This is a pre-copyedited, author-produced version of an article accepted for publication in J Antimicrob Chemother following peer review. The version of record Lloréns-Villar Y, Tusell F, Canut A, Barrasa H, Corral E, Martín A, Rodríguez-Gascón A. Antibiotic susceptibility trend before and after long-term use of selective digestive decontamination: a 16 year ecological study. J Antimicrob Chemother. 2019 Aug 1;74(8):2289-2294. doi: 10.1093/jac/dkz186. PMID: 31065685 is available online at: https://academic.oup.com/jac/article/74/8/2289/5486502?login=false

Antibiotic susceptibility trend before and after long-term use of selective digestive decontamination: a 16-year ecological study

Y. Lloréns-Villar¹, F. Tusell², A. Canut³, H. Barrasa⁴, E. Corral⁴, A. Martín⁴, A. Rodríguez-Gascón^{5,*}

¹ Servicio de Farmacia Hospitalaria. Hospital Universitario de Álava. C/ Olaguibel, 27. 01004 Vitoria-Gasteiz, Spain

² Departamento de Economía Aplicada III. Universidad del País Vasco UPV/EHU. Facultad de Ciencias Económicas y Empresariales. Avda. Lehendakari Aguirre nº 83. 48015 Bilbao, Spain.

³ Servicio de Microbiología. Hospital Universitario de Álava. C/ Francisco Leandro de Viana 1. 01009

⁴ Servicio de Medicina Intensiva. Hospital Universitario de Álava. C/ Olaguibel, 27. 01004 Vitoria-Gasteiz, Spain

⁵ Farmacia y Tecnología Farmacéutica. Facultad de Farmacia. Centro de investigación Lascaray ikergunea, Universidad del País Vasco UPV/EHU. Paseo de la Universidad nº 7. 01006 Vitoria, Spain

^{*} corresponding author: Alicia Rodríguez-Gascón

E-mail: alicia.rodriguez@ehu.eus

Phone: +34 945013094

Fax: +34 945 013040

Running title: Antibiotic susceptibility before and after long-term use of SDD

Synopsis

Objectives: To compare the antimicrobial susceptibility rate in an Spanish ICU before and after the introduction of selective digestive decontamination (SDD), and also to compare with susceptibility data from other Spanish ICUs without SDD.

Methods: We performed a retrospective study in the ICU of the University Hospital of Alava, where SDD was implemented in 2002. SDD protocol consisted of a 2% mixture of gentamycin, colistin and amphotericin B applied on the buccal mucosa, and a suspension of the same drugs in the gastrointestinal; additionally, for the first 3 days, systemic ceftriaxone was administered. From 1998 to 2013 we analyzed the antimicrobial susceptibility rates in 48 antimicrobial/organism combinations. Interrupted time series using a linear dynamic model with the SDD as an intervention was used. Data from other ICUs were obtained from the ENVIN-HELICS national registry.

Results: Only *E. coli* and *P. mirabilis* against amoxicillin/clavulanate, and *E. faecalis* against high concentration of gentamycin, resulted in a significant decrease of the susceptibility rate after the implementation of the SDD, with a drop of 20%, 27% and 32%, respectively. Compared to other Spanish ICUs without SDD, susceptibility rate was higher in the ICU of our hospital in most cases. When it was lower, differences were less than 10%, except for *E. faecium* against high concentration of streptomycin, in which the difference was 19%.

Conclusions: No relevant changes in the overall susceptibility rate after the implantation of the SDD was detected. Susceptibility rates were not lower than the media in the Spanish ICUs without SDD.

Introduction

Selective digestive decontamination (SDD) consists of the topical application of nonabsorbable antimicrobial agents in the oropharynx and gut,¹ and has been associated with better patient outcome.^{2,3} Although high-quality evidence supports the use of SDD, it is not widely used in clinical practice,⁴ The predominant concern seems to be the development of antimicrobial-resistant pathogens. However, in a meta-analysis,⁵ no relation between the use of SDD and the development of antimicrobial resistance was detected. Several reasons justify the limited evidence of the lack of the SDD effect on antimicrobial resistance, including relatively few studies, most of which assessed the effects of SDD at the patient (treated with SDD) level rather than at ICU (ecologic) level, and with limited follow-up time. In addition, there are few studies assessing long-term effects of SDD on multidrug-resistant bacteria in settings of high rates of resistance.⁶ The aim of the present work was to evaluate the antimicrobial susceptibility rates before and after the introduction of the SDD in an ICU of Spain, a country with high level of resistance. A second objective was to compare the susceptibility rates with those reported in other Spanish ICUs without SDD.

