This is the accepted manuscript of the article that appeared in final form in **Ecological Engineering** 103(Part B) : 394-403 (2017), which has been published in final form at <u>https://doi.org/10.1016/j.ecoleng.2016.11.061</u>. © 2016 Elsevier under CC BY-NC-ND license (<u>http://</u>creativecommons.org/licenses/by-nc-nd/4.0/)

# 1 Does land use impact on groundwater invertebrate diversity and functionality in

## 2 alluvial wetlands?

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- 4 Cecilia Español<sup>1</sup>, Francisco A. Comín<sup>1</sup>, Belinda Gallardo<sup>1</sup>, Jingmei Yao<sup>2,3</sup>, José Luis
- 5 Yela<sup>4</sup>, Fabián Carranza<sup>1</sup>, Ane Zabaleta<sup>5</sup>, Julian Ladera<sup>4</sup>, Miren Martínez-Santos<sup>5</sup>,

6 Magali Gerino<sup>2,3</sup>, Sabine Sauvage<sup>2,3</sup>, and José Miguel Sánchez-Pérez<sup>2,3</sup>.

- 7
- 8 <sup>1</sup> Pyrenean Institute of Ecology (IPE-CSIC); Department of Biodiversity and
- 9 Restoration. Avda. Montañana 1005, 50059, Zaragoza, Spain.
- 10 <sup>2</sup> University of Toulouse; INPT, UPS; Laboratoire Ecologie Fonctionnelle et
- 11 Environnement (EcoLab), Avenue de l'Agrobiopole, 31326 Castanet Tolosan Cedex,

12 France.

- <sup>3</sup> CNRS, EcoLab, 31326 Castanet Tolosan Cedex, France.
- <sup>4</sup> University of Castilla-La Mancha (UCLM); School of Environmental Sciences and
- 16 Biochemistry, Avda Carlos III, 45071 Toledo, Spain.
- <sup>5</sup> University of the Basque Country UPV/EHU; Hydrogeology and Environment Group,
- 18 Department of Geodynamics, Sarriena s/n, 48940 Leioa (Basque Country), Spain.
- 19

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- 20 Corresponding author:
- 21 Cecilia Español
- 22 e-mail: cecilia.espanol@csic.es; cieles@msn.com
- 23 Current address: Pyrenean Institute of Ecology (IPE-CSIC), Campus de Aula Dei,
- 24 Avda. Montañana 1005, 50059, Zaragoza, Spain.
- 25 Tel. (+34) 976369393 Fax: (+34) 974363222

## 27 Abstract

28 Land use change, especially the transformation of riparian forest to agricultural fields, 29 plays an important role in groundwater quality. Yet little is known about the effects of 30 land use change on groundwater invertebrate assemblages and diversity. This study 31 assesses for the first time the effect of land use (agricultural vs. forest use) on the 32 groundwater invertebrate community of four river floodplains representative of variable 33 conditions across SW Europe (the Garonne River in France, and the Bidasoa, Ebro and 34 Tajo Rivers in Spain). Groundwater invertebrate and water samples were collected 35 quarterly in 8 to 12 piezometers located in each floodplain over a year. Taxonomic and 36 functional diversity indices and ecosystem services (i.e. biogeochemical filtration and 37 particulate organic matter breakdown) were calculated. The taxonomic and functional 38 diversity of groundwater invertebrate communities increased linearly with the 39 increasing surface occupied by riparian forests and decreased under intensified 40 agriculture use. Moreover, the provision of key ecosystem services related to the 41 biogeochemical filtration and particulate organic matter breakdown also increased 2-42 fold under more natural forest land. According to regression models, this is likely due to 43 the pressure of agricultural practices on groundwater quality, with high concentrations 44 of nitrates and sulphates exerting a negative influence over groundwater invertebrate 45 diversity and their associated ecosystem services. The results of this study have 46 important management implications, and suggest that the presence of large riparian 47 corridors enhances groundwater invertebrate diversity and functionality in floodplains 48 deeply disturbed by agricultural practices.

50	Highlights
51	• We study the effect of land use on groundwater invertebrate community.
52	• Groundwater invertebrate community change between land use types.
53	• Invertebrate diversity and ecosystem services increases with forest occupancy.
54	• Agriculture has a negative influence on groundwater invertebrate communities.
55	• Water quality controls groundwater invertebrate diversity and ecosystem
56	services.
57	
58	Keywords
59	Biogeochemical filtration; biological traits; ecosystem services; particulate organic
60	matter breakdown; floodplain; stygofauna.

## 62 **1. Introduction**

63 Subterranean habitats have been traditionally considered as a biological desert, only 64 interesting as a major source of water resources (Gibert et al., 1994). However, recent 65 studies have demonstrated that groundwater systems are home to a great variety of cryptic and endemic organisms (Avramov, 2014; Deharveng et al., 2009; Sket, 1999), 66 67 which provide important ecosystem services ranging from water purification and 68 bioremediation to water infiltration and transport (Boulton et al., 2008). Unfortunately, 69 the spread of groundwater habitats has also favoured their degradation, mainly because 70 of intensification of agriculture, forest clearance, industrial activities, groundwater 71 extraction, river flow regulation, and waste water discharge (Boulton et al., 2010; 72 Danielopol et al., 2003; Griebler et al., 2010; Tockner et al., 2008). These pressures 73 reduce the hydrological connection between ground and surface waters, and favour the 74 accumulation of pollutants that impair the characteristic biological structure and 75 functionality of groundwaters (Danielopol et al., 2003; Korbel et al., 2013).

