 microanalyser. **Infrared** (FT-IR) spectra (400-4000 cm-1 ) were recorded on a Nicolet FT-IR 6700 spectrometer in KBr pellets.

**Thermogravimetric analysis** (TG/DTA) were performed on a TG-Q500 TA Instruments thermal analyser from room temperature to 800 °C under a synthetic air atmosphere (79 %  $N<sub>2</sub>/21$ % O<sub>2</sub>) at a heating rate of 10 °C min<sup>-1</sup>.

**X-ray powder diffraction** (XRPD) patterns were collected at 25 °C on a Phillips X'PERT powder diffractometer with Cu-Kα radiation ( $\lambda = 1.5418$  Å) over the range  $5^{\circ} < 2\theta < 50^{\circ}$  with a step size of 0.02° and an acquisition time of 2.5 s per step. Indexation of the diffraction profiles were made by means of the FULLPROF program (pattern- matching analysis) based on the space group and the cell parameters found by single crystal X-ray diffraction.

**Scanning electron microscopy (SEM**) images were acquired using either a Hitachi S4100 field emission gun tungsten filament instrument working at 25 kV or a high-resolution Hitachi SU-70 working at 4 kV. Samples were prepared by deposition on aluminium sample holders followed by carbon coating using an Emitech K950X carbon evaporator. EDS (energy dispersive X-ray spectroscopy) data and SEM mapping images were recorded using the latter microscope working at 15 kV and using either a Bruker Quantax 400 or an Esprit 1.9 EDS microanalysis system.

**GC-FID** analysis carried out in a Bruker 430-GC-FID chromatograph (Bruker, Faculty of Science, University of Porto, Porto, Portugal). Hydrogen was used as a carrier gas (55 cm·s−1), and fused silica Supelco capillary columns SPB-5 (30 m  $\times$  0.25 mm i.d.; 25 µm film thickness) were used.

**Photoluminescence Spectroscopy**. The emission and excitation spectra were recorded at ambient-temperature and 12 K using a Fluorolog®-3Horiba Scientific (Model FL3-2T) spectroscope, with a modular double grating excitation spectrometer (fitted with a 1200 grooves/mm grating blazed at 330 nm) and a TRIAX 320 single emission monochromator (fitted with a 1200 grooves/mm grating blazed at 500 nm, reciprocal linear density of 2.6 nm.mm<sup>-1</sup>), coupled to a R928 Hamamatsu photomultiplier, using the front face acquisition mode. The excitation source was a 450 W Xe arc lamp. The emission spectra were corrected for detection and optical spectral response of the spectrofluorometer and the excitation spectra were corrected for the spectral distribution of the lamp intensity using a photodiode reference detector. Timeresolved measurements have been carried out using a 1934D3 phosphorimeter coupled to the Fluorolog®-3, and a Xe-Hg flash lamp (6 μs/pulse half width and 20-30 μs tail) was used as the

excitation source. The low temperature measurements (12 K) were performed using a heliumclosed cycle cryostat with vacuum system measuring ca. 5x10<sup>-6</sup> mbar and a Lakeshore 330 autotuning temperature controller with a resistance heater.

# **A4.2 Chemical characterization**

## **A4.2.1 Elemental Analysis**

**Table A4.1** Elemental analysis of compound **4.1**-**4.8**.



### **A4.2.2 Determination of the metal content by ICP-AES**

**Table A4.2.** Determination of the metal content by ICP and Dy to Ru relationship.







**Figure A4.1.** Infrared spectra of the 5,5'-dimehtyl-2,2'-bipyridine, 2,2'-bipyridine-5,5' dicarboxylic acid, [RuL3]Cl<sup>2</sup> metalloligand and compounds **4.4Ru-Gd** and **4.6Ru-Dy**.

### **A4.2.4 <sup>1</sup>H-Nuclear Magnetic Resonance**

2,2'-bipyridine-5,5'-dicarboxylic acid was prepared following the literature method consisting in the oxidation of 5,5'-dimethyl-2,2'-bipyridyl with  $K_2Cr_2O_7$  in concentrated sulfuric acid. This product has been previously reported in bibliography.[32,33]



**Figure A4.2.**  <sup>1</sup>H-NMR spectra of 2,2'-bipyridine-5,5'-dibarboxylic acid in DMSO-*d6.*

This product has already been reported in bibliography and <sup>1</sup>H-NMR matches with the reported data.[34,35]



Figure A4.3. <sup>1</sup>H-NMR spectra of ruthenium(II) metalloligand in D<sub>2</sub>O made alkaline with NaOD.

# **A4.3 Crystallographic data**



**Table A4.3.** Crystallographic data and structure refinement details of compound **4.6Ru-Dy**.
Ru-metalloligand				Compound 4.6 <sub>Ru-Dy</sub>			
<b>Atom</b>	<b>Atom</b>	Length/Å	<b>Atom</b>	Atom	Length/A		
Ru1	N1 <sup>1</sup>	2.067(2)	Dy1	O <sub>1</sub>	2.34(3)		
Ru1	N <sub>1</sub>	2.067(2)	Dy1	O1 <sup>1</sup>	2.34(3)		
Ru1	N <sub>2</sub>	2.077(2)	Dy1	O <sup>22</sup>	2.31(4)		
Ru1	N2 <sup>1</sup>	2.077(2)	Dy1	O2 <sup>3</sup>	2.31(4)		
Ru1	N3 <sup>1</sup>	2.060(2)	Dy1	O4 <sup>4</sup>	2.23(4)		
Ru1	N1 <sup>1</sup>	2.060(2)	Dy1	O4 <sup>5</sup>	2.23(4)		
	$1-x, +y, 1/2-z$		Dy <sub>2</sub>	$Dy2^7$	3.879(17)		
			Dy <sub>2</sub>	O <sub>7</sub>	2.41(5)		
			Dy <sub>2</sub>	O5 <sup>1</sup>	2.28(6)		
			Dy <sub>2</sub>	O <sub>5</sub>	2.28(6)		
			Dy <sub>2</sub>	O6 <sup>8</sup>	2.34(6)		
			Dy <sub>2</sub>	O6 <sup>7</sup>	2.34(6)		
			Ru1	N <sub>2</sub>	2.07(3)		
			Ru1	N2 <sup>6</sup>	2.07(3)		
			Ru1	N <sub>1</sub>	2.07(3)		
			Ru1	N1 <sup>6</sup>	2.07(3)		
			Ru1	N <sub>3</sub>	2.09(4)		
			Ru1	N3 <sup>6</sup>	2.09(4)		
				1/2+x,1/2+y,+z; <sup>5</sup> 1/2- $y, 1-z$ ; $8+x, 1-y, 1-z$	$1-x, +y, +z$ ; $2+x, 1-y, -z$ ; $3-x, 1-y, -z$ ; $4-x, 1-z$ $x, 1/2+y, +z$ ; $61/2-x, +y, 1/2-z$ ; $7-x, 1-z$		

