

Genetic variants associated with methotrexate-induced mucositis in cancer treatment: a systematic review and meta-analysis

Running title	Genetic variants and methotrexate-induced mucositis
Authors	Hedy Maagdenberg ^{a,b,c,d} , Natanja Oosterom ^{e,f} , Jolanda Zanen ^{c,g} , Donato Gemmati ^h , Rachael E. Windsor ⁱ , Sandra G. Heilf, j, Shakila Jabeen ^{k,l} , Guillermo J. Ruiz-Argüelles ^{m,n} , Oliver Zolk ^o , Susanne Hoerning ^p , Charlotte Sleurs ^q , Elixabet Lopéz-Lopéz ^{r,s} , Mónica Moreno-Galván ^{t,u} , Marry M. van den Heuvel-Eibrink ^e , Anke H. Maitland-van der Zee ^a , and Bruce C. Carleton ^{b,c,d} .
Affiliations	<p>^a Department of Respiratory Medicine, Amsterdam University Medical Center, University of Amsterdam, Amsterdam, the Netherlands.</p> <p>^b BC Children’s Hospital Research Institute, Vancouver, BC, Canada.</p> <p>^c Division of Translational Therapeutics, Department of Pediatrics, Faculty of Medicine, University of British Columbia, Vancouver, BC, Canada.</p> <p>^d Pharmaceutical Outcomes Programme, BC Children’s Hospital, Vancouver, BC, Canada</p> <p>^e Princess Máxima Center for Pediatric Oncology, Utrecht, the Netherlands.</p> <p>^f Department of Clinical Chemistry, Erasmus MC University Medical Center Rotterdam, Rotterdam, the Netherlands.</p> <p>^g Department of Pharmaceutical Sciences, Utrecht University, Utrecht, the Netherlands.</p> <p>^h Department of Translational Medicine, and Centre Hemostasis & Thrombosis, University of Ferrara, Ferrara, Italy.</p> <p>ⁱ Children and Young People’s Cancer Service, University College London Hospitals NHS Foundation Trust, London, United Kingdom.</p> <p>^j Laboratorio de Biología Molecular, Depto. de Bioquímica Clínica, Facultad de Química, Universidad de la Republica, Uruguay.</p> <p>^k Division of Medicine, Department for Clinical Molecular Biology (EpiGen), Akershus University Hospital, Lorenskog, Norway.</p> <p>^l Department of Cancer Genetics, Institute for Cancer Research, Oslo University Hospital, Norway.</p> <p>^m Centro de Hematología y Medicina Interna, Clínica Ruiz de Puebla, Puebla, Mexico,</p> <p>ⁿ Laboratorios Clínicos de Puebla, Clínica Ruiz de Puebla, Puebla, Mexico</p> <p>^o Institute of the Pharmacology of Natural Products and Clinical Pharmacology, University Hospital, Ulm, Germany.</p> <p>^p Department for Pediatric Oncology and Immunology, University Hospital for Children and Adolescents, Erlangen, Germany.</p> <p>^q Department of Pediatric Hemato-Oncology, Katholieke Universiteit Leuven, Leuven, Belgium.</p> <p>^r Department of Genetics, Physical Anthropology and Animal Physiology, Faculty of Medicine and Nursing, University of the Basque Country (UPV/EHU), Leioa, Spain</p> <p>^s BioCruces Bizkaia Health Research Institute, Barakaldo, Spain.</p> <p>^t Departamento de Hemato-Oncología, Hospital Infantil de Mexico, Federico Gomez, Mexico City, Mexico.</p> <p>^u Sociedad Mexicana de Salud Publica, CENAPRECE, SSA, Mexico City, Mexico.</p>

Highlights

- 57 studies were included in this systematic review; 26 in the meta-analysis
- A total of 34 SNPs were associated with (more severe) mucositis in ≥ 1 study
- *MTHFR*c.677C>T was associated with more severe mucositis (OR 2.53, 95%-CI 1.48-4.32)
- *MTRR*c.66A>G was associated with mucositis (OR 2.08, 95%-CI 1.16-3.73)
- Current studies are heterogeneous in design with different case-control definitions

Abstract

Methotrexate (MTX), an important chemotherapeutic agent, is often accompanied with mucositis. The occurrence and severity are unpredictable and show large interindividual variability. In this study, we review and meta-analyze previously studied genetic variants in relation to MTX-induced mucositis.

We conducted a systematic search in Medline and Embase. We included genetic association studies of MTX-induced mucositis in cancer patients. A meta-analysis was conducted for single nucleotide polymorphisms (SNPs) for which at least two studies found a statistically significant association.

A total of 34 SNPs were associated with mucositis in more than one study of the 57 included studies. Two of the seven SNPs included in our meta-analysis were statistically significantly associated with mucositis: *MTHFR* c.677C>T (recessive, grade ≥ 3 vs grade 0-2, OR 2.53, 95%CI [1.48-4.32], False Discovery Rate[FDR]-corrected p-value 0.011) and *MTRR* c.66A>G (overdominant, grade ≥ 1 vs grade 0, OR 2.08, 95%CI [1.16-3.73], FDR-corrected p-value 0.042).

Keywords

Methotrexate, mucositis, gastrointestinal toxicity, pharmacogenomics, pediatric, adult, cancer.

1. Introduction

Methotrexate (MTX), a folate antagonist, is a cornerstone of the treatment of childhood and adult cancers such as acute lymphoblastic leukemia (ALL), osteosarcoma and non-Hodgkin lymphoma. Although MTX has shown its value in the treatment of these cancer types, it is often accompanied with mucositis. Previous studies showed that 10-40% of patients develop oral or gastrointestinal mucositis after intravenous high-dose MTX therapy despite administration of folinic acid (leucovorin) rescue therapy [1–3]. Risk factors for the development of these toxicities remain uncertain and the severity is highly variable

among patients suggesting a strong interindividual susceptibility. Mucositis can impair oral intake of food and liquids and an impaired quality of life during therapy, often requiring dose reductions or cessations of treatment which can interfere with treatment efficacy [1,4,5].