Methods

The study was performed in a ICU of the University Hospital of Alava (Spain), a 800-bed tertiary care teaching facility. This 13-bed ICU cares for surgical, neurosurgical and medical patients. SDD consisted of a 2% mixture gentamycin, colistin and amphotericin B applied on the buccal mucosa, and a suspension of the same drugs (respective doses of 80, 100 and 500 mg) provided in the gastrointestinal tract at 6-h intervals. In addition, for the first 3 days, intravenous ceftriaxone (2 g a day) was administered to all SDD patients. SDD was implanted in 2002 (third quarter), and there were no relevant changes

either in the occupancy rates or in the characteristics of the patients admitted in the ICU before and after the implantation of the SDD.

Bacterial isolates and antimicrobial susceptibility

From 1998 to 2013 we analyzed the bacterial susceptibility rate in 48 antimicrobial/organism combinations (Tables S1 to S11). We also calculated the rate of resistant microorganism acquisition, expressed per 100 patients-day. We included isolates taken for clinical or surveillance purposes, and only the first isolate per patient.

Data of antimicrobial susceptibility from the ENVIN-HELICS national registry⁷ were obtained (report of 2014, corresponding to susceptibility data of 2013). It includes a total of 22,064 patients and 192 ICUs. Less than 5% of these ICUs have implanted the SDD; therefore, these data were used as control (susceptibility rates in absence SDD).

Antibiotic use

Antibacterial consumption, expressed as DDD/100 patient-days, was calculated according to the 2014 version of the ATC/DDD classification.⁸ Prophylactic and therapeutic medication were not distinguished.

Statistical analysis

We developed a statistical model of interrupted time series in which the introduction of the SDD was considered as the intervention. The susceptibility rate λ_t was assumed to be linearly as follows:

 $\log \lambda_t = x_{1t} + x_{2t} (DDD/100 \text{ patient-days})_t + x_{3t} SDD_t$

 $\mathbf{x}_t = \mathbf{T}\mathbf{x}_{t-1} + \mathbf{w}_t$

where T is a fixed matrix and w_t a vector of random noise. The components of the vector are the (unknown) trend of the susceptibility rate of the organism against a given

antimicrobial, x_{1t} , as well as the unknown coefficient x_{2t} of the variable DDDs/100 patientdays, and the coefficient x_{3t} of a 0-1 step variable SDDt designed to capture the effect of SDD.

The total number of isolates (whose distribution given N_t is assumed Poisson N_t λ_t) determines how much each observation in the data set influences the susceptibility rate. To apply this model, the statistical package R version 3.3.1 and in particular package FAS⁹ was used. A significant level of 5% was considered.

On the other hand, trends in the number of isolates, in the rate of resistant microorganism acquisition, and in the antimicrobial consumption were analyzed in the pre- and post-SDD period with linear correlation. An appropriate degree of fit was considered when the correlation coefficient was ≥ 0.7 .¹⁰ A p value < 0.05 was considered statistically significant. IBM[®] SPSS[®] Statistics v. 24 was used.

Results

Out all the combinations studied, only the difference in the susceptibility rate before and after the SDD of *E. coli* and *P. mirabilis* against amoxicillin/clavulanic acid, and *E. faecalis* against high concentration of gentamycin turned out to be significant (p<0.05), the susceptibility rate being lower in the post-SDD period (Table S12). Figure 1 shows the predicted and observed values for these three combinations. The change in the susceptibility of *E. coli* and *P. mirabilis* against amoxicillin/clavulananic acid before and after the implantation of the SDD was 20% and 27%, respectively. In the case of *E. faecalis* against high concentration of gentamycin, the reduction of the susceptibility was 32%. The difference in the susceptibility rate of *E. coli* and *P. mirabilis* against amoxicillin/clavulanic acid before and after the implantation of all beta-lactam consumption in the statistical model (Table

S13). The difference in the susceptibility rate of *E. coli* was also significant with the introduction in the model of the consumption of piperacillin-tazobactam.

