

Hydrogen production by steam reforming of bio-oil/bio-ethanol mixtures in a continuous thermal-catalytic process

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Abstract

The feasibility of the steam reforming of bio-oil aqueous fraction and bio-ethanol mixtures has been studied in a continuous process with two in-line steps: thermal step at 300 °C (for the controlled deposition of pyrolytic lignin during the heating of the bio-oil/bio-ethanol feed) followed by steam reforming in a fluidized bed reactor on a Ni/ α -Al₂O₃ catalyst. The effect of bio-ethanol content in the feed has been analyzed in both the thermal and reforming steps, and the suitable range of operating conditions (temperature and space-time) has been determined for obtaining a high and steady hydrogen yield. Higher ethanol content in the mixture feed improves the reaction indices and reduces coke deposition. Operating conditions of 700 °C and space-times higher than 0.23 g_{catalyst} h (g_{bio-oil+EtOH})⁻¹ are suitable for attaining almost fully conversion of oxygenates (bio-oil and ethanol) and hydrogen yields above 93 %, with low catalyst deactivation.

Keywords: bio-oil, bioethanol, hydrogen production, steam reforming, fluidized reactor

1. Introduction

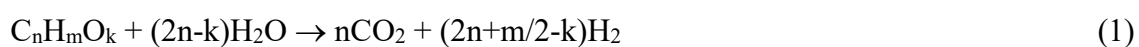
A biomass-based plant (bio-refinery) is the best solution to combine and integrate various processes for converting plant-based biomass to chemicals, energy and materials in order to maximize economic and environmental benefits, while minimizing waste and pollution [1]. Sultana and Kumar [2] found that the delivery cost of a feedstock that combines woody biomass and agricultural biomass is lower than that for a single type of biomass. Consequently, the joint valorization of oxygenated compounds derived from different types of biomasses, such as bio-oil and bio-ethanol (e.g. by catalytic steam reforming for obtaining H₂), is an interesting route for the development of the bio-refinery concept. Bio-oil is produced by fast pyrolysis of woody biomass, such as mill and harvest residues [3], and bio-ethanol can be sustainably obtained by hydrolysis/fermentation of agricultural lignocellulosic biomass [4,5].

The bio-oil is a complex mixture of water and oxygenated compounds (acids, alcohols, ketones, phenols, furans, etc.), whose composition depends on the biomass source and operating conditions of pyrolysis [6]. The viability of bio-oil reforming process is curtailed by the problems associated with re-polymerization of certain bio-oil components (i.e., derivatives of the lignin contained in biomass) that affect reactor operation and cause catalyst deactivation. These problems are mitigated with different strategies, such as the valorization of the bio-oil aqueous fraction, the previous separation of pyrolytic lignin and the co-feeding of methanol [7].

The lignocellulosic ethanol has a great potential as a chemical building block for bio-refineries [8] and its catalytic steam reforming avoids the costly dehydration steps required for other valorization strategies. Furthermore, its production cost has been significantly reduced due to advances in the conversion techniques [9].

Besides, bio-ethanol can replace the methanol commonly used to stabilize the bio-oil during its storage [10], which increases the interest of reforming this mixture.

The stoichiometry of the overall reactions for bio-oil and ethanol steam reforming are given by Eqs. (1) and (2), respectively:



In practice, the hydrogen yield is lower than the stoichiometric maximum due to undesired secondary reactions (thermal decomposition of oxygenates, methanation, reverse-WGS reaction, ethanol dehydration or dehydrogenation).

Nickel-based catalysts have been widely used in the literature for reforming the aqueous fraction of bio-oil [11,12] and ethanol [13-15] due to the high C-C bond-breaking activity and the relatively low cost. Among oxide supports, Al₂O₃-based supports are often used as reforming catalysts because of their mechanical and chemical resistance [16].

This paper analyzes the feasibility of the steam reforming of bio-oil aqueous fraction and bio-ethanol mixtures on Ni/ α -Al₂O₃ catalyst in a continuous process with two steps in line (thermal and catalytic) (Figure 1). This two-step system minimizes the problems inherent to the bio-oil catalytic valorization, caused by the deposition of carbonaceous solid (pyrolytic lignin) during feed preheating [17]. The effect bio-ethanol in the feed has on both the thermal and the reforming steps is analyzed and the suitable range of operating conditions (temperature and space-time) is determined for obtaining steady and high hydrogen yield.

Figure 1

2. Experimental

2.1. Preparation and characterization of bio-oil/bio-ethanol mixtures

Bio-oil was obtained by flash pyrolysis of pine sawdust in a semi-industrial demonstration plant located in Ikerlan-IK4 technology center (Alava, Spain), with a biomass feeding capacity of 25 kg/h [18]. The aqueous fraction (82 wt% water) was obtained by phase separation after adding water to the raw bio-oil in a water/bio-oil mass ratio = 2/1 [19]. For every 100 g of raw bio-oil (composed of 65 g of oxygenated compounds and 35 g water) two fractions are obtained: 15 g of organic fraction and 285 g of aqueous fraction. The bio-oil/bio-ethanol mixtures were prepared by adding aqueous ethanol (82 wt% water) to the bio-oil aqueous fraction in mass ratios ranging from 80/20 up to 20/80. In this paper the mixtures are denoted as B_{*x*}/E_{*y*}, where *x* is the percentage (wt%) of bio-oil aqueous fraction (B) and *y* is the percentage (wt%) of aqueous ethanol (E) in the mixture.

The composition of the raw bio-oil, the bio-oil aqueous fraction and the bio-oil/bio-ethanol mixture B₅₀/E₅₀ (Table 1, on a dry basis) was determined by GC/MS analyzer (*Shimadzu QP2010S device*). Bio-ethanol addition to the bio-oil aqueous fraction only contributes to molecular dilution of the bio-oil oxygenates. The reactivity reported between light alcohols

and organic compounds [20] is not observed probably because the mixture is highly diluted in water (82 wt%). The difference between the ethanol content in the mixture B₅₀/E₅₀ (52 wt%) and the nominal content (50 wt%) is caused by cumulative errors in the mixture preparation (a large quantity to conduct all experiments) and by the chromatographic analysis (the compounds with highest molecular weight are not detected).

Table 1

The elemental composition (C, H, O) of the raw bio-oil, bio-oil aqueous fraction and pyrolytic lignin deposited in the thermal step was determined by elemental analysis (*Leco CHN-932 analyzer* and ultra-microbalance *Sartorius M2P*). The resulting molecular formulas (on a dry basis) are C_{4.3}H_{7.2}O_{2.6} and C_{4.1}H_{7.4}O_{2.7}, for the raw bio-oil and bio-oil aqueous fraction, respectively. The water content of (bio-oil aqueous fraction)/bio-ethanol mixtures (82 wt%) was quantified by Karl Fischer titration in a *870 KF Titrino Plus* device.