Many studies have been conducted on the association between genetic variations in genes involved in MTX pharmacodynamics (e.g. *MTHFR*) or pharmacokinetics (e.g. *SLC19A1*) and the development of mucositis (**Figure 1**). In short, MTX is transported into the cell by the reduced folate carrier 1 (*RFC1/SLC19A1*) and proton-coupled folate transporter (*PCFT/SLC49A1*). *SLCO1B1* is an important MTX transporter in the liver. Inside the cell, MTX is polyglutamated (MTX-PG) by Folylpolylglutamate Synthase (*FPGS*) which augments cellular retention and pharmacological activity of MTX, and is afterwards depolyglutamated by gamma-glutamyl hydrolase (*GGH*). MTX(-PG) inhibits Dihydrofolate Reductase (*DHFR*) and Thymidylate Synthase (*TS*). Often studied and relevant genes in the one-carbon and folate metabolism are Methylenetetrahydrofolate Reductase (*MTHFR*), Methylenetetrahydrofolate Dehydrogenase 1 (*MTHFD1*) and Methionine Synthase/Reductase (*MS/MTRR*) [6,7].

In addition, as coding regions only comprise about 1.5% of the entire genome, awareness was raised regarding the important regulatory functions of non-coding regions such as miRNAs [8]. Therefore, genetic variations in non-coding parts of the genome, such as in miRNA machinery genes and miRNAs have been studied in relation to developing MTX toxicity as well [8,9]. miRNAs play an important role in RNA-silencing and post-transcriptional regulation of gene expression [10]. Genetic variations in these genes and in miRNA products processed by these genes have been studied in relation to MTX-induced toxicity [8].

However, results are often inconsistent and not replicated by other studies. This is likely due to differences in patient characteristics, outcome definitions of toxicity, chemotherapy protocols, thoroughness of clinical phenotyping and often relatively small sample sizes. Several meta-analyses have been conducted for two of the most frequently studied *MTHFR* genetic variations c.677C>T (rs1801133) and c.1298A>C (rs1801131) [11–14]. However, these meta-analyses only included a small and variable subset of the available published studies. This resulted in different conclusions for c.677C>T from no association [11,12] to an increased risk of mucositis in patients with the TT-genotype compared to the CT- and CC-genotype [13,14]. For c.1298A>C three meta-analyses showed no association with an increased risk of developing mucositis [12–14], whereas one study showed an association (AC-genotype vs AA-genotype, Odds Ratio [OR] 1.690, 95%-CI[1.011 - 2.825] and CC-genotype vs AA-genotype, OR 1.889, 95%-CI [1.232-2.896]) between c.1298A>C and developing overall MTX toxicity [11].

To our knowledge no systematic review and meta-analysis of all the studied single nucleotide polymorphisms (SNPs) in relationship to MTX-induced mucositis has been performed. Consequently, it

remains unclear which SNPs have an increased likelihood of involvement in toxicity, and are thus relevant to be studied in more detail given their possible value in clinical prediction models. In this study, we review all previously studied SNPs and conduct a meta-analysis with the aim to identify relevant variants that could be implemented in polygenic clinical mucositis prediction models of toxicity.

2. Methods

This systematic review was carried out according to the Preferred Reporting Items for Systematic Reviews and Meta-Analyses guidelines (PRISMA) [15]. The protocol of this review and meta-analysis was published on PROSPERO, registration number: CRD42018092843.

2.1 Data sources and search strategy

Both the Medline (Ovid) database and Embase were searched from inception until the 9th of August 2019. The search terms used can be found in **Table S1**. References of all included studies, reviews, meta-analyses, and PharmGKB [16] were screened for additional articles.

2.2 Selection criteria

Studies were eligible when fulfilling the following inclusion criteria: (1) genetic association studies (candidate gene and genome wide association studies), (2) MTX chemotherapy, (3) studies that included mucositis or gastrointestinal toxicity (including mucositis) as an adverse outcome of MTX treatment, (4) studies published in English language, and (5) studies including cancer patients. Studies had to contain a description of an analysis and results of the association between genetic variants (1) and the development of mucositis (3). Reviews, case reports, letters to the editor, conference abstracts, preclinical studies (*in vivo* or *in vitro* studies), and studies after a stem cell transplantation/ bone marrow transplantation were excluded. All the identified records were screened for meeting the inclusion criteria based on the title and abstract. If it was not clear whether to include or exclude a study, based on screening the title and abstract, the article was assessed full-text. The study selection was conducted by one reviewer.

2.3 Data extraction

Data extracted from full-text articles were: last name of first author, year of publication, country, study design (Genome Wide Association Study [GWAS] or candidate gene), chemotherapy (MTX dose, protocol phase, administration of folinic acid rescue), patient population (sample size, age, sex, ethnicity, type of cancer), and mucositis (definition, grading system used, prospectively or retrospectively collected and

graded), genetic variants (name of gene, rs number, position, and genotype frequency). If the rs number was not given in the article, PharmGKB [16], SNPedia [17] or PubMed dbSNP [18] were used to find the rs number. For each studied genetic variant, data on the occurrence of mucositis per genotype and per grade, OR and 95% confidence interval (CI) and p-value was collected. When this information was not available the authors were requested to provide this information. The data were extracted by one researcher and checked by a second researcher.

2.4 Risk of bias and quality assessment

Publication bias was assessed by a Funnel plot and Egger's test if more than 10 studies were included in the meta-analysis for one single SNP. The quality and risk of bias of each article was assessed using a scoring system based on the STREGA recommendations [19] from a previously published study [20] (**Table S2**). The assessment was performed by two researchers independently and discrepancies were discussed to reach consensus.