For most microorganisms (Figure S1), the number of isolates was constant over the period of study. Data of antibiotic consumption are presenting in Figure S2 and Table S14. No trend in the consumption of all beta-lactam and all antibiotics was detected either in the pre-SDD or in the post-SDD. No increase in the rate of resistant microorganism acquisition was detected in the post-SDD period (Table S15).

Table 1 shows the susceptibility rate of all antibiotic-organism combinations in 2013 (mean value of the four trimesters) and those from the ENVIN-HELICS report for 2013. In most cases, the susceptibility rate is higher in the ICU of our hospital than those reported for the Spanish ICUs.

Discussion

In this ecological study, we detected evidence of decreasing susceptibility trends after the introduction of the SDD only for *E. coli* and *P. mirabilis* against amoxicillin/clavulanic acid, and *E. faecalis* to high concentration of gentamycin. It is widely assumed that SDD facilitates the selection of enterococci and transmission of hospital-adapted strains,^{11,12} and this may justify the decrease in the susceptibility of *E. faecalis* against high level gentamycin; in fact, a dominant and epidemic *E. faecalis* clone (ST6) was recently detected in the ICU of our hospital, which was related to the use of SDD.¹³ Over the last years, an increase in the rate of resistance to amoxicillin/clavulanic acid among *E. coli* isolates in Europe¹⁴ has been noted, including Spain¹⁵, which has been related to an increase in the consumption of this antibiotic. Therefore, the decrease in the susceptibility of *E.coli* against amoxicillin/clavulanic acid may be explained by the increase in the use of this antibiotic in the community, and not only by the introduction of the SDD. Since the overall consumption of beta-lactams in our ICU was lower in the post-

SDD period, the decrease in the susceptibility of *E. coli* cannot be explained by the use of beta-lactams. The reduction of the susceptibility of *P. mirabilis* to amoxicillin/clavulanic acid in the post-SDD period should be taken with caution, since the level of significance was lower but very close to 0.05, and because the number of isolates was very low (Table S4). It is important to take into account that beta-lactam resistance of *P. mirabilis* has increased in recent years in several countries.¹⁶

For the rest of microorganism-antibiotic combinations, changes in the susceptibility after the introduction of the SDD were not detected. Additionally, no trends in rate of resistant microorganism acquisition in the post-SDD period were found, which confirms that SDD does not contribute to the spread of resistant isolates. It is important to take into account that, in the post-SDD period, other interventions that may have conditioned the susceptibility rates were introduced, such as the International Standard ISO 9001:2000 guidance in 2005, the Bacteremia Zero program in 2009¹⁷, and the "Zero-ventilatorassociated pneumonia (VAP)" bundle in 2011¹⁸.

Contrary to that reported in other studies,⁶ no trends in the number of isolates in the post-SDD period were detected. The overall use of antibiotics in our ICU was lower in the post-SDD period, and it did not increase over time. These results are in agreement with Daneman,⁵ which postulates that the use of prophylactic selective decontamination could even lead to reductions in the need for therapeutic antimicrobials.

We also compared the susceptibility rates of our ICU with those reported in the ENVIN-HELICS national registry (most of the ICUs without SDD).⁷ The susceptibility rate in our hospital was of the same order or even higher that the media in the Spanish ICUs. Only the susceptibility rate of *E. faec*ium against high concentration of streptomycin was lower in our hospital (50% versus 69%).

In conclusion, our study reveals no relevant changes in the overall susceptibility rate after the implantation of the SDD. Additionally, susceptibility rates were not lower than

the media in the Spanish ICUs without SDD. One important shortcoming of our study is that it is retrospective in nature and based on a single institution.

Funding

This work was supported by the University of the Basque Country UPV/EHU (GIU17/32,

PPG17/65).

Transparency declarations

None to declare.