76 Since the early 2000s, the importance of groundwater as a living ecosystem has 77 increased. Scientists and environmental managers are working to develop guidelines 78 and strategies for the conservation of groundwater ecosystems, taking into account their 79 biota and functionality (Gibert and Culver, 2009; Gibert et al., 2009; Gibert and 80 Deharveng, 2002). The majority of studies focus on describing groundwater 81 invertebrate (also called stygofauna) distribution patterns, such as the recent European 82 PASCALIS project (http://pascalis.univ-lyon1.fr/; Castellarini et al., 2007; Gibert et al., 83 2009). Results from this project have identified temperate areas as hot-spots of 84 groundwater biodiversity, and have suggested that the full potential of groundwater 85 ecosystems as sources of biodiversity and ecosystem services is still to be fully

86 comprehended (Deharveng et al., 2009). For this reason, researchers have proposed the 87 use of groundwater invertebrates as bioindicators of ecological groundwater health in 88 the same way that they are used in surface waters (Boulton et al., 2008; Iepure et al., 89 2013). The distribution of groundwater organisms indeed responds to hydrogeological and environmental factors, such as floods, dissolved organic carbon, and the 90 91 concentration of nutrients and pollutants (Castellarini et al., 2007; Danielopol et al., 92 1994; Di Lorenzo et al., 2014; Iepure et al., 2014, 2013;), which makes them suitable 93 candidates to reflect the ecological status of subterranean habitats. Furthermore, 94 groundwater invertebrates contribute to the functioning of groundwater ecosystems, and 95 consequently to the provision of groundwater ecosystem services used by humankind. 96 For example, amphipods and isopods are keystone taxa groups that enhance the 97 decomposition of organic matter, which in turn stimulates bacterial growth and activity, 98 supporting the biodegradation of contaminants (Tomlinson and Boulton, 2010; Ward et 99 al., 1998). These crustaceans also recycle nutrients through the excretion of faecal 100 pellets, and increase the flux of oxygenated water through burrowing (Hakenkamp and 101 Palmer, 2000). However, such functional processes have been poorly studied, 102 preventing the development of consistent ecological criteria for assessing groundwater 103 ecological status. More research is thus needed to ascertain the response of subterranean 104 biodiversity patterns and functionality to both natural and human-induced changes (Culver and Sket, 2000; Deharveng et al., 2009; Hancock et al., 2005; Korbel et al., 105 106 2013).

One of the most interesting subterranean habitats is the hyporheic zone, which is the
area of mixing between surface and subsurface water (White, 1993; Wondzell, 2011).
As surface water contains rich oxygen and organic matter and groundwater contains

110 abundant nutriment elements, the water mix between those two systems has a 111 significant impact on water quality, ecosystems and biogeochemistry cycling (Boulton et al., 1998; Brunke and Gonser, 1997; Krause et al., 2013; Marmonier et al., 2012; 112 113 Sanchez-Pérez et al., 2003; Sánchez-Pérez and Trémolières, 2003; Vervier et al., 2009). 114 These ecosystems support important agricultural activities, and, consequently, 115 groundwater in alluvial plains often suffer from nitrate pollution (Arrate et al., 1997; 116 Almasri and Kaluarachchi, 2007; Liu et al., 2005; Sánchez-Pérez et al., 2003a). Several 117 studies show that the hyporheic zone (i.e. surface-groundwater interface) contributes to 118 nitrogen retention and/or transformation of the land-surface water continuum (Sabater et 119 al., 2003; Weng et al., 2003). This zone supports the purification of water by its ability 120 to eliminate nitrates during their infiltration through the vegetation-soil system to 121 groundwater, but also through diffusion from groundwater to surface water (Sanchez-122 Perez et al., 1991a, 1991b; Takatert et al., 1999). However, little is known about the 123 groundwater biota and its role in these purification processes, and, definitely in the general ecosystem functioning. 124

125 To this end, this study evaluates for the first time the effect of land use on the 126 groundwater invertebrate community of the hyporheic zone in four river floodplains 127 located across SW Europe. First, we hypothesized that forest areas would show higher 128 groundwater invertebrate diversity than agricultural areas, since forests provide high 129 heterogeneity and quality in terms of resources (Brunke and Gonser, 1997; Datry et al., 130 2005). Moreover, we expected the groundwater invertebrate community to change 131 along a land use gradient, showing varying characteristic taxa and functions. For 132 example, the vegetation of forest areas adds debris to the system, increasing the organic 133 matter content in soils and groundwater. Consequently, the presence of detritivores and 134 omnivores, which are essential for organic matter decomposition and nutrient cycling in 135 groundwater ecosystems, is favoured (Avramov, 2014; Boulton et al., 2008; Vannote et 136 al., 1980). In contrast, pollution-tolerant taxa are likely to dominate agricultural areas 137 because of the high levels of fertilizers and manures employed, as was observed by 138 Dumas and Lescher-Moutoué (2001). Thus, our study allows us to evaluate the overall 139 role of riparian corridors as natural biofilters of pollutants from agricultural areas. 140 Ultimately, this study is essential for understanding the local variation in groundwater 141 assemblages and their response to human pressures, contributing to the development of 142 suitable conservation and management policies.

143

## 144 **2. Material and methods**

145 2.1 Study area

For this study, we selected four river floodplains located in South-western Europe: Garonne, Ebro, Bidasoa and Tajo. Floodplains were selected in areas characterized by a combination of agricultural occupation and patches of natural riparian forest that might favour the potential degradation of agricultural pollutants. The study covers a wide gradient of climatic and environmental conditions, from floodplains with little forest use to others with a high forest surface. To our knowledge, no studies on groundwater invertebrates have been performed in any of the four selected floodplains.