**Table A4.4.** Table of the selected bond lengths (Å) for Ru-metalloligand and compound **4.6Ru-Dy**.

Ru-metalloligand						$4.6Ru-Dy$					
Atom	Atom	Atom	Angle/°	Atom	Atom	Atom	Angle/°	Atom	Atom	Atom	Angle/°
N1 <sup>1</sup>	Ru1	N <sub>1</sub>	173.68(11)	N <sub>1</sub>	Ru1	N3 <sup>6</sup>	92.2(13)	O4 <sup>4</sup>	Dy1	$O2^2$	88.2(14)
N <sub>1</sub>	Ru1	N <sub>2</sub>	78.30(8)	N <sub>1</sub>	Ru1	N3	94.9(13)	O4 <sup>4</sup>	Dy1	O2 <sup>3</sup>	149.8(14)
N1 <sup>1</sup>	Ru1	N <sub>2</sub>	96.76(8)	N1 <sup>6</sup>	Ru1	N <sub>1</sub>	171.1(17)	O4 <sup>5</sup>	Dy1	O <sub>1</sub>	78.0(13)
N1 <sup>1</sup>	Ru1	N2 <sup>1</sup>	78.30(8)	N1 <sup>6</sup>	Ru1	N3	92.2(13)	O4 <sup>5</sup>	Dy1	O1 <sup>1</sup>	131.5(13)
N <sub>1</sub>	Ru1	N2 <sup>1</sup>	96.76(8)	N1 <sup>6</sup>	Ru1	N3 <sup>6</sup>	94.9(13)	O4 <sup>5</sup>	Dy1	O2 <sup>2</sup>	149.8(14)
N2 <sup>1</sup>	Ru1	N <sub>2</sub>	79.12(11)	N <sub>2</sub>	Ru1	N <sub>1</sub>	79.7(12)	O4 <sup>5</sup>	Dy1	O2 <sup>3</sup>	88.2(14)
N3	Ru1	N1 <sup>1</sup>	94.04(8)	N <sub>2</sub>	Ru1	N1 <sup>6</sup>	93.6(12)	O4 <sup>5</sup>	Dy1	O4 <sup>4</sup>	87(2)
N3	Ru1	N <sub>1</sub>	90.85(8)	N <sub>2</sub>	Ru1	N3	173.6(13)	O <sub>5</sub>	Dy <sub>2</sub>	O <sub>7</sub>	134.5(15)
N3 <sup>1</sup>	Ru1	N <sub>1</sub>	94.04(8)	N <sub>2</sub>	Ru1	N3 <sup>6</sup>	99.9(13)	O <sub>5</sub>	Dy <sub>2</sub>	O6 <sup>8</sup>	81(2)
N3 <sup>1</sup>	Ru1	N1 <sup>1</sup>	90.84(8)	N2 <sup>6</sup>	Ru1	N <sub>2</sub>	83.9(15)	O <sub>5</sub>	Dy <sub>2</sub>	O6 <sup>7</sup>	149(2)
N3	Ru1	N <sub>2</sub>	101.58(9)	N2 <sup>6</sup>	Ru1	N1 <sup>6</sup>	79.7(12)	O5 <sup>1</sup>	Dy <sub>2</sub>	O <sub>7</sub>	134.5(15)
N3 <sup>1</sup>	Ru1	N2 <sup>1</sup>	101.58(9)	N2 <sup>6</sup>	Ru1	N <sub>1</sub>	93.6(12)	O5 <sup>1</sup>	Dy <sub>2</sub>	O <sub>5</sub>	91(3)
N3	Ru1	N2 <sup>1</sup>	172.33(8)	N2 <sup>6</sup>	Ru1	N3	99.9(13)	O5 <sup>1</sup>	Dy <sub>2</sub>	O6 <sup>7</sup>	81(2)
N3 <sup>1</sup>	Ru1	N <sub>2</sub>	172.33(8)	N2 <sup>6</sup>	Ru1	N3 <sup>6</sup>	173.6(13)	O5 <sup>1</sup>	Dy <sub>2</sub>	O6 <sup>8</sup>	149(2)
N3	Ru1	N3 <sup>1</sup>	78.76(12)	N3	Ru1	N3 <sup>6</sup>	77(2)	O6 <sup>7</sup>	Dy <sub>2</sub>	O <sub>7</sub>	60.5(17)
		$1-x, +y, 1/2-z$		O <sub>1</sub>	Dy1	O1 <sup>1</sup>	78.8(14)	O6 <sup>8</sup>	Dy <sub>2</sub>	O <sub>7</sub>	60.5(17)
				O2 <sup>2</sup>	Dy1	O <sub>1</sub>	125.9(12)	O6 <sup>8</sup>	Dy <sub>2</sub>	O6 <sup>7</sup>	90(3)
				O2 <sup>2</sup>	Dy1	O1 <sup>1</sup>	76.0(12)	N1 <sup>6</sup>	Ru1	N <sub>1</sub>	171.1(17)
				O2 <sup>2</sup>	Dy1	O2 <sup>3</sup>	81.3(18)	N1 <sup>6</sup>	Ru1	N3	92.2(13)
				O2 <sup>3</sup>	Dy1	O1 <sup>1</sup>	125.9(12)	N1 <sup>6</sup>	Ru1	N3 <sup>6</sup>	94.9(13)
				O2 <sup>3</sup>	Dy1	O <sub>1</sub>	76.0(12)				
				O4 <sup>4</sup>	Dy1	O <sub>1</sub>	131.5(13)				
				O4 <sup>4</sup>	Dy1	O1 <sup>1</sup>	78.0(13)				
							1-x,+y,+z; 2-x,1-y,-z; 3+x,1-y,-z; 4-1/2+x,1/2+y,+z; 51/2-x,1/2+y,+z; 61/2-x,+y,1/2-z; 7- x, 1-y, 1-z; 8+x, 1-y, 1-z; 91/2+x, -1/2+y, +z				

Table A4.5. Table of the selected angles (°) for ruthenium metalloligand and compound 4.6<sub>Ru-Dy</sub>.

### **A4.4 Diffuse reflectance**



Figure A4.4. Normalized diffuse reflectance spectrum of  $RuL_3]Cl_2$  and compound 4.3<sub>Ru-Eu</sub>.



### **A4.5 Powder X-ray diffraction analysis**

**Figure A4.5.** Pattern matching analysis and experimental PXRD for complexes **4.1**-**4.8**.

### **A4.6 Continuous Shape Measurements**

CShMs for the coordination environment of **4.6Ru-Dy**. The lowest SHAPE values for each ion is shown highlighted in grey, indicating best fits.

Table A4.6. Table of the continuous Shape Measurements for the RuN<sub>6</sub> andDyO<sub>6</sub> coordination environments.

$HP-6$	$D_{6h}$	Hexagon	
PPY-6	$C_{5v}$	Pentagonal pyramid	
OC-6		Octahedron	
TPR-6	$D_{3h}$	Trigonal prism	
		JPPY-6 $C_{5v}$ Johnson pentagonal pyramid J2	

Complex $4.6_{Ru-Dy}$	$HP-6$	PPY-6	$OC-6$	TPR-6	JPPY-6
Dy1	30.003	15.450	17.049	0.379	19.024
Ru1	26.975	27.199	1.032	14.844	30.469

Table A4.7. Table of the continuous Shape Measurements for the DyO<sub>5</sub> coordination environment.





#### **A4.7 Thermal analysis**

Thermogravimetric analyses have been performed over polycrystalline sample in compound **4.4** in order to check the stability of the products. The TG curves show three main steps of weight loses. The first steps concern to the release of coordinated solvent molecules which are released from room temperature up to 200 °C. The second step refers to the decomposition of the ligands which involves the collapse of the crystal structure, evolving to mixture of Gd<sub>2</sub>O<sub>3</sub> and RuO obtained at 700 °C as the final residue.



**Figure A4.6.** Figure of TG/DTG analysis of compound **4.4Ru-Gd**.

### **A4.8 Thermal evolution**

Thermal evolution has been measured to Ru-Dy\_MOF and has showed that the material keeps its crystallinity up to around 200 ºC. At higher temperatures it stars degradation process until  $Dy_2O_3$  is get as final residue.



**Figure A4.7.** Thermal evolution of as synthesised compound **4.4Ru-Gd** (up) and **4.6Ru-Dy** (bottom).



#### **A4.9 Additional views of the structure**

**Figure A4.8.** View of Ru-metalloligand single crystal (up) and compound **4.6Ru-Dy** along *a*, *b* and *c* crystallographic axis (middle) and "ABAB..." growing of SBU showing hydrogen bonding interaction (bottom)**.**



# **A4.10 Scanning Electron Microscopy**



Spectrum: EE756

Element	Series		unn. C norm. C Atom. C Error (3 Sigma)			
		[wt.%]	[wt.%] [ <b>at.</b> %]			[wt.%]
Europium L-series		19.08	21.06	2.61		2.18
Ruthenium L-series		8.82	9.74	1.82		1.10
Chlorine	K-series	2.99	3.30	1.76		0.46
Potassium K-series		0.01	0.01	0.00		0.09
Carbon	K-series	37.53	41.41	64.98		16.30
Oxvqen	K-series	22.18	24.48	28.83		11.04
	Total:	90.61	100.00	100.00		

**Figure A4.9.** SEM images of compound **4.3Ru-Eu**.

## **A4.11 Extraction and Catalytic Oxidative Desulfurization**

**Table A4.8.** Desulfurization of a multicomponent model diesel catalysed by heterogeneous **4.3Ru-Eu** catalyst at three catalytic loadings.



### **A4.12 Cellular cytotoxicity**

Additionally, with this family of compounds cytotoxicity evaluation was performed. For that purpose, Adenocarcinoma cell line (Caco-2) was selected and cell viability assays conducted. This cell line is widely used as model of the intestinal epithelial barrier and is originally derived from a colon carcinoma.[36]

Compounds **4.4** and **4.6** in 1 to 0.8 mg·mL-1 MOF concentration were incubated for 72 h with the aforementioned cells and regardless the concentration cell viability do not decreases what confirms the lack of cytotoxicity of the synthesised materials.





Subsequently, analogous experiments were conducted with metalloligand and gadolinium and dysprosium metallic salts exhibiting similar behaviour. Results exhibited no toxicity observed in any precursor after 72 h regardless the concentration.



**Figure A4.11.** The cell viability of Caco-2 cells after being incubated with compounds **4.4** and **4.6** and precursors (metalloligand and metallic salts) for 72 h at various MOF concentrations.

# **Appendix 5**

Supporting information of Chapter 5

### **A5.1 Experimental section. General instrumentation**

**Elemental analyses** (C, H, N) were performed on a Leco CHNS-932 microanalyser.

**IR** spectra of the ligand and prepared coordination compound were collected in the region 400-4000 cm−1 on a Nicolet 6700 FTIR (Fourier transform infrared) spectrophotometer (Thermo Phisher Scientific, TX, USA) KBr pellets.