2.2. Catalyst

The α -Al₂O₃ support was obtained by calcining hydrated γ -Al₂O₃ (150-250 μ m particle size) at 1100 °C for 5 h. After the subsequent impregnation with Ni(NO₃)₂·6H₂O and drying at 110 °C for 24 h, the final calcination was carried out at 700 °C for 3 h. The nominal Ni content of the Ni/ α -Al₂O₃ catalyst is 10 wt%. Although it has been found that conversion of oxygenates in the steam reforming increases with Ni content, a limited value of this content (10-15 wt%) is the best option, given that this low content minimizes Ni sintering problems at high temperature [21].

The catalyst was sieved (150-250 μ m) and mixed with an inert solid (CSi carborundum, with 30-50 μ m particle size) in a catalyst/inert mass ratio of 1/4, for improving fluid-dynamic properties of the catalytic bed and reducing axial dispersion. The CSi is a suitable material as diluent in fluidized bed reactors due to its high mechanical resistance and good thermal conductivity [22]. The particle sizes of both materials (of different density) were established in order to have isodromic particles (with similar minimum fluidization velocity). They were determined by means of a fluid-dynamic study (analyzing the evolution of pressure drop with gas linear velocity in a glass reactor at atmospheric pressure and room temperature). Furthermore, it was found that the mixture of both solids within the bed is uniform under the reaction conditions. The catalyst was reduced in the

reactor prior to each reforming reaction, under a H₂-He flow (5 vol % of H₂) for 2 h at 700 °C.

The catalyst properties have been previously described in detail [23]. The physical properties, such as BET surface area (65.5 m²/g), pore volume (0.174 cm³/g) and average pore size (7.4 nm) have been determined from the N₂ adsorption-desorption isotherms, obtained by using a *Micromeritics ASAP 2010C* analyzer. This device was also used for hydrogen chemisorption measurements for quantifying nickel dispersion and specific metal surface area, with values being 2.1 % and 14.1 m²/g_{metal}, respectively. Nickel content (8.93 wt%) was measured by inductively coupled plasma and atomic emission spectroscopy (ICP-AES). The X-ray diffraction (XRD) pattern measured on a *Bruker D8 Advance* diffractometer with a CuK_{α1} radiation showed diffraction lines corresponding to the reflection of Al₂O₃ phase and Ni⁰ phase (at 2θ angles of 44.5°, 51.9° and 76.4°).

The coke deposited on the catalyst after each steam reforming reaction was quantified by temperature programmed oxidation (TPO), conducted in a *TA Instruments Q5000 IR* thermobalance coupled to a mass *Thermostar Balzers Instrument* spectrometer for monitoring the signal corresponding to CO₂ (mass 44).

2.3. Reaction system and operating conditions

The two-step reaction equipment has been previously described in detail for the steam reforming of aqueous bio-oil [23]. The first reactor (thermal processing of the feed) retains the carbonaceous solid (pyrolytic lignin) formed by re-polymerization of certain bio-oil oxygenated components. The volatile compounds leaving this thermal step are subsequently transformed (by catalytic steam reforming) in the second unit (fluidized bed reactor). The controlled deposition of pyrolytic lignin in a specific thermal step prior to the catalytic reactor minimizes the operating problems caused by this deposition and attenuates catalyst deactivation. This fact was previously verified for the catalytic conversion of raw bio-oil into hydrocarbons [24,25]. Besides, the use of a fluidized bed reactor favors a uniform gasification of the coke precursors adsorbed on the catalyst, thereby attenuating the catalyst deactivation [26].

The analysis of the reforming products was carried out on-line with a gas chromatograph (*Agilent Micro GC 3000*) provided with four modules for analysing the following: (1) permanent gases (O₂, H₂, CO, and CH₄) with 5A molecular sieve capillary column; (2) light oxygenates (C₂-), CO₂ and water, with Plot Q capillary column; (3) C₂-C₄

hydrocarbons, with alumina capillary column; (4) oxygenated compounds (C₂₊) with Stabilwax type column.

The flow-rate of (bio-oil aqueous fraction)/bio-ethanol mixtures was 0.1 ml min⁻¹ and was controlled by an injection pump *Harvard Apparatus 22*. The operating conditions were as follows: thermal step, 300 °C; catalytic steam reforming, 500-800 °C; steam-to-carbon ratio at the fluidized bed reactor inlet (S/C), 5.8-10; space-time, 0.10-0.45 g_{catalyst} h (g_{bio-oil+EtOH})⁻¹ (water not included); gas hourly space velocity (G_{C1}HSV), 8200-41500 h⁻¹ (calculated at the reaction temperature as CH₄ equivalent units) and time over frequency (TOF), 6.5-32.3 s⁻¹.

The particle size of the catalyst (150-250 μm) and the linear velocity of the gas under the reaction conditions (around 3.5 cm s⁻¹) are suitable for minimizing the limitations of internal and external diffusion, respectively. This aspect was proven by ensuring that the results were independent of the feed flow-rate for the same space-time. Furthermore, the α-Al₂O₃ support has a mesoporous structure that facilitates the diffusion of bio-oil compounds (with different molecular weights) within the pores. Consequently, the conditions set are suitable for fluidization and for reducing diffusional limitations.

2.4. Reaction indices

The bio-oil and bio-ethanol conversion are calculated by Eq. (4), considering their molar flow-rates (as C units contained) at the inlet, F_{j,inlet}, and outlet (un-reacted bio-oil or ethanol), F_j, of the catalytic reactor:

$$X_j = \frac{F_{j,inlet} - F_j}{F_{j,inlet}} \times 100 \quad (4)$$

The molar flow-rate of the bio-oil fed into the two-unit system is calculated from its mass flow-rate and empirical formula (C_{4.1}H_{7.4}O_{2.7}). The molar flow-rate of the bio-oil at the catalytic reactor inlet (F_{bio-oil,inlet}) is determined from a mass balance in each run by considering the amount of PL deposited in the thermal unit and its elemental composition (Figure 2).

The molar flow-rate of the bio-oil at the reactor outlet (F_{bio-oil}) is calculated from the total mole number at the reactor outlet (quantified by a mass balance to the catalytic reactor) and the molar fraction of oxygenates (excluding ethanol), analyzed by μGC and GC/MS.

The hydrogen yield is calculated by Eq. (5) as a percentage of the maximum allowed by stoichiometry, considering the contribution of both reactants (bio-oil and bio-ethanol), according to Eqs. (1) and (2).

$$Y_{H_2} = \frac{F_{H_2}}{((2 + m/(2n) - k/n) F_{\text{bio-oil,inlet}} + 3F_{\text{EtOH,inlet}})} \times 100 \quad (5)$$

where F_{H_2} is the molar flow of H_2 obtained.

The yield of each carbon-containing byproduct (CO_2 , CO , CH_4 and C_2 - C_4 light hydrocarbons) is quantified by Eq. (6), based on the total C fed into the catalytic reactor (taking into account both the bio-oil aqueous fraction and the bio-ethanol):

$$Y_i = \frac{F_i}{(F_{\text{bio-oil,inlet}} + F_{\text{EtOH,inlet}})} \times 100 \quad (6)$$

where F_i is the molar flow of each i product.