2.5 Data-analysis

A meta-analysis was conducted for all genetic variations for which in at least two studies a statistically significant association was found. By using the four-genetic model strategy described by Horita and Kaneko, the most applicable genetic model (recessive, allelic, dominant and overdominant) was chosen per genetic variation [21]. For this specific genetic model, the odds ratio was estimated and the 95%-CI using a random-effects model (DerSimonian-Laird method). To correct for multiple testing we used a False Discovery Rate (FDR)-correction. For the primary analysis, the case-control (case-comparison) definition of severe mucositis (grade ≥ 3) vs less severe or absent mucositis (grade 0-2) was used, if possible. This definition was selected, because grade ≥ 3 defines a clear and severe mucositis, which can also be easily recognized when graded retrospectively based on notes in patient files and is therefore often used in studies. Only studies with the information available for the specific model and case-control definition were included in the analysis. For each study, SNPs were tested for deviation from Hardy-Weinberg equilibrium (HWE) in a control group. SNPs that were not in HWE were excluded from the meta-analysis. Study heterogeneity was estimated using the Cochran's Q test and reporting of the I-squared statistic. As a sensitivity analysis to test the robustness of the meta-analysis, the meta-analysis was repeated several times, each time excluding one study (Leave-One-Out analysis).

Subgroup analyses were conducted within pediatric patients, patients using intravenous (IV) MTX (often high-dose MTX) and ALL patients. Furthermore, we tested the influence of using another cutoff value for mucositis grade to define a patient as case or control.

Statistical analysis were performed using R version 3.5.0.

3. Results

3.1 Study eligibility and characteristics

In total, 259 unique records were identified in Medline and Embase and one additional record was found by cross reference check of reviews/meta-analyses, see **Figure 2**. After screening of the titles and abstracts, 106 articles were selected. Of these, 49 articles were excluded based on reading the full-text (**Table S3**). In total 57 studies were included in the qualitative synthesis. Of the 57 studies 26 were included in the meta-analysis. The study characteristics of the 57 included studies are depicted in **Table S4**. All studies were published between 2005 and 2019 and most were set up as candidate gene studies with the exception of the study of Treviño et al. that conducted a genome-wide association study [22]. Only two studies, i.e. Treviño et al. and Stocco et al., included an independent replication cohort [22,23].

3.2 Study population

In total 7269 patients were studied for associations between genetic variations and MTX-induced mucositis. The sample size per study ranged from 15 to 484 patients with a median of 89 patients. Of the studies which provided information on the sex of the patients (n=49), 41 studies included more males than females. In 43 of the 57 studies only pediatric patients were included. The other 14 studies included adults (n=7), included adult and pediatric patients (n=6) or age was not specified (n=1). Ethnicity was specifically reported in 18 studies, which was mainly Caucasian/white in 12 studies and Asian (Chinese) in 6 studies [5,12,22–37]. One study reported the genetic ancestry but did not report self-identified ethnicity [38]. Most studies were carried out in Europe (n=24) and Asia (n=19). Seven studies were conducted in North America and three in South America. Most studies were carried out on patients with ALL (n=42). Other cancer types were osteosarcoma (n=6), NHL (n=3) or multiple cancer types within one study (n=6).

3.3 Chemotherapy protocols

A large range of chemotherapy protocols was used in the included studies. Thirty three of the 57 studies only included patients treated with the same chemotherapy protocol. The protocol number and name

were not reported in 7 studies. Most commonly, one or more Berlin-Frankfurt-Münster protocols were included (n=19). Across the protocols, MTX was administered in various dosages and administration formulas. However, most studies focused on the protocol phase in which IV MTX (n=50) was administered, in which the MTX dosages varied from 0.1 to 12 g/m² (n=44 IV MTX of 2-5 g/m²). In all these studies, MTX administration was combined with folinic acid rescue.

3.4 Mucositis

The NCI CTCAE (v1.0–5.0) were used in most studies to grade the severity of mucositis (n=38). In addition, the WHO criteria (n=13), CCG toxicity criteria (n=1) and Oral mucositis index (OMI-score; n=1) were used. Three studies did not use a grading system, but gave a description of what they considered as mucositis being present. Two studies did not report the grading system or a case definition. The cutoff value to classify a patient as mucositis case varied between the studies from mucositis of any grade (grade 1-4) to only severe mucositis (grade 3-4). The case definition was clearly described in most studies; however, the control definition was often missing. Not all studies used a case-control setup to test the association. Some studies used the number of variant alleles or a sum score (adding up all grades of the different courses) in a linear regression model. Other studies used the number of cycles with and without a mucositis event, others the number of patients with at least one event with a grade defined as case during total follow-up. Also, the timing of grading varied (26 prospectively [at the moment the event occurred] and 15 retrospectively [later based on notes in patient files]) and was not always defined (n=16).

3.5 Quality assessment

As none of the studies used a control group (not using MTX), the quality of all the included studies was assessed for only the first four items of **Table S2**. We scored the studies on the reporting of important patient characteristics including at least age, sex, and the number of patients per cancer type, per ALL risk group and per ALL immunophenotype, **Table S5**. In many studies, these patient characteristics were not completely reported. In 27 out of the 48 included studies with ALL patients the number of patients per ALL risk group was not reported. Fourteen of the 48 included ALL studies did not report the ALL immunophenotype. Age, sex, and number of patients were reported in most studies. In 6 studies, one or more of these characteristics were missing.

For the characteristics of treatment, we scored the reporting of the protocol name or MTX dose, route of administration of MTX and number of cycles. All studies except one reported this information.

Thirty-three studies reported whether the distribution of genotypes was in Hardy Weinberg Equilibrium. The quality of genotyping was addressed in 21 studies. The methods for checking the quality included having control samples in the genotyping analysis, repeating the genotyping of all samples with a different method, or reporting the percentage of samples which were successfully genotyped or provided call rates. Description of the genotyping methods used was very limited in 12 studies and in two studies completely missing. In a large number of studies, it was not clear which cells and at which time point (diagnosis or remission) DNA was collected. This information was completely missing in 6 studies. In 31 studies, only the type of material was described. In 19 studies the timing of DNA collection and the type of biological specimen was reported.

A sample size calculation was rarely provided (n=2). In the study by Kotnik et al. from 2017 the sample size calculation was based on the MTX toxicity and genotype association. Correction for multiple testing was carried out in a small number of studies (n=9). Multiple testing correction was not required in seven of the other studies, because of the evaluation of a single SNP.

The geographical locations of the studies were either stated in the article or could easily be deduced based on the author's affiliation. Two studies adjusted for self-reported ethnicity in their analyses.