**Thermogravimetric analysis** (TG/DTA) were performed on a TG-Q500 TA Instruments thermal analyser from room temperature to 800 °C under a synthetic air atmosphere (79 % N<sub>2</sub>/21 % O<sub>2</sub>) at a heating rate of 10 °C min-1 .

**X-ray powder diffraction** (XRPD) patterns were collected at 25 °C on a Phillips X'PERT powder diffractometer with Cu-Kα radiation ( $\lambda = 1.5418$  Å) over the range  $5^{\circ} < 2\theta < 50^{\circ}$  with a step size of 0.02° and an acquisition time of 2.5 s per step. Indexation of the diffraction profiles were made by means of the FULLPROF program (pattern- matching analysis) based on the space group and the cell parameters found by single crystal X-ray diffraction.

**Magnetic susceptibility measurements** were performed on polycrystalline samples of the complexes with a Quantum Design SQUID MPMS-7T susceptometer at an applied magnetic field of 1000 G. The susceptibility data were corrected for diamagnetism estimated from Pascal's tables.[6] the temperature-independent paramagnetism and magnetisation of the sample holder. The ac measurements were performed on a physical property measurement system quantum design model 6000 magnetometer under a 3.5 G ac field and frequencies ranging from 60 to 10000 Hz.

**Photoluminescence measurements** at low temperature were done in an Edinburgh Instruments FLS920 spectrometer using a close cycle helium cryostat enclosed, in an applied vacuum (10–7 bar). For steady state measurements in the UV-Vis range an IK3552R-G HeCd continuous laser (325 nm) was used as excitation source, whereas a Müller‐Elektronik‐Optik SVX1450 Xe lamp was employed to collect the excitation spectra. LDH-P-C-370 laser diode of PicoQuant was employed for recording the decay curves corresponding to the lifetimes of ns range.

**MRI** scans were carried out in a preclinical 7-T magnet (Agilent, Palo Alto, CA, USA) interfaced to Avance III electronics, using a quadrature transmit-receive coil (Bruker, Ettlingen, Germany). *T<sup>1</sup>* values were estimated from images acquired using the rapid acquisition with relaxation enhancement (RARE) sequence with inversion recovery (IT = 50, 200, 400, 800, 1500, 3000, 5500, 8000, 12,000 ms, TE = 7.0 ms, echo train length 2, data matrix size 128  $\times$  64, field of view 30  $\times$  15 mm<sup>2</sup>, slice thickness = 3 mm, 1 scan).

## **A5.2 Chemical characterization**

### **A5.2.1 Elemental Analysis**



**Table A5.1** Elemental analysis of compound **5.1**-**5.14**.

#### **A5.2.2 FT-IR spectroscopy**

Derived by their isostructural nature, compounds **5.1**-**5.11** display very similar FT-IR spectra, that is **5.5<sub>Gd</sub>** was selected as representative material and compare it with the rest gadolinium-based counterparts, **5.12** and **5.13** materials. When examining FT-IR spectrum in detail from higher to smaller frequency, compound **5.11-5.13** display a narrow peak (not clearly visible in all compounds spectra) at around 3653 cm-1 attributed to the N-H stretching vibration of amine group. which is practically hided below the intense broad band around 3404 cm<sup>-1</sup> attributed to O–H bond vibration corresponding to the carboxylate group of the of diclofenac free ligand. At lower frequency, a set of sharp and of medium intensity bands that corresponds to aromatic ring's C–H bond vibrations of the ligand are visible between 3373 cm-1 and 3289 cm-1 . The following intense vibrations located in 1589–1442 cm-1 region are attributed to both the asymmetric stretching vibrations of the carboxylate groups and the aromatic C–C bonds. Moving to lower range of 1373–1268 cm-1 symmetric stretching vibrations of the carboxylate groups appear in the spectrum. The remaining bands that are found at lower frequency can be attributed to the distortions originated in the aromatic ring and the carboxylate groups of the ligands.



**Figure A5.1.** Infrared spectra of the ligand and compound **5.5** and **5.12-5.13.**

# **A5.3 Crystallographic data**



**Table A5.2.** Crystallographic data and structure refinement details of compound **5.7**, **5.12** and **5.14**.

5.7 <sub>Py</sub>				$5.126d-EtoH$			5.14 <sub>Fe</sub>		
Atom	Atom	Length/Å	Atom	Atom	Length/Å	Atom	<b>Atom</b>	Length/Å	
Dy1	O <sub>1</sub> B	2.451(3)	Gd1	O <sub>2</sub> C	2.366(4)	Fe1	O <sub>1</sub> G	1.908(4)	
Dy1	O <sub>1</sub> C	2.442(3)	Gd1	O <sub>2</sub> C <sub>1</sub>	2.564(4)	Fe1	O <sub>1</sub> E	2.027(4)	
Dy1	O <sub>2</sub> A	2.429(3)	Gd1	O <sub>1</sub> A	2.438(4)	Fe1	O <sub>1</sub> B	2.009(5)	
Dy1	O <sub>2</sub> B	2.490(3)	Gd1	O <sub>1</sub> B	2.462(4)	Fe1	O1C1	2.002(5)	
Dy1	O <sub>1</sub> A	2.413(3)	Gd1	O1C1	2.466(5)	Fe1	O <sub>1</sub> A	2.015(5)	
Dy1	O <sub>1</sub> M	2.375(3)	Gd1	O <sub>1</sub> E	2.395(4)	Fe1	O <sub>1</sub> H	2.074(4)	
Dy1	O <sub>1</sub> W	2.362(3)	Gd1	O <sub>2</sub> B	2.520(4)	Fe <sub>2</sub>	O <sub>2</sub> E	2.041(4)	
Dy1	O <sub>2</sub> C	2.549(3)	Gd1	O <sub>2</sub> A	2.464(4)	Fe <sub>2</sub>	O <sub>1</sub> G	1.890(5)	
Dy1	O <sub>2</sub> C <sub>1</sub>	2.327(3)	Gd1	O <sub>1</sub> W	2.392(4)	Fe <sub>2</sub>	O1G1	2.071(5)	
O <sub>2</sub> C	Dy11	2.327(3)	O <sub>2</sub> C	Gd11	2.564(4)	Fe <sub>2</sub>	O <sub>1</sub> D	2.022(5)	
	$11-x,-y,-z$		O <sub>1</sub> C	Gd11	2.466(5)	Fe <sub>2</sub>	O <sub>1</sub> F	2.011(5)	
				$1 - 1 - x, -y, 1 - z$		Fe <sub>2</sub>	O <sub>2</sub> A	2.030(5)	
						Fe3	O <sub>1</sub> G	1.936(5)	
						Fe3	O <sub>2</sub> D	1.999(7)	
						Fe3	O <sub>2</sub> F	2.033(5)	
						Fe3	O11	2.077(6)	
						Fe3	O <sub>2</sub> B	2.021(5)	
						Fe3	O <sub>2</sub> C	2.008(6)	