The selectivity of each product is calculated by Eq. (7):

$$S_i = \frac{F_i}{\sum F_i} \times 100 \quad (7)$$

where $\sum F_i$ is the sum of gaseous products (excluding oxygenated compounds and water) at the reactor outlet.

The experimental errors determined by repeated experiments are $\pm 3\%$ in the H_2 yield values and $\pm 1\%$ in those of C-containing products.

The following sections deal with the features of the steam reforming of bio-oil aqueous fraction and bio-ethanol mixtures by analyzing the effect bio-ethanol content in the feed and operating conditions (reforming temperature and space-time) have on the hydrogen yield and catalyst deactivation.

3. Results and discussion

3.1. Effect of co-feeding ethanol on pyrolytic lignin deposition in the thermal step

The effect bio-ethanol content of the mixture fed into the two-unit system has on the deposition of pyrolytic lignin (in the first thermal unit) was analyzed. Accordingly, the pyrolytic lignin yield (PL yield) formed by re-polymerization of the bio-oil fed was

quantified using Eq. (3). The bio-ethanol is not considered for this calculation because it does not undergo re-polymerization.

$$\text{PL yield(wt\%)} = \frac{\text{pyrolytic lignin deposited (g)}}{\text{oxygenates in bio - oil fed (g)}} \times 100 \quad (3)$$

The PL yield and its elemental composition for different bio-ethanol contents in the mixture fed are shown in Figure 2. These results reveal that the addition of aqueous ethanol (bio-ethanol) to the bio-oil aqueous fraction slightly attenuates the re-polymerization of bio-oil oxygenates and so pyrolytic lignin deposition in the thermal unit. Besides, an increase in bio-ethanol content does not significantly affect pyrolytic lignin composition (only a slight increase in O content and a decrease in C content is observed). These results suggest that the slight attenuation of pyrolytic lignin deposition is a consequence of molecular dilution, which changes the microstructure of the bio-oil emulsion.

The attenuation of lignin deposition observed for these mixtures is lower than that previously observed for raw bio-oil/methanol mixtures [27]. This is probably due to the removal of the fraction containing high molecular weight oxygenates from the raw bio-oil for obtaining the bio-oil aqueous fraction used in this work. This suggests that the stabilizing effect of the alcohol (ethanol or methanol) is more significant when it is added to the raw bio-oil, given that it is a more unstable emulsion than the bio-oil aqueous fraction.

Figure 2

3.2. Catalytic steam reforming of (bio-oil aqueous fraction)/bio-ethanol mixtures

3.2.1. Effect of feed composition

Fig. 3 shows the results of evolution with time on stream of bio-oil and ethanol conversion and the yield of H₂, CO and CH₄ in the steam reforming of bio-oil aqueous fraction (B₁₀₀, Fig. 3a), a 50 wt% mixture of (bio-oil aqueous fraction)/bio-ethanol (B₅₀/E₅₀, Fig. 3b) and bio-ethanol (E₁₀₀, Fig. 3c). Based on these runs the possible differences between the joint reforming of bio-oil and bio-ethanol and their individual reforming are analyzed. The reforming conditions were as follows: 700 °C; space-time, 0.1 g_{catalyst} h (g_{bio-oil+EtOH})⁻¹; G_{C1HSV}, 41500 h⁻¹; Q_{feed}, 0.1 ml (min)⁻¹.

Figure 3

It is observed that bio-oil conversion at zero time on stream (fresh catalyst) is almost full for B₁₀₀ (Fig. 3a) and B₅₀/E₅₀ (Fig. 3b) feeds and slightly decreases with time on stream. Bio-ethanol conversion remains full and steady for 5 h reaction for B₅₀/E₅₀ (Fig. 3b) and E₁₀₀ (Fig. 3c) feeds. Besides, B₁₀₀ and E₁₀₀ feeds yield 84 % and 91 % H₂, respectively, at zero time on stream, whereas B₅₀/E₅₀ feed has a halfway behavior between the individual feeds, with H₂ yield being 89 %. The CO yield obtained from the reforming of B₁₀₀ (≈ 10 %) is lower than that from E₁₀₀ (≈ 22 %) and an intermediate yield (19 %) is obtained for B₅₀/E₅₀ feed. The initial yield of CH₄ is low and very similar for the three feeds studied (≈ 2 %).

These results at zero time on stream evidence the different reaction mechanism for the steam reforming of bio-oil and ethanol. A reaction pathway has been suggested for ethanol steam reforming which includes as follows [28,29]: 1) ethanol dehydrogenation to acetaldehyde, 2) acetaldehyde decomposition to produce CO + CH₄ and 3) WGS reaction and CH₄ reforming to produce hydrogen. According to this, the higher production of CO at zero time on stream obtained in the ethanol reforming may be a consequence of acetaldehyde decomposition and the reforming reaction of the CH₄ produced in this decomposition.

H₂ and CO₂ yields decrease and that of CO increases with time on stream for all the feeds in Fig. 3, while the conversion of bio-oil and ethanol decreases and that of CH₄ increases in a less pronounced way with time on stream. This fact reveals that the WGS reaction is affected more severely by the catalyst deactivation than oxygenates reforming and decomposition reactions. The decrease in H₂ and CO₂ yield and the increase in CO yield with time on stream are more significant for B₁₀₀ feed (Fig. 3a) than for E₁₀₀ feed (Fig. 3c), whereas B₅₀/E₅₀ feed behavior is halfway between those for the individual feeds (Fig. 3b). Catalyst deactivation also affects CH₄ reforming, whose yield increases with time on stream, with this effect being more noticeable for B₁₀₀ feed (from 1 % up to 9 %) than for E₁₀₀ (from 2 % up to 4 %). These results reveal that the catalyst undergoes lower deactivation by feeding ethanol.

The catalyst deactivation in bio-ethanol reforming is caused by Ni site blockage by coke, whose formation is enhanced by the presence in the reaction medium of byproducts, such as ethylene (following oligomerization), methane (dehydrogenation) and (CO Boudouard reaction) [30]. Remiro et al. [26] have proposed a mechanism for coke formation in bio-oil reforming, identifying the steps of coke formation from by-products. They also concluded

that the coke formation capability increases as the reforming activity of the catalyst decreases. For both feeds catalyst deactivation by coke is attenuated under conditions enhancing the reforming/gasification of coke precursors and the formation of filamentous coke instead of encapsulating coke.

In order to analyze in more detail the effect of bio-ethanol content in the feed, the steam reforming of B_X/E_Y mixtures with different mass ratios was carried out (under the same operating conditions as in Fig. 3). Table 2 shows the results obtained for bio-oil and ethanol conversion (X_i), H_2 yield in the reforming unit (Y_{H_2}) and selectivity of reaction products (S_i) at zero time on stream and after 5 h reaction, and the coke content deposited on the catalyst after each reaction (C_c). The overall H_2 yield (Y_{H_2})^G by mass unit of the bio-oil and bio-ethanol fed into the two-step system was also estimated.