References

- 1. Plantinga NL, de Smet AMGA, Oostdijk EAN *et al.* Selective digestive and oropharyngeal decontamination in medical and surgical ICU patients: individual patient data meta-analysis. *Clin Microbiol Infect* 2018; **24**: 505-13.
- 2. Dombrowski SU, Prior ME, Duncan E *et al.* Clinical components and associated behavioural aspects of a complex healthcare intervention: multi-methods study of selective decontamination of the digestive tract in critical care. *Aust Crit Care* 2013; **26**: 173-9.
- 3. Silvestri L, van Saene HKF, Bion J. Antipathy against SDD is justified: No. *Intensive Care Med*. 2018; **44**: 1169-73.
- 4. Reis Miranda D, Citerio G, Perner A *et al*. Use of selective digestive tract decontamination in European intensive cares: the ifs and whys. *Minerva Anestesiol* 2015; **81**: 734-42.
- Daneman N, Sarwar S, Fowler RA *et al.* Effect of selective decontamination on antimicrobial resistance in intensive care units: a systematic review and meta-analysis. *Lancet Infect Dis* 2013; **13**: 328-41.
- Sánchez-Ramírez C, Hípola-Escalada S, Cabrera-Santana M, *et al.* Long-term use of selective digestive decontamination in an ICU highly endemic for bacterial resistance. *Crit Care* 2018; 22: 141.
- ENVIN-HELICS. Sociedad Española de Medicina Intensiva, Crítica y Unidades Coronarias (SEMICYUC). Grupo de Trabajo de Enfermedades Infecciosas. Estudio Nacional de Vigilancia de Infección Nosocomial en UCI. Report of 2014. http://hws.vhebron.net/envinhelics.
- 8. WHO Collaborating Centre for Drug Statistics Methodology, https://www.whocc.no/ddd/definition_and_general_considera/.
- Helske J. Exponential family state space models in R. *Journal of Statistical Software* 2017; 78: 1-39.
- 10. Friedrich LV, White RL, Bosso JA. Impact of use of multiple antimicrobial on changes in susceptibility of gram negative aerobes. Clin Infect Dis 1999; **28**: 1017-24.
- 11. van der Bij AK, Frentz D, Bonten MJ *et al*. Gram-positive cocci in Dutch ICUs with and without selective decontamination of the oropharyngeal and digestive tract: a retrospective database analysis. *J Antimicrob Chemother* 2016;**71**:816-20.
- 12. Zarrilli R, Tripodi MF, Di Popolo A *et al.* Molecular epidemiology of high-level aminoglycosideresistant enterococci isolated from patients in a university hospital in southern Italy. *J Antimicrob Chemother* 2005; **56**: 827-35.
- Muruzábal-Lecumberri I, Girbau C, Canut A *et al.* Spread of an *Enterococcus faecalis* sequence type 6 (CC2) clone in patients undergoing selective decontamination of the digestive tract. *APMIS* 2015; **123**: 245-51.
- 14. Rodríguez-Baño J, Oteo J, Ortega A *et al.* Epidemiological and clinical complexity of amoxicillin-clavulanate-resistant *Escherichia coli. J Clin Microbiol* 2013; **51**: 2414-7.
- 15. Ortega A, Oteo J, Aranzamendi-Zaldumbide M *et al.* Spanish multicenter study of the epidemiology and mechanisms of amoxicillin-clavulanate resistance in *Escherichia coli*. *Antimicrob Agents Chemother* 2012; **56**: 3576-81.

- 16. Aragón LM, Mirelis B, Miró E *et al.* Increase in beta-lactam-resistant Proteus mirabilis strains due to CTX-M- and CMY-type as well as new VEB- and inhibitor-resistant TEM-type beta-lactamases. *J Antimicrob Chemother* 2008; **61**: 1029-32.
- 17. Palomar M, Álvarez-Lerma F, Riera A *et al*. Impact of a national multimodal intervention to prevent catheter-related bloodstream infection in the ICU: the Spanish experience. *Crit Care Med* 2013; **41**: 2364-72.
- Álvarez Lerma F, Sánchez García M, Lorente L *et al*. Guidelines for the prevention of ventilator-associated pneumonia and their implementation. The Spanish "Zero-VAP" bundle. *Med Intensiva* 2014;**38**:226-36.
- 19. Kuch A, Willems RJ, Werner G *et al.* Insight into antimicrobial susceptibility and population structure of contemporary human *Enterococcus faecalis* isolates from Europe. *J Antimicrob Chemother* 2012;**67**:551-8.
- 20. García-Vázquez E, Albendín H, Hernández-Torres A *et al*. Risk factors associated to highlevel resistance to aminoglycosides. *Rev Esp Quimioter* 2013;**26**:203-13.