**Table A5.3.** Table of the selected bond lengths (Å) for compound **5.7, 5.12** and **5.14.**

		5.7 <sub>Py</sub>				$\overline{5.12_{Gd-EtOH}}$				$5.14_{Fe}$	
Atom	Atom	Atom	Angle/°	Atom	Atom	Atom	Angle/°	Atom	Atom	Atom	Angle/°
O <sub>1</sub> B	Dy1	O <sub>2</sub> B	52.86(10)	O <sub>2</sub> C	Gd1	O2C <sup>1</sup>	67.54(17)	O <sub>1</sub> G	Fe1	$\overline{O1E}$	92.44(18)
O <sub>1</sub> B	Dy1	O <sub>2</sub> C	90.63(10)	O <sub>2</sub> C	Gd1	O <sub>1</sub> A	105.22(15)	O <sub>1</sub> G	Fe1	O <sub>1</sub> B	93.61(19)
O <sub>1</sub> C	Dy1	O <sub>1</sub> B	71.74(11)	O <sub>2</sub> C	Gd1	O <sub>1</sub> B	128.50(15)	O <sub>1</sub> G	Fe1	O1C1	98.5(2)
O <sub>1</sub> C	Dy1	O <sub>2</sub> B	99.99(11)	O <sub>2</sub> C	Gd1	O1C <sup>1</sup>	116.30(15)	O <sub>1</sub> G	Fe1	O <sub>1</sub> A	95.32(19)
O <sub>1</sub> C	Dy1	O <sub>2</sub> C	51.87(10)	O <sub>2</sub> C	Gd1	O <sub>1</sub> E	75.12(15)	O <sub>1</sub> G	Fe1	O <sub>1</sub> H	179.0(2)
O <sub>2</sub> A	Dy1	O <sub>1</sub> B	122.16(10)	O <sub>2</sub> C	Gd1	O <sub>2</sub> B	76.68(15)	O <sub>1</sub> E	Fe1	O <sub>1</sub> H	87.63(18)
O <sub>2</sub> A	Dy1	O <sub>1</sub> C	142.15(10)	O <sub>2</sub> C	Gd1	O <sub>2</sub> A	83.33(15)	O <sub>1</sub> B	Fe1	O <sub>1</sub> E	172.80(19)
O <sub>2</sub> A	Dy1	O <sub>2</sub> B	116.46(11)	O <sub>2</sub> C	Gd1	O <sub>1</sub> W	154.29(15)	O <sub>1</sub> B	Fe1	O <sub>1</sub> A	91.2(2)
O <sub>2</sub> A	Dy1	O <sub>2</sub> C	145.89(10)	O <sub>1</sub> A	Gd1	O2C <sup>1</sup>	149.42(15)	O <sub>1</sub> B	Fe1	O <sub>1</sub> H	86.39(18)
O <sub>2</sub> B	Dy1	O <sub>2</sub> C	74.15(10)	O <sub>1</sub> A	Gd1	O <sub>1</sub> B	71.39(15)	O <sub>1</sub> C <sub>1</sub>	Fe1	O <sub>1</sub> E	84.34(19)
O <sub>1</sub> A	Dy1	O <sub>1</sub> B	71.04(10)	O <sub>1</sub> A	Gd1	O1C <sup>1</sup>	136.26(15)	O1C1	Fe1	O <sub>1</sub> B	90.9(2)
O <sub>1</sub> A	Dy1	O <sub>1</sub> C	136.94(11)	O <sub>1</sub> A	Gd1	O <sub>2</sub> B	74.47(15)	O <sub>1</sub> C <sub>1</sub>	Fe1	O <sub>1</sub> A	165.80(19)
O <sub>1</sub> A	Dy1	O <sub>2</sub> A	53.70(11)	O <sub>1</sub> A	Gd1	O <sub>2</sub> A	53.69(15)	O1C1	Fe1	O <sub>1</sub> H	82.43(18)
O <sub>1</sub> A	Dy1	O <sub>2</sub> B	73.60(11)	O <sub>1</sub> B	Gd1	O2C <sup>1</sup>	89.75(15)	O <sub>1</sub> A	Fe1	O <sub>1</sub> E	92.08(19)
O <sub>1</sub> A	Dy1	O <sub>2</sub> C	147.71(10)	O <sub>1</sub> B	Gd1	O1C <sup>1</sup>	72.35(15)	O <sub>1</sub> A	Fe1	O <sub>1</sub> H	83.70(17)
O <sub>1</sub> M	Dy1	O <sub>1</sub> B	147.21(10)	O <sub>1</sub> B	Gd1	O <sub>2</sub> B	52.44(14)	O <sub>2</sub> E	Fe <sub>2</sub>	O1G1	83.86(19)
O <sub>1</sub> M	Dy1	O <sub>1</sub> C	77.53(11)	O <sub>1</sub> B	Gd1	O <sub>2</sub> A	123.12(16)	O <sub>1</sub> G	Fe <sub>2</sub>	O <sub>2</sub> E	94.63(18)
O <sub>1</sub> M	Dy1	O <sub>2</sub> A	77.21(11)	O1C <sup>1</sup>	Gd1	O2C <sup>1</sup>	51.57(14)	O <sub>1</sub> G	Fe <sub>2</sub>	O1G1	178.0(2)
O <sub>1</sub> M	Dy1	O <sub>2</sub> B	146.88(10)	O1C <sup>1</sup>	Gd1	O <sub>2</sub> B	101.44(15)	O <sub>1</sub> G	Fe <sub>2</sub>	O <sub>1</sub> D	96.2(2)
O <sub>1</sub> M	Dy1	O <sub>1</sub> A	130.23(11)	O <sub>1</sub> E	Gd1	O2C <sup>1</sup>	77.56(14)	O <sub>1</sub> G	Fe <sub>2</sub>	O <sub>1</sub> F	94.1(2)
O <sub>1</sub> M	Dy1	O <sub>2</sub> C	79.00(10)	O <sub>1</sub> E	Gd1	O <sub>1</sub> A	130.88(15)	O <sub>1</sub> G	Fe <sub>2</sub>	O <sub>2</sub> A	94.7(2)
O <sub>1</sub> W	Dy1	O <sub>1</sub> B	77.62(10)	O <sub>1</sub> E	Gd1	O <sub>1</sub> B	146.42(16)	O <sub>1</sub> D	Fe <sub>2</sub>	O <sub>2</sub> E	169.0(2)
O <sub>1</sub> W	Dy1	O <sub>1</sub> C	72.93(10)	O <sub>1</sub> E	Gd1	O1C <sup>1</sup>	75.48(15)	O <sub>1</sub> D	Fe <sub>2</sub>	O1G1	85.4(2)
O <sub>1</sub> W	Dy1	O <sub>2</sub> A	76.32(10)	O <sub>1</sub> E	Gd1	O <sub>2</sub> B	146.35(15)	O <sub>1</sub> D	Fe <sub>2</sub>	O <sub>2</sub> A	87.2(2)
O <sub>1</sub> W	Dy1	O <sub>2</sub> B	128.64(11)	O <sub>1</sub> E	Gd1	O <sub>2</sub> A	78.22(15)	O <sub>1</sub> F	Fe <sub>2</sub>	O <sub>2</sub> E	86.9(2)
O <sub>1</sub> W	Dy1	O <sub>1</sub> A	78.63(11)	O <sub>2</sub> B	Gd1	O2C <sup>1</sup>	74.96(14)	O <sub>1</sub> F	Fe <sub>2</sub>	O1G1	84.5(2)
O <sub>1</sub> W	Dy1	O <sub>1</sub> M	82.77(10)	O <sub>2</sub> A	Gd1	O2C <sup>1</sup>	145.95(14)	O <sub>1</sub> F	Fe <sub>2</sub>	O <sub>1</sub> D	94.1(2)
O <sub>1</sub> W	Dy1	O <sub>2</sub> C	124.28(10)	O <sub>2</sub> A	Gd1	O1C <sup>1</sup>	141.05(15)	O <sub>1</sub> F	Fe <sub>2</sub>	O <sub>2</sub> A	170.9(2)
O <sub>2</sub> C <sub>1</sub>	Dy1	O <sub>1</sub> B	128.13(10)	O <sub>2</sub> A	Gd1	O <sub>2</sub> B	116.23(15)	O <sub>2</sub> A	Fe <sub>2</sub>	O <sub>2</sub> E	90.15(19)
O <sub>2</sub> C <sub>1</sub>	Dy1	O <sub>1</sub> C	117.04(11)	O <sub>1</sub> W	Gd1	O2C <sup>1</sup>	122.93(15)	O <sub>2</sub> A	Fe <sub>2</sub>	O1G1	86.6(2)
O <sub>2</sub> C <sub>1</sub>	Dy1	O <sub>2</sub> A	83.42(10)	O <sub>1</sub> W	Gd1	O <sub>1</sub> A	77.02(15)	O <sub>1</sub> G	Fe3	O <sub>2</sub> D	95.2(2)
O <sub>2</sub> C <sub>1</sub>	Dy1	O <sub>2</sub> B	75.65(10)	O <sub>1</sub> W	Gd1	O <sub>1</sub> B	76.89(15)	O <sub>1</sub> G	Fe3	O <sub>2</sub> F	95.3(2)
O <sub>2</sub> C <sub>1</sub>	Dy1	O <sub>1</sub> A	102.79(11)	O <sub>1</sub> W	Gd1	O1C <sup>1</sup>	71.64(15)	O <sub>1</sub> G	Fe3	O1I	176.2(2)
O <sub>2</sub> C <sub>1</sub>	Dy1	O <sub>1</sub> M	76.36(10)	O <sub>1</sub> W	Gd1	O <sub>1</sub> E	84.15(15)	O <sub>1</sub> G	Fe3	O <sub>2</sub> B	92.5(2)
O <sub>2</sub> C <sub>1</sub>	Dy1	O <sub>1</sub> W	153.63(10)	O <sub>1</sub> W	Gd1	O <sub>2</sub> B	127.39(14)	O <sub>1</sub> G	Fe3	O <sub>2</sub> C	94.6(2)
O <sub>2</sub> C <sub>1</sub>	Dy1	O <sub>2</sub> C	67.38(11)	O <sub>1</sub> W	Gd1	O <sub>2</sub> A	77.59(15)	O <sub>2</sub> D	Fe3	O <sub>2</sub> F	89.0(3)

**Table A5.4.** Table of the selected angles (º) for compound **5.7**, **5.12** and **5.14.**



**Table A5.5.** Structural parameters of hydrogen bonds (Å. °) in compound **5.7Dy. a**

$D$ -H $\cdots$ A $b$	D-H	<i>Н. А</i>	D A	D-H…A
O1w-H1wa $\cdots$ O1A(i) 0.85		2.02	2.830(5)	159.7
O1w-H1wb $\cdots$ O1B(i) 0.85		1.99	2.743(4)	145.7

 $a$ Symmetry codes: (i)  $-x$ .  $-y$ .  $-z$ .  $b$ D: donor. A: acceptor

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## **A5.4 Powder X-ray diffraction analysis**

**Figure A5.2.** Figure of the pattern matching analysis and experimental PXRD for complexes **5.1**-**5.6**.



**Figure A5.3.** Figure of the pattern matching analysis and experimental PXRD for complexes **5.7**-**5.12**.



**Figure A5.4.** Figure of the pattern matching analysis and experimental PXRD for gadolinium-based complexes **5.5** and **5.12-5.13**.