Table 2

Bio-ethanol conversion is independent of the feed composition and is almost full throughout 5 h reaction, whereas bio-oil conversion decreases by 10 % and 2 % after 5 h reaction for B_{100} and B_{20}/E_{80} feeds, respectively (Table 2). The initial H_2 yield increases significantly when 20 wt% bio-ethanol is added to the bio-oil aqueous fraction (from 84.4 % to 88.2 % for B_{80}/E_{20} feed), with this increase being attenuated for higher contents of bio-ethanol (91 % for E_{100} feed). Besides, the decrease in H_2 yield after 5 h time on stream (caused by catalyst deactivation) steadily attenuates by increasing the amount of ethanol in the feed, thus H_2 yield decreasing to 35 % for B_{100} feed and to 79 % for B_{20}/E_{80} . Moreover, the difference observed between the H_2 yield obtained in the reforming reactor (Y_{H_2}) and the overall H_2 yield in the two-step system, (Y_{H_2})^G, is lower as the bio-ethanol content in the feed is higher. This should be attributed to the lower deposition of pyrolytic lignin in the thermal step (due to the lower content of bio-oil in the mixture).

The evolution of hydrogen selectivity (S_{H_2}) with the bio-ethanol content in the feed follows a similar trend to the H_2 yield: the initial value is higher as bio-ethanol content is increased (although less pronounced than for yield) and then goes on to decrease with time on stream in a less pronounced way as the bio-ethanol content increases.

The initial selectivity of CO increases steadily with the bio-ethanol content in the feed and that of CO_2 decreases. The initial selectivity of CH_4 is less affected by the bio-ethanol/bio-oil ratio in the feed. The selectivity of CO and CH_4 increases with time on stream due to catalyst deactivation affecting mainly the WGS and CH_4 reforming reactions. This increase

(and thus deactivation) is considerably attenuated by increasing the ethanol content in the feed. This result suggests a higher contribution of bio-oil oxygenate compounds than bio-ethanol to the catalyst deactivation by coke deposition. The results of coke content deposited for the different feeds (Table 2) confirm this hypothesis.

The results in Fig. 3 and Table 2 reveal that bio-ethanol addition has a favorable effect on the bio-oil aqueous fraction reforming because it provides stability by attenuating catalyst deactivation by coke and enhancing a high and steady hydrogen production. Moreover, Table 2 data show that the presence of ethanol in the reaction medium has a beneficial synergistic effect of attenuating coke deposition. Thus, the coke content obtained in the reforming of the B₅₀/E₅₀ mixture is 1.3 wt%, which is lower than the theoretical content (2.15 wt%) that would correspond to the contribution of all components in the feed. This result is caused by a change in coke precursors concentrations in the reaction medium.

Furthermore, it should be considered that the limited internal diffusion of the bio-oil heaviest compounds may contribute to a lower conversion and to a different distribution of the reaction products obtained from the different feeds, which will be more significant at high temperature. The relevance of support diffusivity on the diffusional limitations in ethanol reforming has been reported in the literature (by the Weisz-Prater criterion). This limitation is significant above 350 °C for a Ni/MgAl₂O₄ catalyst [31] but for a Ni/Al₂O₃ catalyst (similar to that used in this paper) there is no diffusional limitation [21] due to the diffusing capacity within the Al₂O₃ support. However, this paper studies the joint reforming of ethanol and the oxygenated mixture constituting the bio-oil, so that the diffusional limitation is presumably heterogeneous and may affect the bio-oil compounds of high molecular weight.

3.2.2. Effect of reforming temperature

The runs were performed in the 500-800 °C range, at low space-time (0.1 g_{catalyst} h (g_{bio-oil+EtOH})⁻¹) and by feeding the mixture of 50 wt% bio-oil aqueous fraction and bio-ethanol (B₅₀/E₅₀). For this feed, 23.7 wt% of pyrolytic lignin (PL) is retained in the thermal step, whose elemental composition is 65.3 wt% C, 4.9 wt% H and 29.8 wt% O (Figure 2). Consequently, the composition of the treated bio-oil that enters the catalytic reactor is C_{3.7}H_{8.2}O_{3.0} and thereby, 2.3 mol of H₂ can be obtained from each mol of C in the bio-oil. The bio-ethanol contained in the feed can be converted to 3 mol of H₂ for every mol of C.

Fig. 4 shows the results at zero time on stream (fresh catalyst, continuous lines) and after 5 h reaction (dashed lines) of bio-oil and bio-ethanol conversion (Fig. 4a), H₂ yield and selectivity (Fig. 4b), CO and CO₂ yields (Fig. 4c) and CH₄ yield (Fig. 4d). HC yield (mainly ethylene and ethane) is not shown because is lower than 1 % in all cases.

Figure 4

Fig. 4a shows that ethanol conversion increases significantly with temperature, being almost full at 700 °C, whereas the increase in bio-oil conversion with temperature is less pronounced and temperatures above 800 °C are required for achieving full conversion of bio-oil with this low value of space-time. The yields of H₂ (Fig. 4b) and CO₂ (Fig. 4c) at zero time on stream follow similar trends with temperature, with both yields increasing significantly from 500 to 600 °C and going through a maximum at 700 °C. The slight decrease in these yields above 700 °C is caused by the reverse-WGS reaction, which is thermodynamically favored at higher temperature, as evidenced by the progressive increase in the CO yield with temperature (Fig. 4c). The low CO yield is a consequence of the catalyst activity for the WGS reaction. The low initial yield of CH₄ ($\approx 2\%$) (Fig. 4d) is a consequence of: i) a more effective competition of oxygenate catalytic reforming reactions than thermal decomposition reactions and ii) the effective CH₄ reforming at the temperature range studied.

The conversion of both bio-oil aqueous fraction and bio-ethanol decrease after 5 h time on stream due to catalyst deactivation, which is notably attenuated by increasing temperature (Fig.4a). The deactivation is severe at 600 °C, so that bio-oil and ethanol conversion decreases by 40 % and 22 %, respectively. The yields of H₂ (Fig. 4b) and CO₂ (Fig. 4c) also undergo significant reduction with time on stream, whereas the yields of CO and CH₄ remain almost constant. At 700 °C and 800 °C, the decrease in bio-oil and ethanol conversion after 5 h time on stream is very low. However, the decrease in H₂ and CO₂ yields is noticeable at both temperatures. It should be noted that the values for H₂ and CO₂ yield after 5 h reaction show a peak at 700 °C. Moreover, the values of H₂ selectivity after 5 h reaction progressively decrease with temperature (Fig. 4b) due to the increase in the yield of CO (Fig. 4c) and CH₄ (Fig. 4d), which is significant above 700 °C.

The aforementioned results evidence that deactivation selectively affects to the different steps of the reaction scheme. The decrease in bio-oil conversion and the fact that at 600 °C CO yield remains constant after 5 h reaction reveal that both the oxygenate reforming

reaction and the WGS reaction are similarly affected by deactivation at this relatively low temperature. Nevertheless, the increase in CO yield after 5 h reaction at 700 °C and 800 °C evidences that catalyst deactivation affects more severely WGS reaction than oxygenate reforming reactions at high temperatures. Deactivation significantly influences the reforming reaction of oxygenates at 800 °C, which favors thermal cracking reactions leading to CH₄ and CO. CH₄ reforming is also strongly affected by deactivation at high temperatures, which contributes to the great increase in CH₄ yield observed at 800 °C after 5 h time on stream (Fig. 4d). Furthermore, the thermodynamics of WGS reaction is hindered by increasing temperature.