The Garonne study area was located 30 km north of Toulouse in South-West France (UTM 355758/4861267; Figure 1, Table 1). This floodplain site is dominated by agriculture, mainly corn, sunflower and poplar plantations (Table 1). Its riparian forest is dominated by poplar (*Populus* spp.), ash (*Fraxinus* spp.) and oak (*Quercus* spp.). The river flow at the study site is very variable, peaking in winter and spring (2880 m<sup>3</sup>·s<sup>-1</sup>, during the study period that corresponds to a 10 years return period) and declining in summer (47  $\text{m}^3 \cdot \text{s}^{-1}$ ).

160 The Ebro study site was located 12 km downstream of Zaragoza, Spain (UTM 161 686179/4607010; Figure 1, Table 1). It is mostly subjected to agricultural land use; 162 mainly corn and irrigated cereals, but it shows an extensive riparian corridor dominated 163 by white poplar (*Populus alba*), European black poplar (*Populus nigra*), white willow 164 (*Salix alba*) and salt cedar (*Tamarix* spp.) (González et al., 2010). River discharge 165 varied from 205 to 1450 m<sup>3</sup>·s<sup>-1</sup> during the study period.

166 The Bidasoa study area was located 7 km upstream of Irun, Spain (UTM 167 602190/4797250, Figure 1). Agriculture (mainly corn and pasture) and livestock 168 practically cover the entire floodplain surface, leaving a tight riparian corridor along the 169 river shore (Table 1). White willow (*Salix alba*), alder (*Alnus glutinosa*) and ash 170 (*Fraxinus excelsior*) dominate this riparian corridor, and the invasive species American 171 pokeweed (*Phytolacca Americana*) is also abundant. River discharge varied between 9 172 and 630 m<sup>3</sup>·s<sup>-1</sup> during the studied period.

The Tajo study site was located 30 km downstream of Toledo, Spain (UTM 380482/4410115; Figure 1, Table 1). This floodplain site is under extensive agriculture; mainly irrigated cereals, corn and pasture (Table 1). Only some isolated patches of riparian forest remain, which are dominated by white poplar (*Populus alba*) and salt cedar (*Tamarix* spp.). The river is highly regulated by dams and water derivations, with a discharge that hardly varied during the study period (10 to 12 m<sup>3</sup>·s<sup>-1</sup>).

The piezometric network in each floodplain was designed considering the floodplain
surface and land use. For this study, we sampled 11 piezometers in the Garonne River,
12 piezometers in the Ebro River, 7 piezometers in the Bidasoa River, and 10

piezometers in the Tajo River, covering different percentages of land use surface (see Figure 1). More detailed information about the piezometric network design, environmental conditions, and hydrodynamics of each studied area can be found in Antiguedad et al. (to be submitted in the same special issue) and Bernard-Jannin et al. (to be submitted in the same special issue).

187 We calculated the surface occupied by each type of land use within a buffer of 50 m 188 around each piezometer using QGIS v2.6.1. For the buffer selection, we took into 189 account the groundwater velocity in the alluvial aquifers (i.e. hydraulic conductivity). For the hydraulic conductivity range  $(10^{-3} \text{ to } 10^{-5} \text{ m}^3 \cdot \text{s}^{-1})$  in our studied areas, we 190 191 estimated that a particle (e.g. nutrients, pollutants) would move between 1 and 100 192 metres per day. It is for this reason that we selected an average surface of influence (i.e. 193 buffer) of 50 metres. Each piezometer was classified as agriculture-dominated when this 194 land use represented  $\geq$  40% of the buffer surface and otherwise as forest-dominated.

## 195 2.2 Taxonomic and functional composition of the invertebrate community

One groundwater invertebrate sample was collected at each piezometer every three months for one year: (i.e. April/May 2013, July/August 2013, October/November 2013 and January/February 2014). This meant that 90-100 L of groundwater were extracted at each piezometer with a manual pump (based on Malard et al., 2002) and filtered through a net with a mesh of 65 µm. Samples were preserved in situ in 70% ethanol. Groundwater invertebrate samples were sorted and identified in the laboratory at least to order level (Table A1, Appendix).

At the taxonomic level, three metrics were calculated: the total abundance of individuals, the total richness of taxa, and the Shannon-Wiener diversity index. To characterize the functional composition of the macroinvertebrate community, we used

206 feeding habits, the biogeochemical filtration capacity, and the particulate organic matter 207 (POM) breakdown capacity. In terms of feeding habits, we used affinity scores from 208 zero to five for each of the following habits: absorber, deposit feeder, shredder, scraper, 209 filter-feeder, piercer, predator, and parasite. We used a fuzzy coding approach based on 210 scores published in Tachet et al. (2002). For those taxa not included in this reference 211 guide, we classified organisms using information from the scientific literature as well as 212 expert consultation (Table A2, Appendix). A score of zero indicated no affinity, and a 213 score of five indicated the highest affinity of the taxon to a particular feeding habit. For 214 taxa identified at higher taxonomic levels than genera, the most frequent score across all 215 taxa belonging to a particular taxonomic group was selected. This may have resulted in 216 a certain underestimation of functional diversity, although according to Dolédec et al. 217 (2000) the overall functional structure of the invertebrate communities is conserved. 218 The matrix containing sites per taxa abundance was multiplied by the taxa per feeding 219 habit matrix to calculate three functional diversity metrics: abundance of individuals 220 showing affinity for each feeding habitat (i.e. functional abundance), total richness of 221 feeding habits (i.e. functional richness), and the Shannon-Wiener diversity of feeding 222 habits (i.e. functional Shannon diversity index). To assess the biogeochemical filtration and POM breakdown capacities of the community, we used the functional scores 223 224 defined by Boulton et al. (2008). Thus, for each taxon, a biogeochemical filtration and 225 POM breakdown efficiency score was assigned: 0: no or unknown direct role, 1: minor 226 role, 2: moderate role, and 3: major role (Table A2, Appendix). We expressed the 227 biogeochemical filtration capacity of the groundwater community as the product of the 228 absolute abundance of each taxon by its efficiency score.