### **A5.5 Continuous Shape Measurements**

CShMs for the coordination environment of **5.7Dy**, **5.12Gd-EtOH** and **5.14Fe**. The lowest SHAPE values for each ion are shown highlighted in grey, indicating best fits.

Table A5.6. Table of the continuous Shape Measurements for the LnO<sub>9</sub> coordination environment.





Table A5.7. Table of the continuous Shape Measurements for the FeO<sub>6</sub> coordination environment.





#### **A5.6 Thermal analysis**

Thermogravimetric analyses have been performed over polycrystalline sample in compounds **5.5, 5.12-5.13** in order to check the stability of the products. The TG curves show three main steps of weight loses. The first steps concern to the release of coordinated solvent molecules which are released from room temperature up to 160 ºC. The second step refers to the decomposition of the ligands which involves the collapse of the crystal structure, evolving to Gd<sub>2</sub>O<sub>3</sub> obtained at 800 °C as the final residue.



**Figure A5.5.** Figure of TG analysis of compound **5.5Gd-MeOH**, **5.12Gd-EtOH** and **5.13Gd-water**.



## **A5.7 Additional views of the structure**

**Figure A5.6.** Most representative intermolecular interactions: H bonds found in compound **5.6Dy** are highlighted in green.

### **A5.8 Magnetic properties**



**Figure A5.7.** Temperature dependence of out-of-phase components of the ac susceptibility in a dc applied field of 1000 Oe for 5.7<sub>Dy.</sub>



**Figure A5.8.**Theoretical orientation of the magnetic moments (green line) for Dy<sup>3+</sup> ions in 5.7<sub>Dy</sub>. Left the asymmetric unit, in right the coordination compound.



**A5.9 Photoluminescent properties**

**Figure A5.9**. <sup>5</sup>D<sub>0</sub> decay curves monitoring the emissions at 619 nm with the excitations selected at 325 nm, respectively for **5.4Eu** at various temperatures; the solid red lines are the best fits using first-order decay functions,  $y = y_0 + A_1 \cdot \exp(-x/\tau_1)$ .



**Figure A5.10.** <sup>5</sup>D<sub>4</sub> decay curves monitoring the emissions at 546 nm with the excitations selected at 325 nm, respectively for **5.6Tb** at various temperatures; the solid red lines are the best fits using first-order decay functions,  $y = y_0 + A_1 \cdot \exp(-x/\tau_1)$ .

### **A5.10 TD-DFT calculations**



**Table A5.8.** Calculated main excitation energies (nm) and singlet electronic transitions and associated oscillator strengths of diclofenac molecule in gas phase.

**Table A5.9.** Calculated main emission energies (nm) and singlet electronic transitions and associated oscillator strengths of diclofenac molecule in gas phase.

Calcd. $\lambda$ (nm)	$Exp. \lambda$ (nm)	<b>Significant contributions</b>	Osc. strength (a.u.)
370	386	$HOMO - 3 \leftarrow LUMO + 1 (96%)$	0.0844
365	386	$HOMO - 3 \leftarrow LUMO + 1 (96%)$	0.0681
486	462	$HOMO - 2 \leftarrow LUMO + 1 (94%)$ $HOMO \leftarrow LUMO + 2(5\%)$	0.0099











**Figure A5.13.** Lowest Unoccupied Molecular Orbitals of diclofenac molecule involved in the singlet excitation transitions.



**Figure A5.14.** Lowest Unoccupied Molecular Orbitals of diclofenac molecule involved in the singlet emission transitions**.**

### **A5.11 Encapsulation in liposomes and characterization**

Physical-chemical characterization was performed by dynamic light scattering (DLS) and the determination of metal content using inductively coupled plasma mass spectrometer (ICP-MS).

#### **A5.11.1 Liposomes characterization: DLS and Z potential**



**Table A5.10.** Characterization of liposomes.

#### **A5.11.2 Quantification of metal content by ICP-MS**

**Table A5.11.** Metal content determined by ICP-MS.



#### **A5.11.3 Determination of phosphates by Rouser method**

Rouser method was applied to determine the lipid concentration transformed into liposomes from the initial phosphate concentration. For that purpose, phosphate free buffers were required for lipid rehydration process. In view of the synthetic problems encountered when directly rehydrating liposomes with HEPES or HPLC water, we decided to rehydrate with PBS and subsequently exchange solvent by washing several times with phosphate-free buffers. Additionally, as reference material we performed the same procedure with PBS buffer and exchange it with HEPES and HPLC water.



**Table A5.12.** Phosphate concentration in mM for Gd, Eu and Fe liposomes after being rehydrated with PBS and solvent exchange procedure was performed for liposomes and the buffer. Exhibited results are averages of the triplicated data.

In view of the obtained results, it can be concluded that the approach to exchange liposome's rehydration solvent by centrifugation was efficient and we successfully performed it, since as it is summarized in Table A5.9 the phosphate concentration in reference or blank material was practically negligible.

# **Appendix 6**

Supporting information of Chapter 6.A1
## **A6.1 Experimental section. General instrumentation**

**Elemental analyses** (EA) of synthetized novel catalyst compound **Y/Eu-MOF** performed on an Elementar vario EL cube in the CHN mode.

**IR** (ATR) spectra of 3-amino-4-hydroxybenzoic acid ligand and compound **Y/Eu-MOF**  collected in the region 400–4000 cm−1 on a Nicolet 6700 FTIR (Fourier transform infrared) spectrophotometer (Thermo Phisher Scientific, TX, USA) KBr pellets.

**NMR measurements**: NMR spectra were measured in a Bruker Avance III 300 spectrometer equipped with a direct double SmartProbe BBFO 1H/BB(19F) probe. Chemical shifts are reported in parts per million (ppm) relative to residual solvent peak (CDCl<sub>3</sub>, <sup>1</sup>H: 7.26 ppm; 13C: 77.16 ppm). Coupling constants are reported in Hertz. Multiplicity is reported with the usual abbreviations (s: singlet, bs: broad singlet, d: doublet, dd: doublet of doublets, ddd: doublet of doublet of doublets, t: triplet, td: triplet of doublets, q: quartet, dq: doublet of quartet, p: pentet, sex: sextet, hept: heptet, m: multiplet). Quantitative NMR acquisition parameters. <sup>1</sup>H NMR determination of product conversion was carried out by comparing signals arising from both CH of aldehyde or CH<sup>3</sup> of ketone **1** and products **2**, **3** or **4**. The standard acquisition parameters were one-dimensional pulse sequence which includes a 30º flip angle (Bruker zg30), recycle time (D1  $=$  30 s), time domain (TD = 27k), number of scans (NS = 32), acquisition time (AQ = 2.05 s), transmitter (frequency) offset  $(O1P = 6.0$  ppm), and spectral width  $(SW = 22.0$  ppm).

**ICP-AES analysis** of catalyst 6.1<sub>Y-Eu</sub> was conducted in Horiba Yobin Yvon Activa atomic emission spectrometer equipped with a glass and Teflon nebulizer system, which enables samples from acidic digestion to be determined using HF. This equipment also provides the option of coupling a hydride generation system which enables elements such as As, Hg and Sb,.etc. to be determined, in very low levels of concentration (ppb). The equipment is controlled by a control computer with Activa Analyst 5.4 software, which enables it to interact with the equipment at all times. It permits sequential multi-elemental analysis and also enables numerous analytical requirements to be met due to the large linear interval that characterizes this technique, which in turn facilitates the analysis of majority and minority elements.

**SEM-EDX**, Scanning Electron Microscopy (SEM Leo 1430 VP) with microanalysis of elements (EDX): Scanning electron microscopy, SEM Leo 1430VP, linked to a system for the microanalysis of elements via energy-dispersive X-ray spectroscopy, Inca 350, version 17 (hereinafter, SEM-EDX). Scanning electron microscopy enables the identification of elements with low atomic numbers, including carbon.

**X-ray Diffraction Data Collection** and Structure Determination. Single-crystal diffraction data were collected at 100(2) K on a Bruker X8 APEX II and Bruker D8 Venture with a Photon detector equipped with graphite monochromated MoKα radiation ( $\lambda = 0.71073$  Å). The data reduction was performed with the APEX2 software[37] and corrected for absorption using

SADABS.[2] These structures were solved by direct methods using the SHELXT program[38] and refined by full-matrix least-squares of F2 including all reflections with SHELXL-2018/3 program.[39] All calculations for these structures were performed using the WINGX crystallographic software package.[4]

**Powder X-ray diffractions** (XRPD) patterns were collected on a Philips X'PERT powder diffractometer with Cu Kα radiation (λ = 1.5418 Å) over the range of 5 < 2*θ* < 50° with a step size of 0.02° and an acquisition time of 2.5 s per step at 25 °C. Indexation of the diffraction profiles were carried out using the FULLPROF program,[1] on the basis of the space group and cell parameters found for isostructural compounds by single crystal X-ray diffraction.