3.6 Genes studied in relationship with mucositis

A large number of SNPs involved in the pharmacodynamics (n=12) and in the pharmacokinetics of MTX (n=11) were studied for their association with mucositis or gastrointestinal toxicity (**Table S6**). In addition, 12 SNPs in non-coding pre-microRNAs and miRNA processing genes were also studied by Gutiérrez-Camino et al. [9,39] and López-López et al. [8]. In total 34 SNPs in 28 (miRNA processing) genes/pre-miRNAs were at least once statistically significantly associated with mucositis (p<0.05). Of these, 7 SNPs were replicated in at least one other study (**Table 1**), and were included in our meta-analysis.

3.6.1 MTHFR c.677C>T (rs1801133)

Multiple studies (n=38) have studied the association between MTHFR c.677C>T and mucositis [1,4,5,24,27–29,31,34–38,40–64]. In eight studies a statistically significant association was found [27,34,37,42,46,47,57,62]. A total of 16 studies provided information on the number of cases and controls per genotype using the grade ≥ 3 versus grade 0-2 definition as mucositis endpoint and were included in the meta-analysis [1,31,34–37,46–48,50,55,56,58–60,62]. The study of Xie et al. was excluded from the meta-analysis, because of deviation from HWE in the control group (grade 0-2) [36]. The data of the other 15 studies were fitted best using a recessive model. Patients with the TT-genotype had a higher chance of

developing mucositis compared to the CT- and CC-genotype (Odds Ratio [OR] 2.53, 95%-CI [1.48-4.32], FDR-corrected p-value 0.011) (**Figure 3**). However, the heterogeneity was relatively large ($I^2=45\%$, moderate heterogeneity) and was mainly caused by the study of Tantawy et al. [62]. When this study was excluded the I^2 dropped to 8% and the OR was 1.86 (1.29-2.67) with an FDR-corrected p-value of 0.011 (**Figure S7**). There was no clear sign of publication bias based on the Funnel plot or with the Egger's test ($p=0.116$), with the exception of the study of Tantawy et al. The Leave-One-Out analysis showed a stable OR, including Tantawy et al., varying between 1.86 to 2.88 for which none of the 95%-CI's was including the value 1 (**Figure S8**).

Three subgroup analyses were conducted (**Figures S9-11**). The first was of studies in which patients received IV MTX ($n=14$). The association with mucositis was less pronounced, but still reached statistical significance after correction for multiple testing (OR 2.32, 95%CI [1.37-3.92], FDR-corrected p-value of 0.023). The second included studies with pediatric patients ($n=8$) which resulted in an OR of 2.90 (1.22-6.89) with an FDR-corrected p-value of 0.130. The third comprised studies with ALL patients only ($n=7$), in which the OR was 3.31 (1.09-10.04) with an FDR-corrected p-value of 0.242.

Applying different case-control definitions resulted in a non-significant association for the grade ≥ 1 versus grade 0 definition and a statistically significant association of MTHFR c.677C>T for the grade ≥ 3 versus grade 0-1 (excluding grade 2) mucositis definition with a OR of 2.39 (1.35-4.25), of which the FDR-corrected p-value was 0.026 (**Figures S12 and S13**).

3.6.2 *MTRR c.66A>G (rs1801394)*

In total, four studies were identified in which the association between *MTRR c.66A>G* and mucositis was studied [1,27,49,63]. In two studies a statistically significant association was found [27,49]. Only for the case-control definition of grade ≥ 1 versus grade 0 three of the four studies provided the number of events per genotype [1,27,49].

Applying the four-genetic model strategy to the 3 studies resulted in an overdominant model (AG vs GG+AA). This model showed an OR of 2.08 (1.16-3.73) with an FDR-corrected p-value of 0.042 (**Figure 4**). The heterogeneity was low (I^2 of 11%, $p=0.33$) and there was no indication of publication bias using 3 studies). The Leave-One-Out analysis showed that only when the study of Faganel Kotnik et al. was excluded the association was still statistically significant (OR 2.59, 95%CI [1.33-5.03]) (**Figure S14**). However, when one of the other two studies was excluded, no statistically significant association was found.

3.6.3 *SL19A1* c.80G>A (*rs1051266*)

Of the 18 studies conducted for *SL19A1* c.80G>A [1,4,12,26,27,29,30,34,35,47,49,50,54–56,59,61,67], three found a statistically significant association [29,55,59]. A total of 10 studies were included for the primary analysis using the grade ≥ 3 versus grade 0–2 case-control definition [1,30,34,35,47,50,55,56,59,67]. The four-genetic model strategy showed that the allelic model was fitting the data best. The A-allele was associated with a decrease in mucositis when compared to the G-allele (OR 0.52, 95%CI [0.32–0.85], FDR corrected p-value was 0.09, **Figure 5**). However, the heterogeneity was huge ($I^2=69\%$), mainly caused by the study of Salazar et al. When this study was excluded, the heterogeneity almost completely disappeared ($I^2=3\%$) and the association became statistically significant even after FDR correction (OR 0.68, 95%CI [0.52–0.89], FDR-corrected p-value was 0.037, **Figure S15**). The Leave-One-Out analysis showed a stable OR, varying between 0.48 and 0.68 and the 95%CI's did not include the value 1, **Figure S16**. Visual inspection of the funnel plot showed no clear indication of publication bias, except for the fact that the study of Salazar et al. was clearly an outlier in the funnel plot. Three subgroup analyses were conducted. The first covered the studies in which patients received IV MTX ($n=9$) and resulted in an OR of 0.48 (95%CI [0.29–0.79], FDR-corrected p-value = 0.036, **Figure S17**). The second included the pediatric studies ($n=6$) and the third comprised patients with ALL only ($n=5$). Both ORs were close to the overall OR, but not statistically significant after FDR correction for multiple testing (pediatric: OR 0.45, 95%CI [0.23–0.89], FDR-corrected p-value = 0.134, **Figure S18**; ALL: OR 0.56, 95%CI [0.27; 1.18], FDR-corrected p-value = 0.639, **Figure S19**).

No statistically significant association was found when changing the case-control definition into ≥ 1 versus 0 or ≥ 3 vs 0–1 (excluding grade 2) (**Figure S20 and S21**).