Taxonomic and functional diversity metrics were computed with the 'vegan' (Oksanen
et al., 2013) and 'ade4' (Thioulouse et al., 1997) packages of R software, 2.15.3 (R
Core Team, 2013).

## 232 2.3 Environmental variables

Water samples were collected at each piezometer with a submersible pump. To ensure that the water sample corresponded to the aquifer and not stagnant water accumulated in the piezometer, we previously extracted groundwater until conductivity and oxygen values were constant (Griebler et al., 2010; Sánchez-Pérez, 1992). Water samples were transferred into a 1.5 L PVC bottle previously washed in acid (ClH 0.1 N), and placed on ice for transportation to the laboratory (see total number of samples in Table 1).

239 The alkalinity of unfiltered water samples was estimated within 4 h of collection by 240 automatic titration with H<sub>2</sub>SO<sub>4</sub> 0.04 N (APHA, 1989). Total suspended solids, total 241 dissolved solids and organic matter content were determined by the gravimetric method, 242 i.e. filtering of samples through pre-combusted (450°C, 4 h) Whatman GF/F glass-fibre 243 filters following standard protocols (APHA, 1989). Filtered water aliquots were stored 244 at -20 °C and used within one month for the following analyses. Ion chromatography 245 (Metrohm 861 Advanced Compact IC; APHA, 1989) was applied to determine ammonia  $(NH_4^+)$ , nitrate  $(NO_3^-)$  and sulphate  $(SO_4^{2-})$  concentration. Modified Berthelot 246 247 reaction using salicytate and dichloroisocyanurate was used to determine the amount of 248 ammonia  $(NH_4^+)$  by the colorimetric method (Krom, 1980) for Bidasoa River samples. 249 Soluble reactive phosphorus (SRP) was measured by the ascorbic acid method and 250 determined by the colorimetric method (APHA, 1989). Total dissolved phosphorus 251 (TDP) was also estimated by the ascorbic acid method, but potassium persulphate digestion was performed beforehand (90 min, 115 °C) (APHA, 1989). Finally, water 252

temperature, pH, conductivity, dissolved oxygen, and oxidation-reduction potential
(ORP) were recorded in situ with portable probes. The depth of the water table was also
measured with a sound piezometric probe.

#### 256 2.4 Statistical analysis

All statistical analyses were based on log-transformer data (with the exception of water pH, left untransformed) to normalize distributions and linearize relationships. However, water physicochemical parameters and diversity metrics still showed a non-normal distribution (Kolmogorov-Smirnoff test, P < 0.05). Thus, non-parametric Mann-Whitney *U* (for two samples) and Kruskal-Wallis (for k samples) tests were applied to identify significant differences in diversity metrics between rivers, land use types within each river, and seasons.

The response of diversity indices (both taxonomic and functional) and ecosystem services (biogeochemical filtration and POM breakdown capacities) to increasing forest surface was plotted and evaluated by simple linear regression after checking that no increase in fitness was achieved when quadratic or logistic functions were used.

268 To evaluate changes in the composition of groundwater invertebrate assemblages in 269 response to land use types, a similarity test was used. This test allows detection of the 270 most characteristic taxa and functions associated to each land use, as well as those that 271 better explain the dissimilarity between land use types. Similarity tests were performed 272 for each floodplain separately by means of two-way SIMPER analysis with squared-273 root transformed abundances, Bray-Curtis similarity distance, and a cut-off for low 274 contributions set at 90%. Taxa and feeding traits that contributed more to dissimilarity 275 (i.e. with values higher than one of the quotient between average dissimilarity and

standard deviation [Av. Dissimilarity/SD > 1]), were considered significant in terms of
discriminating land use types (Clarke and Warwick, 2001; Ruhí et al., 2012).

278 Linear mixed-effect models (LME, Laird and Ware, 1982) were used to identify the 279 environmental characteristics that controlled taxonomic and functional diversity. This 280 statistical technique was used to avoid the co-dependence effect introduced by repeated 281 measurements over time and across the four riparian areas (Demidenko, 2004). 282 Physicochemical parameters (non-correlated, Spearman rank test P < 0.6) and forest 283 surface percentage were included as fixed effects in LME models. Sampling season and 284 floodplain identity were included as random factors. Model selection followed an 285 automatic stepwise forward regression selection of predictors, based on the lowest 286 Akaike Information Criteria (AIC) that quantify the goodness of fit of multiple 287 alternative models.

Non-parametric analyses of variance (Mann-Whitney U test and Kruskal-Wallis test)
and linear regressions were performed with SPSS version 18.0 (<sup>©</sup>SPSS, Inc., Chicago,
IL). SIMPER tests were performed with PRIMER v. 6.0 for Windows. LME models
were computed with the 'nlme' (Pinheiro et al., 2013), 'MuMIn' (Barton, 2013),
'MASS' (Venables and Ripley, 2002), 'lme4' (Bates et al. 2013), and 'effects' (Fox,
2003) packages in R version 2.15.3 (R Core Team, 2013).

294

## **3. Results**

296 3.1 Groundwater invertebrate community and associated ecosystem services

297 Diversity indices and the associated ecosystem services (i.e. biogeochemical filtration

and POM breakdown) showed a linear positive response with increasing forest land use.

299 Moreover, the percentage of forest land use was able to explain between 14 and 52% of

300 the diversity indices and associated ecosystem services of groundwater communities301 (Figures 2, 3, and A1, Appendix).