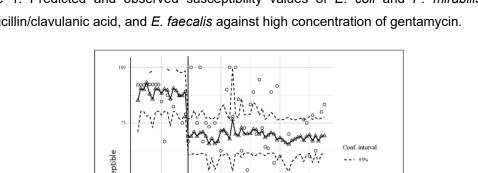
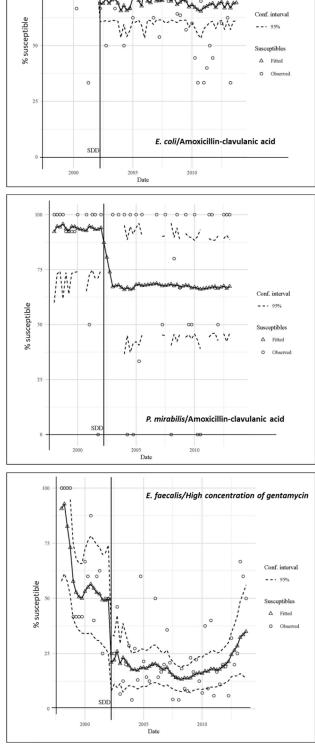


Figure 1. Predicted and observed susceptibility values of E. coli and P. mirabilis against amoxicillin/clavulanic acid, and *E. faecalis* against high concentration of gentamycin.



Microorganism	Antimicrobial agent	Susceptibili	eptibility rate in 2013 (%)	
Gram-negative organisms				
Enterobacterales		HUA [*]	ENVIN-HELICS	
Enterobacter spp.	Cefotaxime	80	50	
	Imipenem	100	89	
E. coli	Amoxicillin/clavulanicacid	64	66	
	Cefepime	88	85	
	Cefotaxime	77	83	
	Ceftazidime	88	84	
	Ciprofloxacin	59	67	
	Gentamycin	78	85	
	Levofloxacin	87	77	
	Piperacillin/tazobactam	93	85	
Klebsiella pneumoniae	Cefotaxime	100	62	
	Ceftazidime	83	60	
P. mirabilis	Amoxicillin/clavulanicacid	100	76	
	Cefepime	100	94	
	Cefotaxime	100	98	
	Ceftazidime	100	100	
	Ciprofloxacin	100	82	
	Gentamycin	100	82	
	Levofloxacin	100	100	
	Piperacillin/tazobactam	100	100	
<i>Serratia</i> spp.	Cefepime	100	100	
	Cefotaxime	100	77	
	Ceftazidime	100	85	
	Ciprofloxacin	100	78	
	Gentamycin	100	95	
	Levofloxacin	100	91	
	Piperacillin/tazobactam	100	80	
Non-fermenters		100	00	
Acinetobacter spp	Imipenem	0	15	
P. aeruginosa	Amikacin	86	82	
	Cefepime	88	61	
	Ceftazidime	88	67	
	Ciprofloxacin	61	47	
	Colistin	90	94	
	Imipenem	67	49	
	Levofloxacin	67	41	
	Meropenem	67	51	
	•			
	Piperacillin/tazobactam	67	66	
	Tobramycin Gram-positive organisms	56	n.a.	
E. faecalis	High concentration of gentamycin (500 mg/L)	51	44 ¹	
E. laecans	High concentration of streptomycin (1,000 mg/L)	57	44 42 ¹	
		100	42 98	
	Vancomycin Daptomycin	96	97	
	Linezolid	90 96	97	
E foodium	Tigecycline High concentration of gentamycin (500 mg/L)	- 53	- 54 ²	
E. faecium			54 69 ²	
	High concentration of streptomycin (1,000 mg/L)	50		
	Vancomycin Daptomycin	100 83	100	
		83 92	90	
			97	
S. aureus	Tigecycline	-	-	
	Cloxacillin	92	100	
	Vancomycin	100	97	
	Daptomycin	100	100	
	Linezolid	100	100	
0.10	Tigecycline	92	100	
CoNS	Cloxacillin	12	15	
	Vancomycin	100	97	
	Daptomycin	96	100	
	Linezolid	96	95	
	Tigecycline	92	100	

Table 1.Compared values of susceptibility rate between the ICU of the HUA and data in the ENVIN-HELICS report.7

*HUA: mean value of the four quarters

n.a.: not available

¹Not available in the ENVIN-HELICS report. Obtained from Kuch*et al*¹⁹. 2012. Data from 2006-2009 ²Not available in the ENVIN-HELICS report. Obtained from García-Vázquez *et al*²⁰.