3.6.4 *TYMS* 2R>3R (*rs34743033*)

Of the twelve studies conducted for *TYMS* 2R>3R [27,29,34,35,43,44,47,49,54,56,63,65], seven studies reported an effect size or number of mucositis events in the publication [27,29,43,44,56,63,65]. For six studies additional information was provided after our request [27,34,35,47,56,65]. All studies showed a lower risk of mucositis for the 3R3R (+2R/3R) genotype compared to other genotype(s), except for the study by Oosterom et al. However, in only 2 of the 12 studies this effect was statistically significant [27,56]. For five studies information was available per genotype and for each grade [34,35,47,56,65]. The four-genetic model strategy showed that the overdominant model best fitted the data. However, the result was close to the border between the overdominant and dominant model. Therefore, we have tested both the overdominant and dominant model. Both models did not show any statistically significant association

between *TYMS* 2R>3R and mucositis, and showed a low study heterogeneity ($I^2=17\%$ and $I^2=0\%$) (**Figure S22 and S23**).

3.6.5 *ABCC2* c.24C>T (*rs717620*)

Six studies reported on the *ABCC2* c.24C>T SNP in relationship with mucositis [1,12,33,35,56,66]. Two studies found a statistically significant association [12,33]. Three studies provided the data per grade and per genotype [1,35,56]. In total, four studies were included in the analysis using the ≥ 3 versus 0-2 case-control definition [1,33,35,56]. The dominant model (CT +TT vs CC) was fitting the data best, but the meta-analysis did not show any association. The heterogeneity was very high ($I^2=82\%$). This was probably caused by the deviant findings of Liu et al. [33]. When this study was excluded the heterogeneity disappeared ($I^2=0\%$), but there was still no statistically significant association found between the *ABCC2* c.24C>T and mucositis (**Figure S24 and S25**).

3.6.6 *SLCO1B1* c.1865+4846T>C (*rs11045879*)

Five studies analyzed the association between *SLCO1B1* c.1865+4846T>C and mucositis [12,22,23,56,67]. Two studies found a statistically significant association between *SLCO1B1* c.1865+4846T>C and mucositis [22,23]. Four studies were included in the meta-analysis using the ≥ 3 versus 0-2 case-control definition [22,23,56,67]. One of these studies both reported the results for the discovery and replication cohort and both cohorts were separately included in the meta-analysis [23]. The data fitted the over-dominant model best using the four-genetic model strategy. However, the result was close to the border between the over-dominant and dominant model. Both the over-dominant and dominant model showed no statistically significant association of the *SLCO1B1* c.1865+4846T>C and mucositis (**Figure S26 and S27**).

3.6.7 *miR-1206* A>G (*rs2114358*)

Gutiérrez-Camino et al. [9] and López-López et al. [8] studied the association of *miR-1206* A>G with mucositis. They both showed a statistically significant association of the GG-genotype and a higher chance of developing mucositis compared to patients with the AA- or AG-genotype. The studies used different case-control definitions ≥ 3 vs 0-2 (Gutiérrez-Camino et al.) and ≥ 2 versus 0-1 (López-López et al.). With the provided information we were able to meta-analyze the two studies using the ≥ 2 versus 0-1 definition. Although the result of the four-genetic model strategy was close to the border with the recessive model, they fitted the over-dominant model better. We have tested both models, but did not find any statistically significant association with mucositis (**Figure S28 and S29**).

3.6.8 Other SNPs identified in review

In addition to the seven genetic variations studied in our meta-analysis, we identified 15 additional SNPs in this literature review that were significantly associated with mucositis. However, these SNPs had been studied in only one study. Most had ORs with a 95%-CI close to 1. The exception was *GGH* c.452C>T (rs11545078) with a very low OR of 0.11 (0.02-0.56), **Table 2**.

In the GWAS of Treviño et al. two SNPs were found to be statistically significant associated with clinically relevant MTX-induced gastrointestinal toxicity [22]: *SLCO1B1* c.1865+4846T>C (rs11045879), included in our meta-analysis, and *SLCO1B1* c.1865+248G>A (rs4149081; OR 15.3, 95%CI [7.9-24.6], p = 0.03). Only the study of López-López also studied the latter SNP, but did not find a significant association [12].

4. Discussion

In the present review, we included 57 articles studying a large number of genetic variations involved in the pharmacodynamics and pharmacokinetics of MTX in relation to mucositis. Twenty-six articles were included in our meta-analysis. Seven genetic variations were found to be associated with mucositis in at least two independent studies. Of these, two SNPs – *MTHFR* c.677C>T and *MTRR* c.66A>G – were found to be statistically significant in our meta-analysis, and one – *SLC19A1* c.80G>A – was statistically significant after reducing heterogeneity by excluding one outlying study.

The data show that carriers of the *MTHFR* c.677C>T TT-genotype and *MTRR* c.66A>G AG-genotype were at increased risk and carriers of the *SLC19A1* c.80G>A A-allele were decreased risk of developing (severe) mucositis compared to patients with (less severe) or no mucositis. Methylenetetrahydrofolate Reductase (*MTHFR*), Methionine Synthase Reductase (*MTRR*) and Solute Carrier Family 19 member 1 (*SLC19A1*) - also known as the reduced folate carrier 1 (*RFC1*) – are important enzymes and a transporter respectively in folate and MTX metabolism. *MTHFR* plays a central role in folate metabolism by catalyzing the conversion of 5,10-methylenetetrahydrofolate (THF) to 5-methylTHF, which is the primary circulating form of folate and is utilized in many subsequent reactions important for purine- and thymidine synthesis and homocysteine remethylation pathway [68]. *MTHFR* c.677C>T causes alanine to be replaced by valine and is associated with a thermolabile *MTHFR* enzyme with reduced activity [68,69]. *MTRR* is involved in remethylating homocysteine to methionine. Methionine is an essential amino acid, necessary for protein synthesis and one carbon metabolism [70]. *MTRR* c.66A>G causes isoleucine to be replaced by methionine and is associated with a decreased enzyme activity [71]. In our study, the overdominant model (AG vs. AA+GG) was significant, whereas we expected to observe an effect of the GG-genotype or G-allele. Given

that studies included in our meta-analysis were relatively small, we recommend this variant to be tested in a large independent validation cohort. *SLC19A1* is one of the primary transporters involved in transporting MTX into the cell. *SLC19A1* c.80G>A results in the substitution of a histidine for an arginine in the protein sequence and has been associated with possibly altered uptake of MTX and higher plasma folate levels [72].