# **A6.2 Chemical characterization**

### **A6.2.1 Elemental Analysis**





#### **A6.2.2 Determination of the metal content by ICP-AES**

**Table A6.2.** Determination of the metal content by ICP and Y to Eu relationship.



#### **A6.2.3 FT-IR spectroscopy**

FTIR spectrum of **Y/Eu-MOF** display a narrow peak at around 3617 cm-1 attributed to the N–H stretching vibration of the amine group of 3-amino-4-hydroxbenzoate ligand. Also, an intense and broad band attributable to O–H bond of free ligand along with a set of weak bands between 3325 and 2917 cm-1 corresponding to the C–H vibrations of the aromatic ring of the 3-amino-4-hydroxybenzoate ligand are visible in the spectrum. Peaks in 1683–1408cm<sup>-1</sup> region are associated with asymmetric stretching vibrations of the carboxylate groups and the aromatic C–C and C–N bonds. The group of signals at lower range, 1390–1263 cm-1 , can be linked to symmetric stretching vibrations of the carboxylate groups. Remaining bands found at lower frequencies originated by distortions in the aromatic and carboxylate group of the ligand. Note that vibrational bands M–O and M–N bonds are observed below 660 cm-1 .



**Figure A6.1.** Figure of the infrared spectra of the ligand and Y/Eu-MOF.

# **A6.3 Crystallographic data**



**Table A6.3.** Crystallographic data and structure refinement details of compound **6.1Y-Eu**.

Atom	Atom	Length/Å	
Eu1	Eu1 <sup>2</sup>	3.5335(7)	
Eu1	Y2	3.9093(3)	
Eu1	Y2 <sup>3</sup>	3.9093(3)	
Eu <sub>2</sub>	Y1	3.9093(3)	
N1	Eu1	2.507(3)	
N1	Y1	2.507(3)	
O <sub>1</sub>	Eu2	2.341(2)	
O <sub>1</sub>	Y1	2.512(2)	
O <sub>1</sub>	Eu1	2.512(2)	
O <sub>1</sub>	Y2	2.341(2)	
O <sub>2</sub>	Y2 <sup>1</sup>	2.419(2)	
O <sub>3</sub>	Y2 <sup>1</sup>	2.428(2)	
O <sub>4</sub>	Eu1	2.3830(19)	
O <sub>4</sub>	Eu1 <sup>2</sup>	2.3831(19)	
O <sub>4</sub>	Y2	2.321(3)	
O <sub>4</sub>	Eu <sub>2</sub>	2.321(3)	
O <sub>4</sub>	Y1	2.3830(19)	
O <sub>5</sub>	Y2	2.332(3)	
O <sub>5</sub>	Eu2	2.332(3)	
		$1+y$ , -x+y, 1-z; $2+x$ , +y, 1/2-z; $31-y$ ; +x-y, +z	

**Table A6.4.** Table of the selected bond lengths (Å) and angles (°) for compound **6.1Y-Eu**.









**A6.4 Powder X-ray diffraction analysis**

**Figure A6.2.** Figure of the pattern matching analysis and experimental PXRD for Y/Eu-MOF.

# **A6.5 Continuous Shape Measurements**

CShMs for the coordination environment of **6.1Y-Eu**. The lowest SHAPE values for each ion is shown highlighted in grey, indicating best fits.

$EP-9$	$D_{9h}$	Enneagon
OPY-9	$C_{8v}$	Octagonal pyramid
HBPY-9	$D_{7h}$	Heptagonal bipyramid
JTC-9	$C_{3v}$	Johnson triangular cupola J3
JCCU-9	$C_{4v}$	Capped cube J8
CCU-9	$C_{4v}$	Spherical-relaxed capped cube
JCSAPR-9	$C_{4v}$	Capped square antiprism J10
CSAPR-9	$C_{4v}$	Spherical capped square antiprism
JTCTPR-9	$D_{3h}$	Tricapped trigonal prism J51
TCTPR-9	$D_{3h}$	Spherical tricapped trigonal prism
JTDIC-9	$C_{3v}$	Tridiminished icosahedron J63
$HH-9$	$C_{2v}$	Hula-hoop
MFF-9	$C_{\rm s}$	Muffin

Table A6.5. Table of the continuous Shape Measurements for the MN<sub>3</sub>O<sub>6</sub> coordination environment.



Table A6.6. Table of the continuous Shape Measurements for the MO<sub>8</sub> coordination environment.







# **A6.6 Scanning Electron Microscopy**







# **A6.7 Optimization of the hydroboration reaction conditions**

Table A6.7. Optimization of the reaction conditions in the hydroboration reaction.<sup>a</sup>



<sup>a</sup> Reaction carried out using acetophenone (28 µL, 0.25 mmol), HBPin (40 µL, 0.275 mmol) in 0.5 mL of the corresponding solvent at room temperature and under nitrogen inert atmosphere during 24 h. bConversions (relative to acetophenone) determined by <sup>1</sup>H NMR of the reaction crude.

## **A6.8 Characterization Data of Products**

#### *Cyanosilylated carbonyl compounds catalysed by 6.1Y-Eu*

**2-Phenyl-2-((trimethylsilyl)oxy)acetonitrile (2a).** This product has been previously reported.[10] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 7.50–7.35 (m, 5H, ArH), 5.50 (s, 1H, *CH*CN), 0.23 (s, 9H, TMS) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 136.2 (C*ipso*), 129.3(ArCH), 128.9 (ArCH), 126.3 (ArCH), 119.1 (CN), 63.6 (CH), -0.29 (TMS) ppm.

**2-(4-Methoxyphenyl)-2-((trimethylsilyl)oxy)acetonitrile (2b).** This product has been previously reported.[12] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 7.38 (d, *J* = 8.6 Hz, 2H, ArH), 6.92 (d, *J* = 8.6 Hz, 2H, ArH), 5.43 (s, 1H, *CH*CN), 3.82 (s, 3H, OMe), -0.21 (s, 9H, TMS) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 160.3 (C*ipso*), 128.3 (C*ipso*), 127.9 (ArCH), 119.3 (CN), 114.2 (ArCH), 63.3 (CH), 55.3 (OCH3), -0.24 (TMS) ppm.

**2-(4-Chlorophenyl)-2-((trimethylsilyl)oxy)acetonitrile (2c).** This product has been previously reported.[14] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 7.45–7.40 (m, 5H, ArH), 5.49 (s, 1H, *CH*CN), 0.26 (s, 9H, TMS) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 135.3 (C*ipso*), 134.8 (C*ipso*), 129.1 (ArCH), 127.7 (ArCH), 118.8 (CN), 63.0 (CH), -0.30 (TMS) ppm.

**2-(Pyridin-2-yl)-2-((trimethylsilyl)oxy)acetonitrile (2d).** This product has been previously reported.[15] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 8.60–8.55 (m, 1H, ArH), 7.79 (dt, *J* = 7.7, 1.7 Hz, 1H, ArH), 7.59 (d, *J* = 7.7 Hz, 1H, ArH), 7.35–7.25 (m, 1H, ArH), 5.58 (s, 1H, *CH*CN), 0.26 (s, 9H, TMS) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 155.4 (C*ipso*), 149.3 (ArCH), 137.5(ArCH), 124.0 (ArCH), 120.5 (ArCH), 118.6 (CN), 65.1 (CH), -0.37 (TMS) ppm.

**2-((Trimethylsilyl)oxy)butanenitrile (2e)**. This product has been previously reported.[17] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 4.34 (t, *J* = 6.3 Hz, 1H, CH), 1.85–1.75 (m, 2H, CH2), 1.04 (t, *J*  = 7.4 Hz, 3H, CH<sub>3</sub>), 0.21 (s, 9H, CH<sub>3</sub> x 3) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl<sub>3</sub>): δ 119.9 (CN), 62.7  $(CH)$ , 29.6  $(CH<sub>2</sub>)$ , 8.9  $(CH<sub>3</sub>)$ , 0.4  $(TMS)$  ppm.

**2-Phenyl-2-((trimethylsilyl)oxy)propanenitrile (3a).** This product has been previously reported.[19] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 7.60–7.50 (m, 2H, ArH), 7.45–7.30 (m, 3H, ArH), 1.86 (s, 3H, CH3), 0.18 (s, 9H, TMS) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 142.0 (C*ipso*), 128.68 (ArCH), 128.66 (ArCH), 124.6 (ArCH), 121.6 (CN), 71.6 (C), 33.5 (CH3), 1.03 (TMS) ppm.