The results of coke content deposited on the catalyst after 5 h reaction at the different temperatures are set out in Table 3. As observed, coke deposition on the catalyst (and thus catalyst deactivation) reduces as temperature is increased by enhancing the gasification reactions involving the coke deposited [26]. These results are consistent with those in Fig. 4, where a significant deactivation is observed at temperatures below 700 °C. The loss of catalyst activity observed at 800 °C is not due to coke deposition, which is negligible at this temperature, but to the sintering of the metallic function as proven in previous works [23,26]. It was found that the Ni crystal size of the catalyst used at 700 °C and regenerated by coke combustion is the same as that of the fresh catalyst (8 nm), thus confirming that the only cause of deactivation at this temperature is coke deposition. However, Ni sintering is remarkable at 800 °C and the Ni particle size of the catalyst used at this temperature is 15 nm (after removing the coke). Accordingly, 700 °C is a suitable temperature for minimizing deactivation and avoiding metal sintering, which allows the catalyst to be used continuously in reaction-regeneration cycles.

Table 3

Based on the above results, the optimum temperature for reforming mixtures of bio-oil and bio-ethanol is 700 °C, since at this temperature the maximum H₂ yield is attained with low deactivation of the Ni/ α -Al₂O₃ catalyst after 5 h reaction. This stability is promoted by coke gasification (which is low below 700 °C) and the low Ni metal sintering (which is significant above 700 °C).

3.2.3. Effect of space-time

The effect of space-time was analyzed at 700 °C in the 0.1-0.5 g_{catalyst} h (g_{bio-oil+EtOH})⁻¹ range, which corresponds to a gas hourly space velocity (G_{C1}HSV) between 8200 and

41500 h⁻¹ and TOF between 6.5 and 32.3 s⁻¹ feeding the B₅₀/E₅₀ mixture. Fig. 5 shows the results at zero time on stream (fresh catalyst, continuous lines) and after 5 h reaction (dashed lines) for bio-oil and ethanol conversion (Fig. 5a), H₂ yield and selectivity (Fig. 5b), CO₂ and CO yields (Fig. 5c) and CH₄ yield (Fig. 5d).

Figure 5

An increase in space-time enhances the reforming reactions of both bio-oil and ethanol, thus increasing their conversion (Fig. 5a) and also the WGS reaction extent, as derived from the progressive decrease in the initial values of CO yield (Fig. 5c). An increase in space-time to 0.23 g_{catalyst} h (g_{bio-oil+EtOH})⁻¹ results in a slight increase in the initial values of H₂ (Fig. 5b) and CO₂ (Fig. 5c) yields and enhancement of CH₄ reforming reaction (Fig. 5d). Nevertheless, a subsequent increase in space-time to 0.5 g_{catalyst} h (g_{bio-oil+EtOH})⁻¹ does not significantly improve conversion and H₂ yield.

The decrease with time on stream in oxygenate conversion (Fig. 5a) and H₂ yield (Fig. 5b) is noticeable at space-times below 0.23 g_{catalyst} h (g_{bio-oil+EtOH})⁻¹. For higher values of space-time, both oxygenate conversion and the H₂ yield hardly decrease in 5 h time on stream. CO yield increases from 19 % to 34 % for 0.1 g_{catalyst} h (g_{bio-oil+EtOH})⁻¹ (Fig. 5c) due to the deactivation of the WGS reaction. The increase in CH₄ yield with time on stream to 9 % for the lowest value of space-time studied is explained by the cracking reactions to give CH₄ and CO, which are favored by catalyst deactivation (lower activity for the reforming reactions). In addition, the catalyst deactivation affects CH₄ reforming reaction (Fig. 5d), which contributes to significantly decreasing H₂ yield (Fig. 5b).

These kinetic results are consistent with the coke contents deposited on the catalyst for the different values of space-time (Table 4), i.e., they significantly decrease as space-time is increased.

Table 4

Based on the above results under the operating condition studied, a space-time of 0.23 g_{catalyst} h (g_{bio-oil+EtOH})⁻¹ is enough for achieving high conversion of reactants in the (bio-oil aqueous fraction)/bio-ethanol mixture, with the maximum yield and selectivity of H₂ and low deactivation of the catalyst along 5 h reaction.

Furthermore, it should be noted that many of the afore-mentioned results correspond to severe deactivation conditions, and therefore they are useful to assess clearly the effect of operating conditions and bio-ethanol co-feeding on the deactivation and also to delimit the

range of conditions suitable for minimizing catalyst deactivation. The deactivation is also attenuated by using the proposed two-step reaction equipment, which facilitates the separation of compounds that repolymerize as pyrolytic lignin (in the first step), thus avoiding their deposition on the catalyst. Bearing in mind scaling up, the two-step process allows continuous operation, which may be performed without interruption by using a fluidized reactor with circulating catalyst. The partially deactivated catalyst would be fed back to the reactor after being regenerated in an external unit (for coke combustion). This technology is currently implemented for the conversion of methanol into olefins (MTO process) and has been proposed for the catalytic pyrolysis of biomass [32].

4. Conclusions

The joint steam reforming of the bio-oil aqueous fraction and bio-ethanol over a Ni/ α -Al₂O₃ catalyst is a promising route for obtaining H₂ from two different biomass-derived feedstocks. Bio-ethanol addition promotes the steam reforming of the bio-oil aqueous fraction by slightly attenuating the pyrolytic lignin deposition (during bio-oil heating) and stabilizing the steam reforming reaction by decreasing the catalyst deactivation by coke, given that the composition of reaction medium hinders coke formation. The reaction system with two in-line steps (thermal and catalytic) is suitable to avoid pyrolytic lignin deposition in the fluidized bed reforming reactor, and therefore longer reactions may be carried out with low catalyst deactivation under suitable reaction conditions.

Operating conditions of 700 °C and space-times higher than 0.23 g_{catalyst}h(g_{bio-oil+EtOH})⁻¹ are suitable for reforming the bio-oil aqueous fraction and bio-ethanol mixture (50 wt%), given that almost full oxygenate conversion and a high hydrogen yield (above 93 %) are attained with low catalyst deactivation. The catalyst used at 700 °C fully recovers its activity after being regenerated by coke combustion.