The only GWAS published by Treviño et al. showed that *SLCO1B1* c.1865+4846T>C T-allele (OR 16.4, 95%CI[8.7-26.7]) and c.1865+248G>A G-allele (OR 15.3, 95%CI [7.9-24.6]) were associated with gastrointestinal toxicity [22]. For *SLCO1B1* c.1865+4846T>C we did not find a significant association in our meta-analysis, in which four candidate-gene studies and the GWAS results were taken into account [Figure S26-27]. For *SLCO1B1* c.1865+248G>A, another smaller candidate-gene study by López-López et al. also showed the lack of a significant association with mucositis [12]. For both SNPs it would be interesting to study their association with mucositis further in larger patient cohorts. In addition, *GGH* c.452C>T (OR 0.11, 95%CI[0.02-0.56]) showed a large effect in one study and would therefore be valuable to be tested in an independent validation cohort or in a GWAS setting [25].

In our review, we observed that the main focus in the field of pharmacogenomics of MTX-induced mucositis was on pediatric (43/57 studies) ALL (42/57 studies) patients using IV MTX at doses varying between 2 to 5 g/m² (44/57 studies). When conducting a subgroup analysis in these three categories (IV MTX - pediatric patients - ALL patients) the number of studies included in each decreased. Consequently, statistical significance decreased after FDR-correction. Still, effect sizes remained similar and showed the same directions of effects. These subgroup analyses further emphasized that creating large homogeneous cohorts with similar patient and treatment characteristics are needed to be able to extrapolate results to certain populations. For *MTRR* c.66A>G studies were too few to be able to conduct subgroup analyses.

A large variety of chemotherapy protocols – including often concomitant chemotherapy – and endpoints was used. Differences between treatment protocols could lead to differences in the risk of mucositis. Furthermore, the definition of a mucositis case was different between the studies, which might have influenced the results. In our study, we tried to take this factor into account by only analyzing studies with a clear and comparable case-control definition and by mostly focusing on grade 3 or higher. A grade 3 mucositis requires medical intervention, such as a hospitalization, extra pain management or a nasal tube to aid nutrition [73]. Given that these interventions - especially when studied retrospectively - are often mentioned in the patient record file, it becomes less likely that cases were missed. For *MTHFR* c.677C>T and *SLC19A1* c.80A>G, it was possible to analyze other case-control definitions than the grade 3 or higher

versus grade 0-2 definition, because of available data per grade. When using the grade 1 or higher versus grade 0, the effect became less pronounced for both SNPs. When using the grade 3 or higher versus grade 0-1 (excluding grade 2) definition, the effect became more pronounced for *SLC19A1* c.80A>G. This comparison was unfortunately impossible for *MTRR* c.66A>G, where only grade 1 or higher versus the grade 0 definition were available to be analyzed. It remains an important factor to take into account in future studies that mucositis should preferably be assessed in a prospective, valid and reliable manner. For future pharmacogenomic studies in which the mucositis is prospectively graded, it would be interesting to use a case definition of grade ≥ 2 instead of grade ≥ 3 , because it is clinically relevant to find genetic risk factors that predict an increased risk of developing grade ≥ 2 mucositis. High-risk patients can then be closely monitored to allow timely interventions to prevent severe mucositis from occurring. For controls only grade 0 should be used to compare real controls (patients without any grade of mucositis). For our meta-analysis we didn't use this definition for cases (grade ≥ 2 mucositis) and controls (grade 0 mucositis), because this would have substantially reduced the number of included studies.

In a large proportion of the included studies, important baseline information on the cancer type, immunophenotype or risk classification was missing. In over one-fifth of the studies, the description of the laboratory methods used was very limited or completely missing. In more than half of the studies information on the quality of genotyping was missing. Sample size calculations were almost never provided and corrections for multiple testing were only carried out in a small proportion of the studies. Finally, only two studies adjusted for self-reported ethnicity. We therefore concluded that the number of high-quality studies remains limited and that abovementioned factors are important to be taken into account and to be described in future studies.

With regard to the results of our meta-analysis, it is important to note that effect sizes range between ORs of 0.52 and 2.63, never exceeding a 2.6-fold increased or decreased risk MTX-induced mucositis. Most effect sizes of risk alleles in meta-analyses are relatively small. This is often observed in pharmacogenomics of complex traits such as the multifactorial process of developing MTX-induced mucositis [74,75], but also could be due to improper study design issues as noted. Studies were often underpowered, studied cohorts were heterogeneous with regard to clinical and treatment characteristics and all but one [22] cohort studied genes in a candidate gene approach. It is known, that these types of studies are suboptimal for studying multiple genetic variants and rare genetic variants leading to extreme phenotypes [74,76,77]. In future studies, large sample sizes with clearly defined genetic materials, as well as prospectively collected treatment and toxicity data of good quality allowing for subgroup analyses,

should be studied to further elucidate the role of the five polymorphisms identified in this study – *MTHFR* c.677C>T, *MTRR* c.66A>G, *SLC19A1* c.80G>A, *SLCO1B1* c.1865+4846T>C and *SLCO1B1* c.1865+248G>A – and to identify possible new polymorphisms. In these studies, complex gene-gene and gene-environment interactions should be taken into account as the strategy of studying single variant associations has not proven to be very successful [75]. To date, oral and gastrointestinal mucositis/toxicity were often studied in one combined measure of toxicity, due to power issues. In future studies it might be valuable to study these two phenotypes separately since genetics might influence oral mucositis and gastrointestinal mucositis/toxicity differently.

In conclusion, we reviewed all reported genetic variation studies regarding the development of MTX-induced mucositis in the adult and pediatric cancer population. *MTHFR* c.677C>T is a valuable candidate to study in future genetic prediction models. The role of *MTRR* c.66A>G, *SLC19A1* c.80G>A, *SLCO1B1* c.1865+4846T>C and c.1865+248G>A remains unclear and should be further studied in large, homogenous cohorts with clear case-control definitions.