**2-(4-Methoxyphenyl)-2-((trimethylsilyl)oxy)propanenitrile (3b).** This product has been previously reported.[22] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 7.46 (d, *J* = 8.7 Hz, 2H, ArH), 6.91 (d, *J*  $= 8.7$  Hz, 2H, ArH), 3.82 (s, 3H, OMe), 1.85 (s, 3H, CH<sub>3</sub>), 0.16 (s, 9H, TMS) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 159.7 (C*ipso*), 133.9 (C*ipso*), 126.0 (ArCH), 121.7 (CN), 113.8 (ArCH), 71.2 (C), 55.2 (OCH3), 33.3 (CH3), 1.00 (TMS) ppm.

**2-(4-Chlorophenyl)-2-((trimethylsilyl)oxy)propanenitrile (3c).** This product has been previously reported.[22] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 7.48 (d, *J* = 8.5 Hz, 2H, ArH), 7.37 (d, *J*  $= 8.5$  Hz, 2H, ArH), 1.83 (s, 3H, CH<sub>3</sub>), 0.19 (s, 9H, TMS) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl<sub>3</sub>): δ 140.6 (C*ipso*), 134.5 (C*ipso*), 128.8 (ArCH), 126.0 (ArCH), 121.2 (CN), 71.0 (C), 33.4 (CH3), 1.00 (TMS) ppm.

**2-(2,4-Difluorophenyl)-2-((trimethylsilyl)oxy)propanenitrile (3d).** This product has been previously reported.[28] <sup>1</sup>H NMR (500.13 MHz, CDCl3): δ 7.56 (td, *J* = 8.8, 6.4 Hz, 1H, ArH), 6.95– 6.90 (m, 1H, ArH), 6.86 (ddd, *J* = 11.1, 8.8, 2.5 Hz, 1H, ArH), 1.92 (s, 3H, CH3), 0.27 (s, 9H, TMS) ppm. <sup>13</sup>C NMR (125.77 MHz, CDCl3): δ 163.2 (dd, *J* = 250.8, 12.0 Hz, C*ipso*-F), 159.4 (dd, *J* = 252.5, 12.0 Hz, C*ipso*-F), 127.8 (dd, *J* = 9.7, 4.3 Hz, ArCH), 125.0 (dd, *J* = 11.2, 3.9 Hz, C*ipso*-F), 120.4 (CN), 111.2 (d, *J* = 21.2 Hz, ArCH), 104.9 (t, *J* = 25.6 Hz, ArCH), 68.0 (d, *J* = 2.0 Hz, C), 30.8 (d, *J* = 2.9 Hz, CH3), 1.08 (TMS) ppm.

**2-(Pyridin-2-yl)-2-((trimethylsilyl)oxy)propanenitrile (3e)**. This product has been previously reported.[29] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 8.62 (d, *J* = 4.7 Hz, 1H, ArH), 7.77 (t, *J*  $= 7.8$  Hz, 1H, ArH), 7.60 (d, J = 7.8 Hz, 1H, ArH), 7.30–7.25 (m, 1H, ArH), 1.93 (s, 3H, CH<sub>3</sub>), 0.26 (s, 9H, TMS) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 160.0 (C*ipso*), 149.0 (ArCH), 137.2 (ArCH), 123.4 (ArCH), 121.3 (CN), 118.9 (ArCH), 72.9 (C), 31.2 (CH3), 1.06 (TMS) ppm.

**2-Methyl-2-((trimethylsilyl)oxy)butanenitrile (3f).** This product has been previously reported.[22] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 1.85–1.65 (m, 2H, CH2), 1.55 (s, 3H, CH3), 1.04 (t, *J* = 7.4 Hz, 3H, *CH3*CH2), 0.23 (s, 9H, TMS) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 121.9 (CN), 70.2 (C), 36.4 (CH<sub>2</sub>), 28.4 (CH<sub>3</sub>), 8.6 (CH<sub>3</sub>), 1.21 (TMS) ppm.

**1-((Trimethylsilyl)oxy)cyclohexane-1-carbonitrile (3g).** This product has been previously reported.[22] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 2.10–2.00 (m, 2H, CH2), 1.75–1.70 (m, 2H, CH<sub>2</sub>), 1.70–1.45 (m, 6H, CH<sub>2</sub> x 3), 1.30–1.20 (m, 2H, CH<sub>2</sub>), 0.23 (s, 9H, TMS) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 121.9 (CN), 70.6 (C), 39.3 (CH2), 24.5 (CH2), 22.6 (CH2), 1.38 (TMS) ppm.

#### *Hydroborated carbonyl compounds catalysed by 6.1Y-Eu*

**1-Phenylethan-1-ol** (**4a**). This product has been previously reported.[40] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 7.30–7.20 (m, 5H, ArH), 4.82 (q, *J* = 6.5 Hz, 1H, CH), 1.83 (br s, 1H, OH), 1.42 (d, *J* = 6.5 Hz, 3H, CH3) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 145.8 (C*ipso*), 128.5 (ArCH), 127.5 (ArCH), 125.3 (ArCH), 70.4 (CH), 25.1 (CH3) ppm.

**1-(4-Isobutylphenyl)ethan-1-ol** (**4b**). This product has been previously reported.[41] <sup>1</sup>H NMR (400 MHz, CDCl3): δ 7.27 (d, *J* = 8.0 Hz, 2H, ArH), 7.12 (d, *J* = 8.0 Hz, 2H, ArH), 4.86 (q, *J* = 6.4 Hz, 1H, CH), 2.46 (d, *J* = 7.2 Hz, 2H, CH2), 2.12 (br s, 1H, OH), 2.00–1.75 (m, 1H, CH),

1.48 (d, *J* = 6.5 Hz, 3H, C*H*3CHOH), 0.90 (d, *J* = 6.4 Hz, 6H, CH<sup>3</sup> x 2). <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 143.0, 141.0, 129.2, 125.2, 70.3, 45.1, 30.2, 25.0, 22.4 ppm.

**1-(4-Methoxyphenyl)ethan-1-ol** (**4c**). This product has been previously reported.[40] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 7.30 (d, *J* = 8.6 Hz, 2H, ArH), 6.88 (d, *J* = 8.6 Hz, 2H, ArH), 7.85 (q, *J* = 6.5 Hz, 1H, CH), 3.8 (s, 3H, MeO), 1.48 (d, *J* = 6.5 Hz, 3H, CH3) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 159.0 (C*ipso*), 138.0 (C*ipso*), 130.6 (ArCH), 113.6 (ArCH), 70.0 (CH), 55.2 (OMe), 25.0 (CH3) ppm.

**1-(4-Chlorophenyl)ethan-1-ol** (**4d**). This product has been previously reported.[40] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 7.30–7.25 (m, 5H, ArH), 4.88 (q, *J* = 6.5 Hz, 1H, CH), 1.85 (br s, 1H, OH), 1.47 (d, *J* = 6.5 Hz, 3H, CH3) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 144.2 (C*ipso*), 133.0 (C*ipso*), 128.6 (ArCH), 126.8 (ArCH), 69.7 (CH), 25.2 (CH3) ppm.

**1-(3-Chlorophenyl)ethan-1-ol (4e**). This product has been previously reported.[42] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 7.40–7.35 (m, 1H, ArH), 7.30–7.25 (m, 3H, ArH), 4.91 (q, *J* = 6.4 Hz, 1H, CH), 1.83 (br s, 1H, OH), 1.51 (d, *J* = 6.4 Hz, 3H, CH3) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 147.8 (C*ipso*), 134.4 (C*ipso*), 129.8 (ArCH), 127.6 (ArCH), 125.6 (ArCH), 123.5 (ArCH), 69.8 (CH), 25.3 (CH3) ppm.

**1-(2-Chlorophenyl)ethan-1-ol (4f)**. This product has been previously reported.[42] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 7.61 (d, *J* = 7.7 Hz, 1H, ArH), 7.35–7.30 (m, 2H, ArH), 7.25–7.20 (m, 1H, ArH), 5.31 (q, *J* = 6.4 Hz, 1H, CH), 2.2 (br s, 1H, OH), 1.51 (d, *J* = 6.4 Hz, 3H, CH3) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 143.0 (C*ipso*), 131.6 (C*ipso*), 129.4 (ArCH), 128.3 (ArCH), 127.2 (ArCH), 126.4 (ArCH), 66.9 (CH), 23.5 (CH3) ppm.