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Nomenclature

C_c	coke content, wt %
$F_{j,\text{inlet}}$	molar flow rate of j (aqueous bio-oil or ethanol) at the reactor inlet, in C equivalent units, mol h ⁻¹
F_j	molar flow rate of j (aqueous bio-oil or ethanol) at the reactor outlet, in C equivalent units, mol h ⁻¹
F_i	molar flow rate of i product (H ₂ , CO, CO ₂ , CH ₄ , C ₂ -C ₄) at the reactor outlet, mol h ⁻¹
$G_{C_1\text{HSV}}$	gas hourly space velocity, in CH ₄ equivalent units, h ⁻¹
PL	pyrolytic lignin deposited in the thermal unit, wt%
Q_{feed}	feed flow rate, ml min ⁻¹
S/C	steam-to-carbon ratio
S_i	selectivity of i product (H ₂ , CO, CO ₂ , CH ₄ , C ₂ -C ₄), %
X_j	conversion of j (bio-oil and bio-ethanol), %
Y_{H_2}	hydrogen yield in the catalytic reforming step, %
$(Y_{\text{H}_2})^G$	overall hydrogen yield in the two-unit system, %
Y_i	yield of i product (H ₂ , CO, CO ₂ , CH ₄ , C ₂ -C ₄), %
τ	space-time, g _{catalyst} h (g _{bio-oil+EtOH}) ⁻¹ (water not included)

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Figure Captions

- Figure 1.** Hydrogen production layout according to the joint steam reforming of bio-oil and bio-ethanol obtained from biomass.
- Figure 2.** Effect of ethanol content in the feed on the amount (wt%) and elemental composition of the pyrolytic lignin (PL) deposited in the thermal step at 300 °C.
- Figure 3.** Evolution with time on stream of oxygenate conversion and product yields. Reaction conditions: 700 °C; S/C, 10; space-time, 0.1 g_{catalyst} h (g_{bio-oil+EtOH})⁻¹; G_{C1}HSV, 41500 h⁻¹. Graph a: feed, B₁₀₀; Graph b: feed B₅₀/E₅₀; Graph c: feed, E₁₀₀.
- Figure 4.** Effect of temperature on the values at zero time on stream (fresh catalyst) and after 5 h reaction for bio-oil and ethanol conversions (a), H₂ yield and selectivity (b), CO and CO₂ yields (c), and CH₄ yield (d). Reaction conditions: feed, B₅₀/E₅₀; S/C, 7; space-time, 0.1 g_{catalyst} h (g_{bio-oil+EtOH})⁻¹; G_{C1}HSV, 41500 h⁻¹.
- Figure 5.** Effect of space-time on the values at zero time on stream (fresh catalyst) and after 5 h reaction for bio-oil and ethanol conversions (a), H₂ yield and selectivity (b), CO and CO₂ yields (b), and CH₄ yield (d). Reaction conditions: feed B₅₀/E₅₀; 700 °C; S/C, 7.