CRedit author statement

Hedy Maagdenberg: Conceptualization, Methodology, Investigation, Formal analysis, Visualization. Writing- Original Draft **Natanja van Oosterom:** Methodology, Validation, Resources, Writing- Original Draft **Jolanda Zanen:** Methodology, Investigation, Writing- Review & Editing **Donato Gemmati:** Resources, Writing - Review & Editing **Rachael E. Windsor:** Resources, Writing - Review & Editing **Sandra G. Heil:** Supervision, Writing - Review & Editing **Patricia Esperón:** Resources, Writing - Review & Editing **Shakila Jabeen:** Resources, Writing - Review & Editing **Guillermo J. Ruiz-Argüelles:** Resources, Writing - Review & Editing **Oliver Zolk:** Resources, Writing - Review & Editing **Susanne Hoerning:** Resources, Writing - Review & Editing **Charlotte Sleurs:** Resources, Writing - Review & Editing **Elixabet Lopéz-Lopéz:** Resources, Writing - Review & Editing **Mónica Moreno-Galván:** Resources, Writing - Review & Editing **Mary M. van den Heuvel-Eibrink:** Supervision, Writing - Review & Editing **Anke-Hilse Maitland-van der Zee** Methodology, Supervision, Writing - Review & Editing **Bruce Carleton:** Conceptualization, Methodology, Supervision, Writing - Review & Editing.

Conflict of interest

All authors approved the manuscript and agree with its submission to Critical Reviews in Oncology/ Hematology. Dr. Maitland-van der Zee reports research grants from GSK and Boehringer Ingelheim outside the submitted work. Furthermore, she received personal fees paid to the AmsterdamUMC from Boehringer Ingelheim and Astra Zeneca. All other authors have no conflict of interests.

Funding

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

Acknowledgements

We like to thank K. Seidemann, W.E. Evans, B. Faganel Kotnik, I. Badell and J. Salazar, for providing additional information.

References

1. den Hoed MAH, López-López E, te Winkel ML, Tissing W, de Rooij JDE, Gutiérrez-Camino Á, et al. Genetic and metabolic determinants of methotrexate-induced mucositis in pediatric acute lymphoblastic leukemia. *Pharmacogenomics J*. 2015 Jun;15(3):248–54.
2. Relling MV, Fairclough D, Ayers D, Crom WR, Rodman JH, Pui CH, et al. Patient characteristics associated with high-risk methotrexate concentrations and toxicity. *J Clin Oncol Off J Am Soc Clin Oncol*. 1994 Aug;12(8):1667–72.
3. Sonis ST, Elting LS, Keefe D, Peterson DE, Schubert M, Hauer-Jensen M, et al. Perspectives on cancer therapy-induced mucosal injury: pathogenesis, measurement, epidemiology, and consequences for patients. *Cancer*. 2004 May 1;100(9 Suppl):1995–2025.
4. Shimasaki N, Mori T, Samejima H, Sato R, Shimada H, Yahagi N, et al. Effects of methylenetetrahydrofolate reductase and reduced folate carrier 1 polymorphisms on high-dose methotrexate-induced toxicities in children with acute lymphoblastic leukemia or lymphoma. *J Pediatr Hematol Oncol*. 2006 Feb;28(2):64–8.
5. Liu S-G, Li Z-G, Cui L, Gao C, Li W-J, Zhao X-X. Effects of methylenetetrahydrofolate reductase gene polymorphisms on toxicities during consolidation therapy in pediatric acute lymphoblastic leukemia in a Chinese population. *Leuk Lymphoma*. 2011;52(6):1030–40.
6. Khan ZA, Tripathi R, Mishra B. Methotrexate: a detailed review on drug delivery and clinical aspects. *Expert Opin Drug Deliv*. 2012 Feb;9(2):151–69.
7. Schmiegelow K. Advances in individual prediction of methotrexate toxicity: a review. *Br J Haematol*. 2009;146(5):489–503.
8. López-López E, Sánchez-Toledo J, Uriz JJ, Ballesteros J, García-Miguel P, Navajas A, et al. Pharmacogenetics of MicroRNAs and MicroRNAs biogenesis machinery in pediatric acute lymphoblastic leukemia. *PLoS ONE*. 2014;9(3).
9. Gutiérrez-Camino A, Oosterom N, den Hoed MAH, López-López E, Martín-Guerrero I, Pluijm SMF, et al. The miR-1206 microRNA variant is associated with methotrexate-induced oral mucositis in pediatric acute lymphoblastic leukemia. *Pharmacogenet Genomics*. 2017;27(8):303–6.
10. Roberts TC. The microRNA Machinery. *Adv Exp Med Biol*. 2015;887:15–30.
11. Spyridopoulou KP, Dimou NL, Hamodrakas SJ, Bagos PG. Methylene tetrahydrofolate reductase gene polymorphisms and their association with methotrexate toxicity: a meta-analysis. *Pharmacogenet Genomics*. 2012;22(2):117–33.
12. López-López E, Ballesteros J, Piñan MA, Sánchez De Toledo J, García De Andoin N, García-Miguel P, et al. Polymorphisms in the methotrexate transport pathway: A new tool for MTX plasma level prediction in pediatric acute lymphoblastic leukemia. *Pharmacogenet Genomics*. 2013;23(2):53–61.