302 In general, the taxonomic diversity and composition of groundwater invertebrate 303 communities showed differences between river and land use types (Table 1, Figures 4 304 and A2, Appendix). Particularly, more than 30% of the community in forest areas was 305 composed of crustacea (mainly Copepoda and Amphipoda), and Oligochaeta was one of 306 the most abundant taxa in agricultural areas. Moreover, SIMPER analysis showed that 307 Amphipoda was one of the taxa that contributed most to the dissimilarity between land 308 use types in three out of the four studied floodplains (Garonne, Ebro, and Bidasoa). The 309 abundance of Ostracoda was particularly important in the dissimilarity between land 310 uses in the Tajo River floodplain (Table 3, and A3, Appendix). At the functional level, 311 the most abundant (ca. 50%) functional feeding group corresponded to deposit feeders 312 (Figure 5), which also contributed most to the dissimilarity between land use types in all 313 of the studied floodplains (SIMPER analysis, Table 3 and A4, Appendix). The 314 proportion of shredders, scrapers, and filter-feeders was generally higher in forest than 315 in agricultural areas (Figure 5). Diversity indices and the associated ecosystem services 316 remained practically unchanged between sampling campaigns (i.e. seasons, Figure A3, 317 Appendix).

318

## 319 *3.2 Environmental factors controlling diversity indices and ecosystem services*

In general, concentration values of dissolved oxygen and nitrate in piezometers located
in agricultural areas were almost double those in forest areas, whereas conductivity and
DOC increased under forest land use (Table 2).

According to LME, the percentage of forest area and water quality (mainly, nitrate, sulphate, phosphate, and DOC concentrations) explained between 51 and 71% of the response of diversity indices and the associated ecosystem services (Table 4; Figure A4, Appendix). Diversity indices and ecosystem services had a positive response to the increase in the forest use surface and the groundwater content of nitrates, sulphates, and DOC. Likewise, phosphate concentration had a negative influence on diversity indices (Table 4; Figure A4, Appendix).

330

## 331 **4. Discussion**

332 In this study we demonstrate that the diversity and functionality of groundwater 333 invertebrates increase with the area occupied by riparian corridors in floodplains 334 affected by intensive agricultural development. This is probably because riparian forests 335 provide greater availability of quality resources (such as organic matter) to groundwater 336 organisms and unpolluted groundwater. More importantly, riparian corridors supported 337 organisms that provide important ecosystem services, such as biogeochemical filtration 338 and POM breakdown. Similar results were observed in other studies (Boulton et al., 339 2008; Korbel and Hose, 2011), where changes in the composition, abundance, and 340 richness of the groundwater fauna were directly related to ecosystem functions. Our 341 study therefore highlights the need to conserve or even increase the forest area in 342 floodplains affected by intense agricultural use as a means to create buffer areas that 343 favour groundwater biodiversity and, consequently, the biogeochemical degradation of 344 pollutants coming from agricultural areas.

345

346 *4.1 How does land use affect groundwater biodiversity and functionality?* 

347 Groundwater invertebrate assemblages showed notable changes between land use types, 348 at both taxonomic and functional levels. For instance, Amphipoda and Copepoda were 349 the most abundant taxa in the forest areas, whereas Oligochaeta and Ostracoda were 350 more frequent in agricultural areas. In the Bidasoa floodplain, where agricultural and 351 livestock activities are intensively developed, we mainly found species associated with 352 poor water quality, such as Oligochaeta, Diptera and Nematoda (see Figure 4). In 353 contrast, Ostracoda, often associated with low groundwater level fluctuations (Malard et 354 al., 1996), was the most abundant taxon in the Tajo River, which is consistent with the 355 low river discharge fluctuations registered during the study period (10 to 12  $\text{m}^3 \cdot \text{s}^{-1}$ ). 356 Similar results were found by Danielopol (et al., 2003), who suggested that river 357 regulation combined with the negative effect of organic pollution strongly alters 358 groundwater habitats. This is partly explained through river regulation which reduces 359 water energy, thereby increasing the transport large amounts of fine sediments (e.g. silt 360 and clay) that are accumulated in the aquifer, favouring its siltation. This siltation 361 reduces the porosity of alluvial aquifers and, consequently, reduces the space to host 362 fauna and reduces water and energy exchange between rivers and alluvial aquifers. 363 Moreover, the high content of organic pollution together with the characteristic low 364 oxygen concentration of alluvial aquifers favours the creation of chemical-reducing conditions that also eliminate groundwater fauna. Consequently, free-moving 365 366 crustaceans such as stygobiotic copepods and isopods are replaced by epigean animals 367 (i.e. surface animals), mostly nematodes and oligochaetes, which are able to live under 368 these conditions (Danielopol et al., 2003).

Unlike agricultural use, forest use does not add pollutants to groundwater, favouring the
provision of better-quality habitats and resources. Forest areas can thus act as local hot-

371 spots of food resources for groundwater invertebrates, thanks mainly to the organic 372 matter provided by the vegetation (Boulton and Foster, 1998). More particularly, 373 organic matter is degraded by surface fauna, increasing dissolved organic carbon 374 content in soils and infiltrated water. Consequently, dissolved organic carbon content 375 increases in groundwater under forest areas and is used as a food resource for many 376 taxa, such as Amphipoda and Isopoda. The increase of this kind of taxon also favours 377 better ecosystem functioning. For example, shredder crustaceans are keystone species 378 for nutrient cycling in groundwater ecosystems (Avramov, 2014; Boulton et al., 2008) 379 because of their role in bioturbation and the compaction of fine sediments into faecal 380 pellets (Boulton et al., 2008). Such biological processes boost the substrate availability 381 for bacteria, which in turn enhance denitrification processes and the transformation of 382 micropollutants in groundwaters, thereby contributing to healthy waters (Gibert and 383 Deharveng, 2002; Tomlinson and Boulton 2010; Ward et al., 1998; Yao et al., 384 submitted). Consequently, floodplains dominated by forest land use, such as the Ebro 385 and Garonne, showed the greatest biogeochemical filtration and POM breakdown 386 capacities. It thus follows that the loss of groundwater invertebrates in agricultural 387 areas, particularly of keystone species such as amphipods and isopods, can have serious 388 repercussions for the functioning of groundwater ecosystems (Boulton et al., 2008; 389 Korbel et al., 2013) and the associated supporting and provisioning ecosystem services 390 (e.g. purification of water, sustainability of surface ecosystems, provision of fit/clean 391 water for agricultural use) (Danielopol et al., 2003).