FIGURES

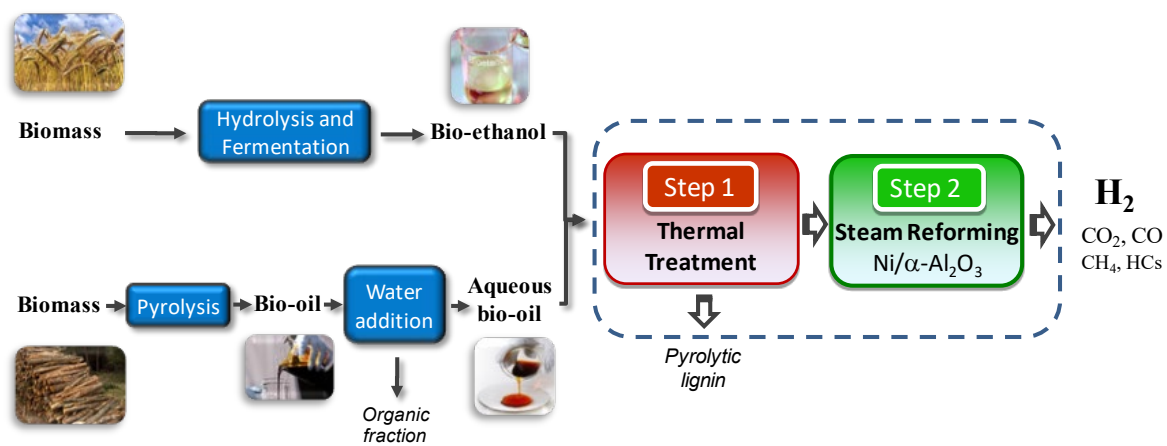


Figure 1

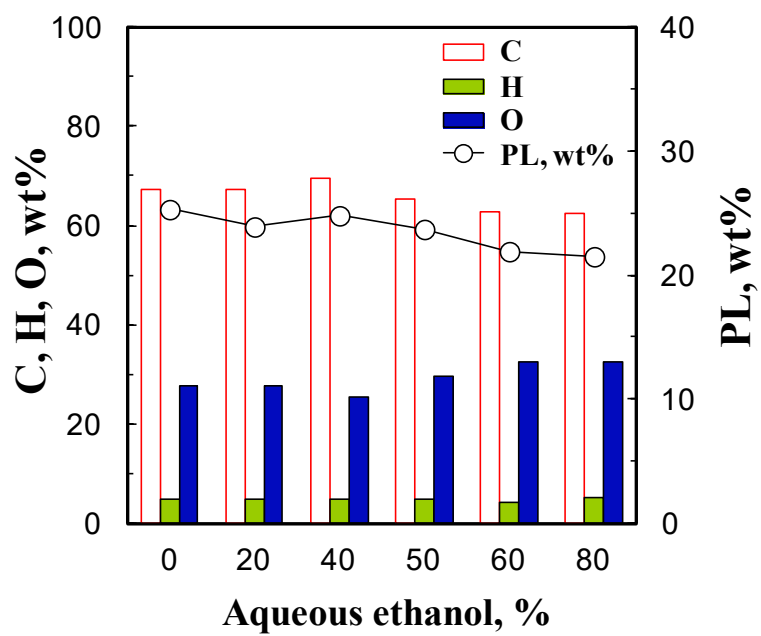


Figure 2

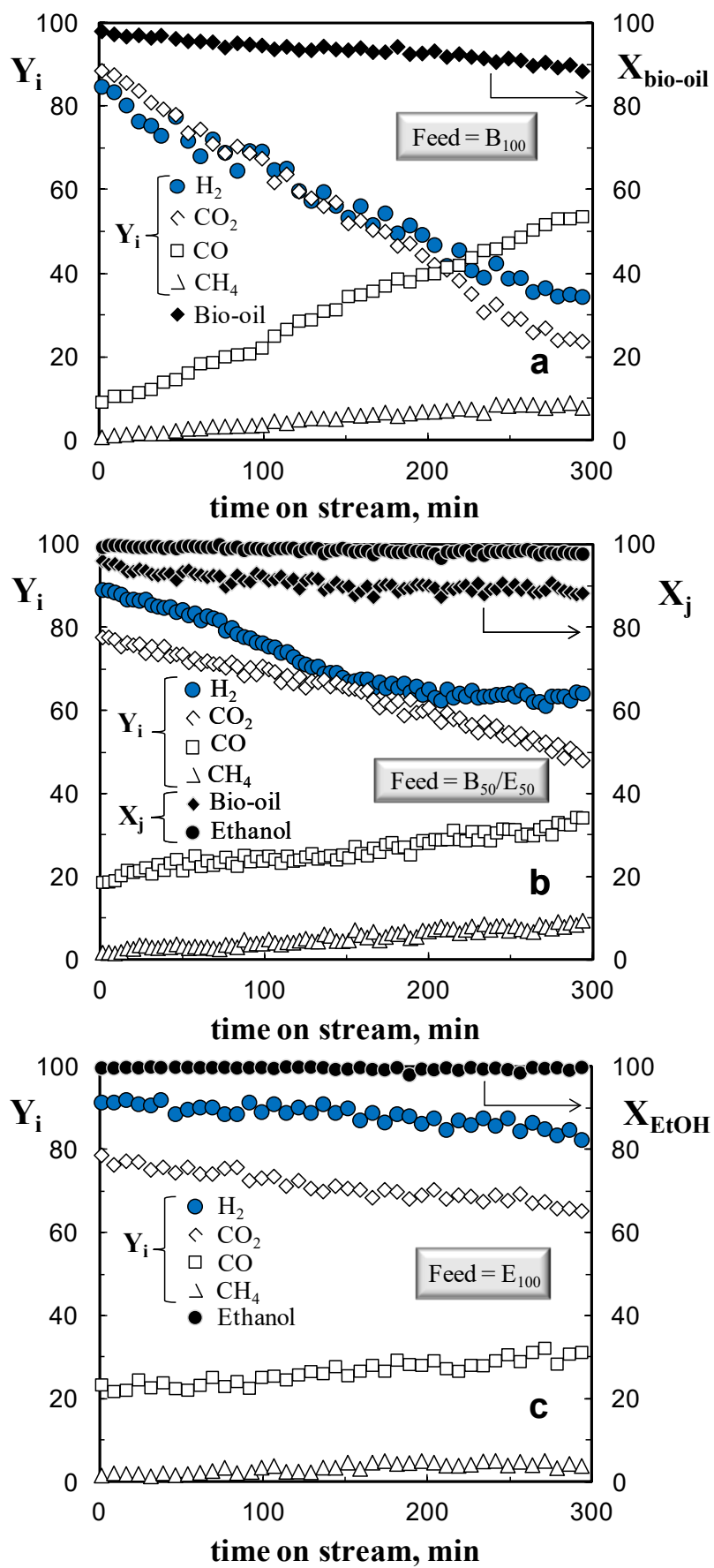


Figure 3

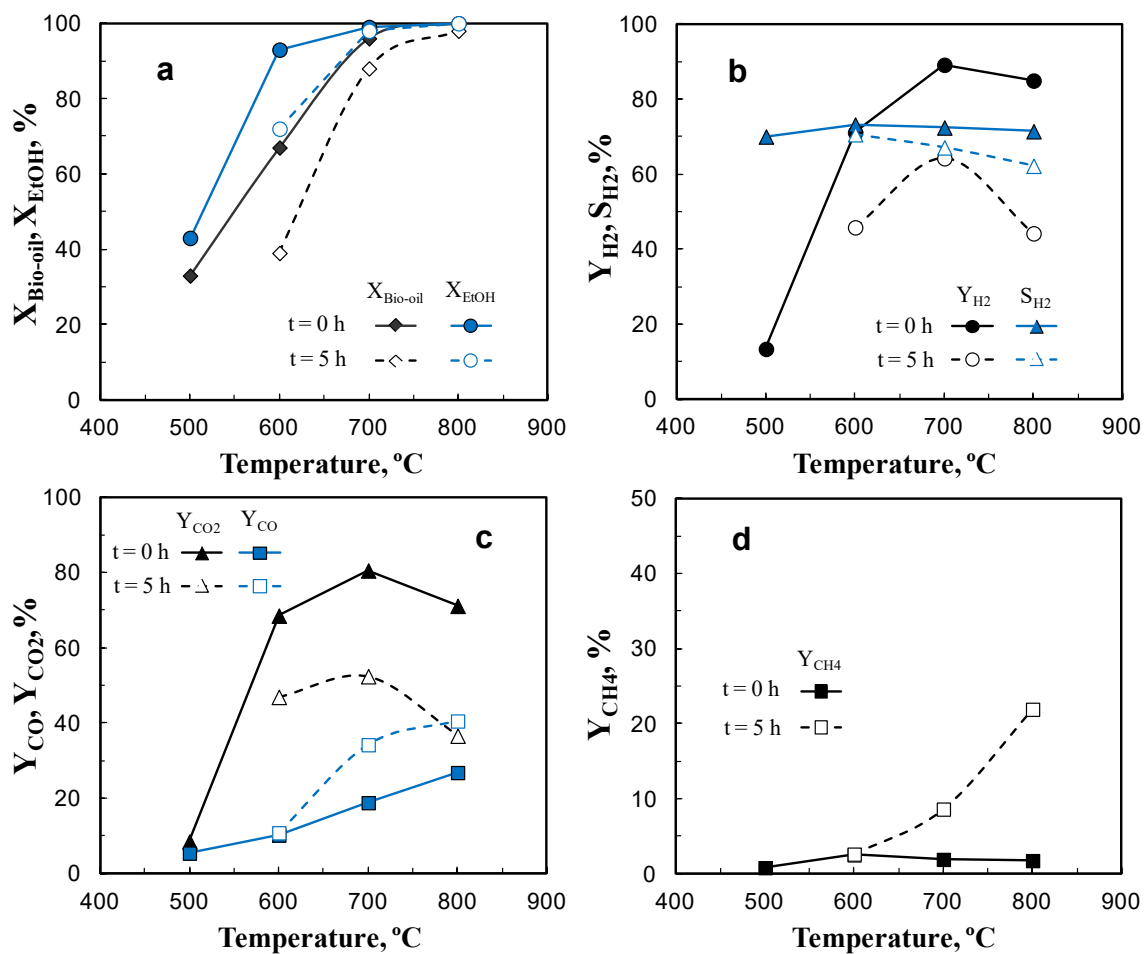


Figure 4

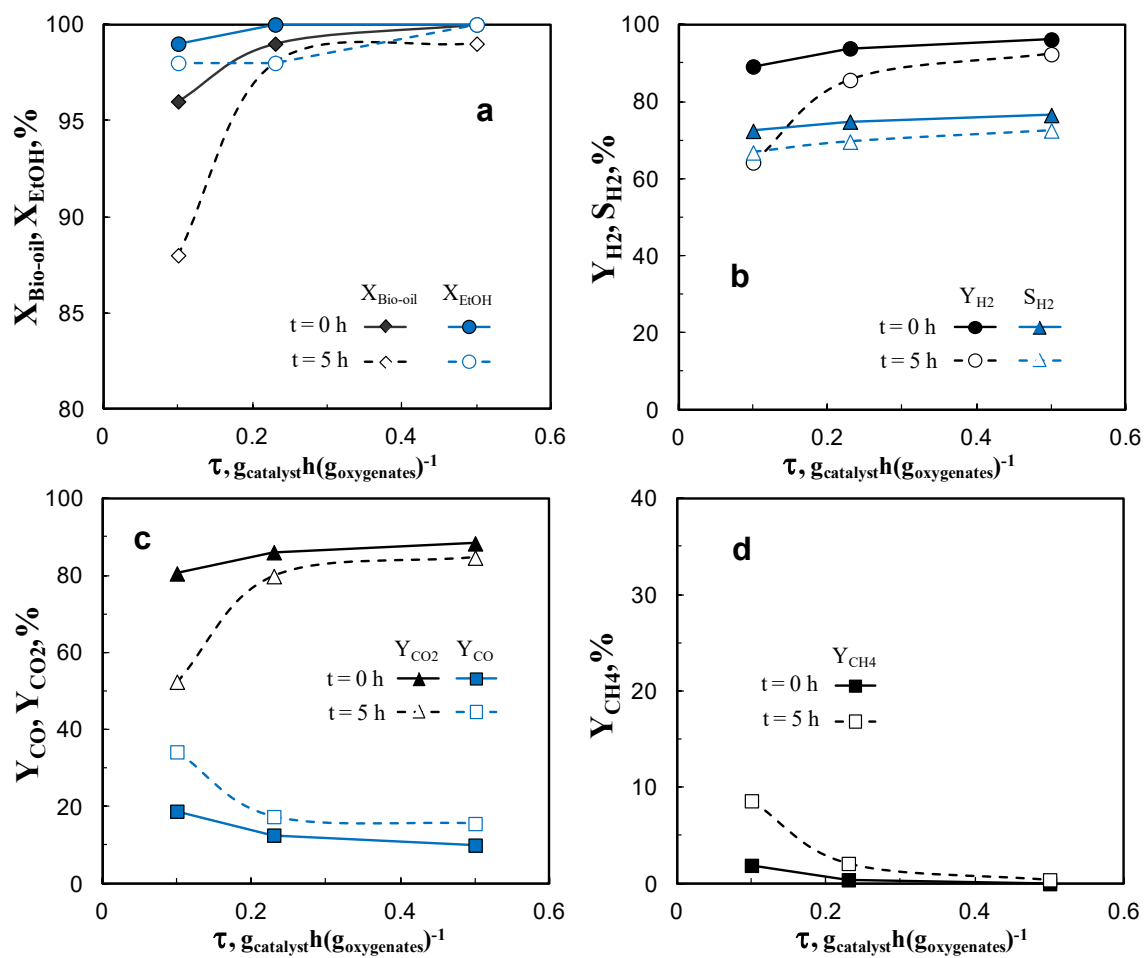


Figure 5

TABLES

Table 1. Composition (wt %) of raw bio-oil (rB), bio-oil aqueous fraction (B) and the mixture with 50 wt% of aqueous ethanol (B₅₀/E₅₀).

Compound/Group	rB	B	B₅₀/E₅₀
Ethanol	0.0	0.0	52.5
Acetic acid	12.8	19.1	9.2
Formic acid	2.1	2.7	1.1
Other acids	1.9	4.7	2.4
Hydroxyacetaldehyde	8.5	1.8	0.8
Others aldehydes	6.5	5.5	2.3
Acetone	5.5	1.0	0.5
1-hydroxy-2-propanone	16.3	8.7	3.9
Other ketones	3.8	8.1	4.6
Esters	5.1	3.1	1.3
Ethers	1.4	0.3	0.2
Alcohols	3.7	4.6	2.0
Phenols	16.6	13.4	6.1
Hexose	1.9	2.7	1.6
Levoglucofuranose	11.0	19.6	9.2
Others	0.4	1.1	0.7
Unidentified	2.5	3.6	1.6

Table 2. Effect of bio-oil/ethanol mass ratio on the values at zero time on stream (fresh catalyst) and after 5 h reaction of bio-oil conversion, ethanol conversion, H₂ yields (catalytic and overall) and reaction products selectivity. Ni/ α -Al₂O₃ catalyst, 700° C, S/C = 6-10, $\tau = 0.1 \text{ g}_{\text{catalyst}} \text{ h}(\text{g}_{\text{oxygenates}})^{-1}$, $G_{\text{C1HSV}} = 41500 \text{ h}^{-1}$.

Mixture	Bio-oil aqueous fraction/aqueous ethanol (wt%)							
	100/0	80/20	60/40	50/50	40/60	20/80	0/100	