**1-(2,4-Difluorophenyl)ethan-1-ol (4g)**. This product has been previously reported.[43] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 7.55-7.45 (m, 1H, ArH), 6.95-6.85 (m, 1H, ArH), 6.85-6.75 (m, 1H, ArH), 5.25-5.10 (m, 1H, CH), 1.52 (d, J = 6.5 Hz, 3H, CH<sub>3</sub>) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl<sub>3</sub>): δ 162.1 (dd, *J* = 247.8, 12.2 Hz), 159.6 (dd, *J* = 248.1, 11.9 Hz), 128.6 (dd, *J* = 13.7, 3.8 Hz), 127.5 (dd, *J* = 9.5, 6.5 Hz), 111.2 (dd, *J* = 20.9, 3.5 Hz), 103.6 (t, *J* = 25.4 Hz), 63.9 (d, *J* = 2.5 Hz), 24.0 ppm. <sup>19</sup>F-NMR (CDCl3, 282.37 MHz): δ -112.1 (d, *J* = 6.6 Hz), -116.1 (d, *J* = 7.1 Hz) ppm.

**2,2,2-Trifluoro-1-(4-fluorophenyl)ethan-1-ol** (**4h**): This product has been previously reported.[44] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 7.55–7.45 (m, 2H, ArH), 7.15–7.10 (m, 2H, ArH), 5.15–5.00 (m, 1H, CH), 2.78 (br s, 1H, OH) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 163.4 (d, <sup>1</sup>*J*C-F = 248.4 Hz), 129.8 (C*ipso*), 129.3 (d, <sup>3</sup>*J*C-F = 8.3 Hz), 124.1 (q, <sup>1</sup>*J*C-F = 282.2 Hz), 115.6 (d, <sup>2</sup>*J*C-F = 21.5 Hz), 72.1 (q, <sup>2</sup>*J*C-F = 32.1 Hz) ppm. <sup>19</sup>F-NMR (CDCl3, 282.5 MHz): δ -78.6 (d, *J* = 6.1 Hz, 3F), -111.9 (m, 1F) ppm.

**1-(Pyridin-2-yl)ethan-1-ol (4i)**. This product has been previously reported.[45] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 8.55 (d, *J* = 4.8 Hz, 1H, ArH), 7.75–7.65 (m, 1H, ArH), 7.30 (d, *J* = 8.1 Hz, 1H, ArH), 7.30–7.20 (m, 1H, ArH), 4.91 (q, *J* = 6.5 Hz, 1H, CH), 4.35 (br s, 1H, OH), 1.52 (d, *J* = 6.5 Hz, 3H, CH3) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 163.0 (C*ipso*), 148.1 (ArCH), 136.8 (ArCH), 122.2 (ArCH), 119.8 (ArCH), 68.8 (CH), 24.25 (CH3) ppm.

**Diphenylmethanol (4j):** This product has been previously reported.[41] <sup>1</sup>H NMR (400 MHz, CDCl3): δ 7.45-7.25 (m, 10 H, Ar-H), 5.87 (s, 1 H, CH), 2.52 (s, 1 H, OH) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 143.8 (C*ipso*), 128.5 (ArCH), 127.6 (ArCH), 126.5 (ArCH), 76.2 (CH) ppm.

**(***E***)-1,3-Diphenylprop-2-en-1-ol (4k):** This product has been previously reported.[46]<sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 7.55–7.20 (m, 10H, ArH), 6.73 (d, *J* = 15.9 Hz, 1H, C*H*-Ph), 6.42 (dd, *J* = 15.9, 6.5 Hz, 1H, C*H*=CH-Ph), 5.42 (d, *J* = 6.5 Hz, 1H, CH-OH), 2.19 (br s, 1H, OH) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 124.7 (C*ipso*), 136.5 (C*ipso*), 131.5, 130.5, 128.6 (ArCH), 128.5 (ArCH), 127.76, 127.74 (ArCH), 126.6 (ArCH), 126.3, 75.1 (CH) ppm.

**Butan-2-ol (4m)**. This product has been previously reported.[47] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 3.75–3.70 (m, 1H, CH), 1.67 (br s, 1H, OH), 1.50–1.45 (m, 2H, CH2), 1.18 (d, *J* = 6.2 Hz, 3H, C*H*3-CH), 0.93 (t, *J* = 7.5 Hz, 3H, C*H*3-CH2) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 69.4  $(CH)$ , 32.0  $(CH<sub>2</sub>)$ , 22.8  $(CH<sub>3</sub>)$ , 9.9  $(CH<sub>3</sub>)$  ppm.

**Cyclohexanol (4n):** This product has been previously reported.[48,49] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 3.70–3.55 (m, 1H, CH), 2.05 (br s, 1H, OH), 2.00–1.85 (m, 2H, CH2), 1.80–1.65 (m, 2H, CH2), 1.60–1.45 (m, 2H, CH2), 1.40–1.20 (m, 4H, CH<sup>2</sup> x 2) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl<sub>3</sub>): δ 70.3 (CH), 35.5 (CH<sub>2</sub>), 25.4 (CH<sub>2</sub>), 24.1 (CH<sub>2</sub>) ppm.

**1,2,3,4-Tetrahydronaphthalen-1-ol (4o):** This product has been previously reported.[41] <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 7.50–7.40 (m, 1 H, ArH), 7.30–7.05 (m, 3 H, ArH), 4.85–4.75 (m, 1 H, CH-OH), 2.90–2.70 (m, 2 H, CH<sub>2</sub>), 2.10–1.70 (m, 5 H, CH<sub>2</sub> x 2, CH) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 138.8 (C*ipso*), 137.1 (C*ipso*), 129.0 (ArCH), 128.7 (ArCH), 127.6 (ArCH), 126.2 (ArCH), 68.1 (CH), 32.3 (CH2), 29.2 (CH2), 18.8 (CH2) ppm.

**(1***r***,3***r***,5***r***,7***r***)-Adamantan-2-ol (4p):** This product has been previously reported. <sup>1</sup>H NMR (300.13 MHz, CDCl3): δ 3.95-3.85 (m, 1H, CH), 2.50–1.50 (m, 15H) ppm. <sup>13</sup>C NMR (75.48 MHz, CDCl3): δ 74.5 (CH), 37.5 (CH2), 36.2 (CH2), 34.5 (CH), 31.0 (CH2), 27.5 (CH), 27.0 (CH) ppm.

## **A6.9 Catalyst recyclability**

**Recyclability of the catalyst in the cyanosilylation reaction:** In a 1 mL vial with a septum screw capped equipped with a stirring bar, the catalysts **Y/Eu-MOF** (6.4 mg, 0.5 mol%) were weighed. Then, the corresponding amount of acetophenone (56 µL, 0.5 mmol) followed by TMSCN (68 µL, 0.55 mmol, 1.1 equiv.) were added and the reaction was stirred under inert  $N_2$ atmosphere at room temperature overnight. After this time, 1.5 mL of DCM was added to the reaction mixture and transferred to a centrifuge tube for the separation of the catalyst. The centrifugation of the mixture was carried out at 12300 rpm during 5 min. After that, the solution was eliminated, and the catalyst was washed with DCM (2 x 1.5 mL). Later on, the catalyst was dried under vacuum and reused in the next cycle of the reaction with the same reaction conditions previously described.



**Scheme A6.1.** Reaction conditions used for the study of recyclability of Y/Eu-MOF catalysts in the cyanosilylation reaction.

**Recyclability of the catalyst in the hydroboration reaction:** In a 1 mL vial with a septum screw capped equipped with a stirring bar, the catalysts **Y/Eu-MOF** (9.2 mg, 0.5 mol%) were weighed. Then, the corresponding amount of acetophenone (84 µL, 0.75 mmol) followed by HBPin (120 µL, 0.825 mmol, 1.1 equiv.) were added and the reaction was stirred under inert N<sub>2</sub> atmosphere at room temperature overnight. After this time, 1.5 mL of DCM was added to the reaction mixture and transferred to a centrifuge tube for the separation of the catalyst. The centrifugation of the mixture was carried out at 12300 rpm during 5 min. After that, the solution was eliminated, and the catalyst was washed with DCM (2 x 1.5 mL). Later on, the catalyst was dried under vacuum and reused in the next cycle of the reaction with the same reaction conditions previously described.



**Scheme A6.2.** Reaction conditions used for the study of recyclability of Y/Eu-MOF catalysts in the hydroboration reaction.

## **A6.10 Leaching test**

**Leaching test**: after the first and second reaction of the recyclability test was complete in the hydroboration reaction, the reaction was centrifuged and the supernatant was filtered through a plug of celite and dried under vacuum. Later, 1-(pyridin-2-yl)ethan-1-one (86 µL, 0.75 mmol) and HBPin (120 μL, 0.825 mmol, 1.1 equiv.) were added to the crude of the corresponding reaction cycle and the reaction was stirred under inert N2 atmosphere at room temperature during 24 h. After that time, an aliquot was analysed by <sup>1</sup>H NMR obtaining in 38 % and 13 % of product **4j**, corroborating that the leaching of Y or Eu take place.



**Scheme A6.3**. Leaching test carried out after the first and second cycle.



## **A6.11 TOF of 6.1Y-Eu**





**Figure A6.5.** Analysis of the TOF (h<sup>-1</sup>) obtained in the hydroboration reaction acetophenone at different times of reaction with **6.1Y-Eu** (0.5 mol%) with the optimized reaction conditions.

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