392 On the other hand, increase of nitrates, phosphates, and sulphates as a consequence of 393 agricultural fertilization and pesticides application influenced the groundwater 394 invertebrate community (Tables 2 and 4). These compounds can inhibit the

395 development of key taxa (mainly amphipods and isopods) at high concentrations 396 (Fakher et al., 1998), and favour the presence of other pollution-tolerant taxa such as 397 Oligochaeta. Other studies (Di Lorenzo et al., 2014; Iepure et al., 2013; Yao et al., 398 submitted) also observed a negative influence of human use (e.g. agricultural, urban or 399 industrial) on groundwater invertebrate communities that was mainly associated with 400 the deterioration of water quality and ecosystem functioning. In contrast, other authors 401 (Boulton et al., 1998; Castellarini et al., 2007; Di Lorenzo et al., 2005; Dole-Olivier et 402 al., 2009; Dumas and Lescher-Moutoué, 2001) found little evidence that land use and 403 the associated changes in water chemistry affect the composition of stygobiotic 404 assemblages. These authors emphasized the role of hydrological (e.g. hydraulic 405 conductivity) and geomorphological (e.g. pore size) factors instead. This discordant 406 result can be explained by the strong agricultural pressure in our studied areas, 407 characterized by long periods of low discharges that usually coincide with fertilization 408 and irrigation periods. Diffuse pollution from agricultural areas may thus override the 409 effects of other hydrological and geomorphological determinants. However, the role of 410 hydrological factors on groundwater invertebrate communities should not be 411 disregarded, especially given its importance for surface waters (Español et al., 2015; 412 Gallardo et al., 2008, 2009).

413

414 *4.2 Conclusions* 

The results of this study demonstrate that land use impacts on groundwater biota and their associated ecosystem services. We observed a negative influence of agricultural land use on invertebrate diversity and associated ecosystem services, which is attributed to a high concentration of nitrates and sulphates. In contrast, riparian corridors provide

419 food resources (dissolved organic matter) and better water quality (low nitrate and 420 sulphate concentration) that enhance biota development and, consequently, the 421 associated ecosystem services (e.g. POM breakdown and biogeochemical filtration 422 capacities). Therefore, management measures to protect groundwater ecosystems should 423 include the conservation, creation, and eventually expansion of riparian forest corridors 424 in floodplains occupied by intensive agriculture. In addition, the rationalization of 425 agricultural fertilization activities and pesticides application are needed to reduce 426 diffuse contamination of subterranean habitats.

427 In terms of biomonitoring, our results suggest that groundwater invertebrate abundance, 428 taxonomic richness, POM breakdown capacity, and biogeochemical filtration capacity 429 are appropriate indicators for comparing the composition and functionality of alluvial 430 aquifers and assessing the effect of human pressures. However, more detailed regional 431 long-term studies are needed for better understanding of the structure and function of 432 groundwater ecosystems, which remain poorly investigated. Such basic studies are 433 essential for developing the ecological criteria proposed by the European standard (EU-434 GWD, 2006) for assessing groundwater ecosystem status at the same level as surface 435 aquatic ecosystems.

436

## 437 **5. Acknowledgements**

This study was performed as part of the EU Interreg SUDOE IVB programme (ATTENAGUA - SOE3/P2/F558 project, <u>http://www.attenagua-sudoe.eu</u>) and funded by ERDF. The main author was funded by the Aragon government (Research Group E61 on Ecological Restoration and B079/09 pre-doctoral grant) and AISECO (student grant). This work has been made possible thanks to the work of the rest of

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- Figure 1. Study area location and piezometric network of each studied river floodplain.
   The study area is composed by four river floodplains located in South-western Europe
   (Garonne, Ebro, Bidasoa and Tajo Rivers). Samples were collected in the piezometric
   network (8 to 12 piezometers) of each floodplain.
- 5



Figure 2. Relationship between taxonomic and functional diversity indices and the percentage of surface occupied by forest (buffer 50 m) in the floodplain of four rivers in 

South-western Europe. 



- 12 Figure 3. Relationship between ecosystem services (i.e. biogeochemical filtration and
- 13 POM breakdown capacities) and the percentage of surface occupied by forest (buffer 50



14 m) in four river floodplains in South-western Europe.

16 Figure 4. Taxonomic composition of the groundwater invertebrate community of four 17 river floodplains according to their dominant land use type. Bars represent the relative 18 abundance (average percentage) of each taxon in the community for each land use type. 19 Significant differences between land use types are indicated by \* (P < 0.05, Mann-20 Whitney U test) and \*\* (P < 0.01).



Figure 5. Functional feeding habits of groundwater invertebrates in four river floodplains according to their land use. Bars represent relative abundance (average percentage). Significant differences between land use types are indicated by \* (P < 0.05, Mann-Whitney U test) and \*\* (P < 0.01).