	B ₁₀₀	B ₈₀ /E ₂₀	B ₆₀ /E ₄₀	B ₅₀ /E ₅₀	B ₄₀ /E ₆₀	B ₂₀ /E ₈₀	E ₁₀₀	
X _{Bio-oil}	0 h	0.98	0.98	0.96	0.96	0.97	0.99	-
	5 h	0.88	0.92	0.90	0.88	0.96	0.97	-
X _{EtOH}	0 h	-	1	1	1	1	1	1
	5 h	-	0.95	0.97	0.98	0.99	0.99	0.99
Y _{H₂} , %	0 h	84.4	88.2	88.9	89.1	89.8	89.2	91.0
	5 h	35.1	49.6	62.5	64.2	73.1	78.9	82.4
(Y _{H₂}) ^G , %	0 h	59.3	68.8	74.1	78.1	81.2	84.9	91.0
	5 h	19.0	38.7	52.1	56.4	66.1	75.4	82.4
S _{H₂} , %	0 h	68.8	71.6	72.1	72.4	73.0	73.1	72.2
	5 h	47.4	59.9	65.4	67.1	68.8	70.1	71.2
S _{CO₂} , %	0 h	26.2	23.7	22.8	21.8	20.9	20.0	19.1
	5 h	11.9	15.3	16.4	16.7	17.7	18.9	18.8
S _{CO} , %	0 h	4.7	4.5	4.8	5.2	5.9	6.7	8.0
	5 h	33.9	19.8	14.4	12.6	10.3	8.9	9
S _{CH₄} , %	0 h	0.3	0.2	0.3	0.4	0.2	0.2	0.7
	5 h	5.5	4.3	3.3	3.2	3.0	2.0	1.0
S _{H_Cs} , %	0 h	0	0	0	0	0	0	0
	5 h	1.3	0.7	0.5	0.4	0.2	0.1	0.1
C _c , wt%	3.8	2.9	1.7	1.3	1.1	0.9	0.5	

Table 3. Effect of reforming temperature on coke contents. Conditions: B_{50}/E_{50} , $S/C = 7$, $\tau = 0.1 \text{ g}_{\text{catalyst}} \text{ h}(\text{g}_{\text{bio-oil}+\text{EtOH}})^{-1}$.

<u>T, °C</u>	<u>C_c, wt %</u>
500	6.8
600	2.3
700	1.3
800	n.d.

Table 4. Effect of space-time (τ , $\text{g}_{\text{catalyst}}\text{h}(\text{g}_{\text{bio-oil}+\text{EtOH}})^{-1}$) on coke contents after 5 h of reaction. Conditions: B_{50}/E_{50} , $S/C = 7$, $T = 700$ °C.

τ	C_c , wt %
0.10	1.34
0.23	0.65
0.50	n.d.

Comentado [A.R.1]: Añadir columna con TOF