27

- 1 Table 1. Study site characteristics, dominating land use type, diversity metrics (mean  $\pm$
- 2 SD), and provided ecosystem services (mean  $\pm$  SD) in four river floodplains (Garonne,

3 Ebro, Bidasoa and Tajo Rivers). N = sample size. Significant differences between river

- 4 floodplains are indicated by \*\* (P < 0.05, Kruskal-Wallis test).
- 5

	Units	Garonne	Ebro	Bidasoa	Tajo
		(N = 44)	(N = 48)	(N = 28)	(N = 40)
Piezometers	n°	11	12	7	10
Total Floodplain Area	На	1200	80	17	62
Habitat					
Agricultural area	На	960	30	15	52
Forest area	На	240	50	2	8
Hydrological conditions					
Hydraulic conductivity (aquifer)	m/day	86	1 - 3	50 - 100	8-9
River discharge (average)	m <sup>3</sup> /s	195	250	25	11
Specific flow (average)	m <sup>3</sup> /s·km <sup>2</sup>	$1.4 \cdot 10^{-2}$	$5.3 \cdot 10^{-3}$	$3.7 \cdot 10^{-2}$	$4.3 \cdot 10^{-4}$
Diversity indices					
Abundance *	n°indiv/100L	$776 \pm 645$	$354\pm560$	$11 \pm 10$	$30 \pm 44$
Taxonomic richness *	n° taxa	$7 \pm 1$	$6 \pm 1$	$3\pm 2$	$2 \pm 1$
Taxonomic Shannon diversity index *		$0.83\pm0.39$	$1.13\pm0.25$	$0.70\pm0.52$	$0.38\pm0.36$
Functional richness *	n° categories	$5 \pm 2$	$6 \pm 1$	$5 \pm 2$	$2 \pm 1$
Functional Shannon diversity index *		$0.77\pm0.16$	$1.24\pm0.10$	$1.07\pm0.40$	$0.52\pm0.41$
Ecosystem services					
Biogeochemical filtration capacity *		$1539 \pm 1272$	$639 \pm 1014$	$24 \pm 27$	$60 \pm 88$
POM breakdown capacity *		$873\pm717$	$635 \pm 1077$	$18 \pm 17$	$34 \pm 53$

8 Table 2. Groundwater physicochemical features (mean  $\pm$  SD) of four floodplains 9 located in the Garonne, Ebro, Bidasoa and Tajo Rivers under different land use types. N 10 = sample size. Significant differences between land use types in each river floodplain 11 are indicated by \* (P<0.05, Mann-Whitney *U* test).

12

		Garonne		Ebro		Bidasoa		Тајо	
Physicochemical	Units	Agricultural	Forest use (N	Agricultural	Forest use (N	Agricultural	Forest use	Agricultural	Forest use (N
features (Abbr.)		use $(N = 8)$	= 36)	use $(N = 12)$	= 36)	use $(N = 24)$	(N = 4)	use $(N = 24)$	= 16)
Depth (Depth)	m	2.26 ± 0.77 *	$3.28 \pm 1.28 *$	$3.90 \pm 0.71*$	$2.49\pm0.45*$	$4.17\pm0.76$	$3.99 \pm 0.73$	$3.53 \pm 1.20*$	$2.13 \pm 0.73^{*}$
Temperature	°C	$14.2 \pm 1.7$	$13.79 \pm 1.23$	$16.3 \pm 1.4*$	$14.0\pm1.8*$	$15.1 \pm 2.0$	$14.1 \pm 1.3$	$17.5\pm1.3$	$17.6\pm2.4$
(Temp)									
pH (pH)		$7.02\pm0.14$	$6.97\pm0.12$	$6.92\pm0.49*$	$7.36\pm0.28*$	$6.49 \pm 0.28$	$6.49 \pm 0.13$	$7.14\pm0.12$	$7.07\pm0.15$
Dissolved oxygen	%	$65.60 \pm 10.53$	$29.95\pm29.20$	$24.06 \pm 19.74$	$16.72\pm15.94$	37.66 ±	$38.00 \pm$	$42.34 \pm$	$26.10 \pm$
(O2)		*	*			17.80	8.94	32.67*	29.96*
Conductivity	µS/cm	$945 \pm 115$	$901 \pm 225$	$3464 \pm 646*$	$1618 \pm 959*$	$367 \pm 101$	$397 \pm 31$	$2234\pm410^*$	$2493 \pm 439 *$
(EC)									
Oxidation-	mV	$252 \pm 35$	$195 \pm 91$	$104 \pm 21*$	$68 \pm 34*$	$98 \pm 66$	$163 \pm 50$	$118 \pm 36$	$116 \pm 39$
reduction									
potential (ORP)									
Dissolved organic	mg/L	$0.69 \pm 0.45$	$1.22\pm0.98$	$13.95 \pm 3.47*$	$8.58 \pm 5.58*$	$3.31 \pm 1.22$	$3.08\pm0.85$	$1.61 \pm 0.60$	$1.99 \pm 0.87$
carbon (DOC)									
Alkalinity (Alk)	meq/L	$5.34 \pm 0.35 *$	6.02 ± 1.13 *	$6.39 \pm 0.44*$	$5.32 \pm 0.93*$	$3.15 \pm 1.01$	$3.23\pm0.32$	$5.78 \pm 1.01$	$5.98 \pm 1.31$
Phosphate (PO4)	µg/L	$9.28 \pm 8.53$	$9.70\pm9.59$	$2.68 \pm 1.89^*$	$16.47 \pm$	$5.13 \pm 3.33$	$3.65 \pm 1.52$	$182.08 \pm$	$160.00 \pm$
					20.48*			150.79	165.81
Ammonium	µg/L	$7.90 \pm 4.21$	$106.48 \pm$	$0.004 \pm$	$10.50 \pm$	3212.68 ±	$176.04 \pm$	199.94 ±	239.68 ±
(NH4)			269.98	0.014*	38.73*	9516.56	161.71	362.97	350.90
Nitrate (NO3)	mg/L	$76.88 \pm 15.96$	$47.26 \pm 38.73$	$34.68 \pm 4.85*$	$10.10 \pm 8.25*$	$9.23 \pm 8.84$	$8.71 \pm 3.36$	$32.25 \pm 12.58$	$38.66 \pm 16.16$
		*	*						
Sulphate (SO4)	mg/L	$67.38 \pm 10.79$	$64.81 \pm 29.66$	$1410.96 \pm$	$451.18 \pm$	21.96 ±	32.16 ±	924.13 ±	$1009.03 \pm$
				154.57*	421.78*	12.33	4.22	371.81	384.17
Silica oxide	mg/L	$13.01 \pm 1.00$	$11.43 \pm 3.97$	$18.66 \pm 4.06*$	$7.75 \pm 4.96^*$	$9.36 \pm 2.92$	12.98 ±	$15.23 \pm 3.16$	$14.81 \pm 3.50$
(SiO2)							4.33		
12									

Table 3. Taxa and feeding traits that contributed the most to dissimilarity (Av.
Diss/SD>1) between land use types (agricultural use areas vs. forest use areas).
Statistics obtained through a two-way SIMPER analysis with factors date and land use.

	GARONNE		EBRO		BIDASOA		TAJO	
xonomic level	(average	dissimilarity	(average	dissimilarity	(average	dissimilarity	(average	dissimilarity
	49.3%)		52.7%)		74.1%)		71.1%)	
	Taxa	Diss/SD	Taxa	Diss/SD	Таха	Diss/SD	Taxa	Diss/SD
	Copepoda	1.46	Amphipoda	1.39	Oligochaeta	1.67	Ostracoda	1.44
Та	Amphipoda	1.31	Copepoda	1.22	Amphipoda	1.02		
	(average	dissimilarity	(average	dissimilarity	(average	dissimilarity	(average	dissimilarity
	38.4%)		37.1%)		41.1%)		53.7%)	
	FFG	Diss/SD	FFG	Diss/SD	FFG	Diss/SD	FFG	Diss/SD
vel								
vel	Deposit feeder	1.60	Deposit feeder	1.44	Deposit feeder	1.38	Deposit feeder	1.54
al level	Deposit feeder Predator	1.60 1.52	Deposit feeder Shredder	1.44 1.43	Deposit feeder Shredder	1.38 1.24	Deposit feeder Scraper	1.54 1.09
tional level	Deposit feeder Predator Shredder	1.60 1.52 1.25	Deposit feeder Shredder Scraper	1.44 1.43 1.36	Deposit feeder Shredder Absorber	1.38 1.24 1.33	Deposit feeder Scraper Predator	1.54 1.09 1.04
<b>Functional level</b>	Deposit feeder Predator Shredder	1.60 1.52 1.25	Deposit feeder Shredder Scraper Predator	1.44 1.43 1.36 1.34	Deposit feeder Shredder Absorber Filter feeder	1.38 1.24 1.33 1.57	Deposit feeder Scraper Predator	1.54 1.09 1.04
Functional level	Deposit feeder Predator Shredder	1.60 1.52 1.25	Deposit feeder Shredder Scraper Predator Absorber	1.44 1.43 1.36 1.34 1.34	Deposit feeder Shredder Absorber Filter feeder Predator	1.38 1.24 1.33 1.57 1.47	Deposit feeder Scraper Predator	1.54 1.09 1.04
Functional level	Deposit feeder Predator Shredder	1.60 1.52 1.25	Deposit feeder Shredder Scraper Predator Absorber	1.44 1.43 1.36 1.34 1.34	Deposit feeder Shredder Absorber Filter feeder Predator Parasite	1.38 1.24 1.33 1.57 1.47 1.41	Deposit feeder Scraper Predator	1.54 1.09 1.04

Table 4. Results from LME models linking physicochemical variables with taxonomic and functional diversity indices and ecosystem services. All selected explanatory variables were statistically significant at P < 0.05. Total sample size = 160.  $d^2$  = variance of the random intercept;  $\alpha$  = variance of the fixed intercept;  $\rho$  = Spearman correlation coefficient between observed and predicted values, used as a measure of goodness of fit.

	Explanatory variables	Coefficient	Intercept	Spearman correlation test
Taxonomic diversity indices				
Abundance	% Forest area	0.30	$d^2 = 1.10^2$	$\rho = 0.71$
	NO <sub>3</sub>	0.28	$\alpha = 15.00$	P < 0.01
	Temp	- 4.55		
Taxonomic richness	% Forest area	0.09	$d^2 = 0.33^2$	$\rho = 0.57$
	$SO_4$	0.06	$\alpha = 3.21$	P < 0.01
	Temp	- 0.79		
Taxonomic Shannon	DOC	0.07	$d^2 = 0.10^2$	$\rho = 0.58$
diversity index	% Forest area	0.04	$\alpha = 1.50$	P < 0.01
	$O_2$	- 0.05		
	Temp	- 0.37		
Functional diversity indices				
Functional richness	% Forest area	0.08	$d^2 = 0.16^2$	$\rho = 0.51$
	$PO_4$	- 0.03	$\alpha = 2.99$	P < 0.01
	Temp	- 0.54		
Functional Shannon	DOC	0.07	$d^2 = 0.10^2$	$\rho = 0.61$
diversity index	EC	- 0.07	$\alpha = 0.91$	P < 0.01
	% Forest area	0.03		
Ecosystem services				
Biogeochemical filtration	% Forest area	0.30	$d^2 = 1.13^2$	$\rho = 0.69$
capacity	NO <sub>3</sub>	0.31	$\alpha = 15.38$	P < 0.01
	Temp	- 4.50		
POM breakdown capacity	% Forest area	0.34	$d^2 = 1.11^2$	$\rho = 0.70$
	NO <sub>3</sub>	0.28	$\alpha = 13.86$	P < 0.01
	Temp	- 4.11		

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Supplementary Material Click here to download Supplementary Material: Appendix\_att4.doc