13. Zhao M, Liang L, Ji L, Chen D, Zhang Y, Zhu Y, et al. MTHFR gene polymorphisms and methotrexate toxicity in adult patients with hematological malignancies: a meta-analysis. *Pharmacogenomics*. 2016 Jun 1;17(9):1005–17.
14. Zhu C, Liu YW, Wang SZ, Li XL, Nie XL, Yu XT, et al. Associations between the C677T and A1298C polymorphisms of MTHFR and the toxicity of methotrexate in childhood malignancies: a meta-analysis. *Pharmacogenomics J*. 2018 22;18(3):450–9.
15. Moher D, Liberati A, Tetzlaff J, Altman DG, PRISMA Group. Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. *BMJ*. 2009 Jul 21;339:b2535.
16. Klein T, Altman R, Whirl-Carrillo M, Gong L, Sangkuhl K, Barbarino J. PharmGKB: Stanford University. [Internet]. 2018. Available from: <https://www.pharmgkb.org>
17. Cariaso M, Lennon G. SNPedia: a wiki supporting personal genome annotation, interpretation and analysis. *Nucleic Acids Res*. 2012 Jan;40(Database issue):D1308-1312.
18. Bethesda (MD): National Library of Medicine (US). PubMed. [Internet]. Available from: <https://www.ncbi.nlm.nih.gov/pubmed/>
19. Little J, Higgins JPT, Ioannidis JPA, Moher D, Gagnon F, von Elm E, et al. Strengthening the reporting of genetic association studies (STREGA): an extension of the STROBE Statement. *Hum Genet*. 2009 Mar;125(2):131–51.
20. Leusink M, Onland-Moret NC, de Bakker PIW, de Boer A, Maitland-van der Zee AH. Seventeen years of statin pharmacogenetics: a systematic review. *Pharmacogenomics*. 2016;17(2):163–80.
21. Horita N, Kaneko T. Genetic model selection for a case-control study and a meta-analysis. *Meta Gene*. 2015 Sep;5:1–8.
22. Treviño LR, Shimasaki N, Yang W, Panetta JC, Cheng C, Pei D, et al. Germline genetic variation in an organic anion transporter polypeptide associated with methotrexate pharmacokinetics and clinical effects. *J Clin Oncol Off J Am Soc Clin Oncol*. 2009;27(35):5972–8.
23. Stocco G, Yang W, Crews KR, Thierfelder WE, Decorti G, Londero M, et al. PACSIN2 polymorphism influences TPMT activity and mercaptopurine-related gastrointestinal toxicity. *Hum Mol Genet*. 2012 Nov 1;21(21):4793–804.
24. Aplenc R, Thompson J, Han P, La M, Zhao H, Lange B, et al. Methylenetetrahydrofolate reductase polymorphisms and therapy response in pediatric acute lymphoblastic leukemia. *Cancer Res*. 2005;65(6):2482–7.
25. Chen X, Wen F, Yue L, Li C. Genetic polymorphism of γ -glutamyl hydrolase in Chinese acute leukemia children and identification of a novel double nonsynonymous mutation. *Pediatr Hematol Oncol*. 2012 May;29(4):303–12.
26. Faganel Kotnik B., Dolžan V., Grabnar I., Jazbec J. Relationship of the reduced folate carrier gene polymorphism G80A to methotrexate plasma concentration, toxicity, and disease outcome in childhood acute lymphoblastic leukemia. *Leuk Lymphoma*. 2010;51(4):724–6.

27. Faganel Kotnik B., Grabnar I., Bohanec Grabar P., Dolžan V., Jazbec J. Association of genetic polymorphism in the folate metabolic pathway with methotrexate pharmacokinetics and toxicity in childhood acute lymphoblastic leukaemia and malignant lymphoma. *Eur J Clin Pharmacol.* 2011;67(10):993–1006.
28. Karathanasis NV, Stiakaki E, Goulielmos GN, Kalmanti M. The role of the methylenetetrahydrofolate reductase 677 and 1298 polymorphisms in Cretan children with acute lymphoblastic leukemia. *Genet Test Mol Biomark.* 2011;15(1–2):5–10.
29. Kishi S, Cheng C, French D, Pei D, Das S, Cook EH, et al. Ancestry and pharmacogenetics of antileukemic drug toxicity. *Blood.* 2007;109(10):4151–7.
30. Kotnik BF, Jazbec J, Grabar PB, Rodriguez-Antona C, Dolžan V. Association between SLC19A1 gene polymorphism and high dose methotrexate toxicity in childhood acute lymphoblastic leukaemia and non Hodgkin malignant lymphoma: Introducing a haplotype based approach. *Radiol Oncol.* 2017;51(4):455–62.
31. Lambrecht L, Sleurs C, Labarque V, Dhooge C, Laenen A, Sinnaeve F, et al. The role of the MTHFR C677T polymorphism in methotrexate-induced toxicity in pediatric osteosarcoma patients. *Pharmacogenomics.* 2017;18(8):787–95.
32. Liu S-G, Gao C, Zhang R-D, Jiao Y, Cui L, Li W-J, et al. FPGS rs1544105 polymorphism is associated with treatment outcome in pediatric B-cell precursor acute lymphoblastic leukemia. *Cancer Cell Int.* 2013;13(1):107.
33. Liu Y, Yin Y, Sheng Q, Lu X, Wang F, Lin Z, et al. Association of ABCC2 -24C>T polymorphism with high-dose methotrexate plasma concentrations and toxicities in childhood acute lymphoblastic leukemia. *PLoS One.* 2014;9(1):e82681.
34. Ongaro A, De Mattei M, Della Porta MG, Rigolin G, Ambrosio C, Di Raimondo F, et al. Gene polymorphisms in folate metabolizing enzymes in adult acute lymphoblastic leukemia: effects on methotrexate-related toxicity and survival. *Haematologica.* 2009;94(10):1391–8.
35. Windsor RE, Strauss SJ, Kallis C, Wood NE, Whelan JS. Germline genetic polymorphisms may influence chemotherapy response and disease outcome in osteosarcoma: a pilot study. *Cancer.* 2012;118(7):1856–67.
36. Xie L, Guo W, Yang Y, Ji T, Xu J. More severe toxicity of genetic polymorphisms on MTHFR activity in osteosarcoma patients treated with high-dose methotrexate. *Oncotarget.* 2018 Feb 20;9(14):11465–76.
37. Xu L, Wang L, Xue B, Wang S. MTHFR variant is associated with high-dose methotrexate-induced toxicity in the Chinese osteosarcoma patients. *J Bone Oncol.* 2018 Nov;13:143–7.
38. de Carvalho DC, Wanderley AV, Dos Santos AMR, Fernandes MR, Cohen Lima de Castro A de N, Leitão LPC, et al. Pharmacogenomics and variations in the risk of toxicity during the consolidation/maintenance phases of the treatment of pediatric B-cell leukemia patients from an admixed population in the Brazilian Amazon. *Leuk Res.* 2018;74:10–3.

39. Gutiérrez-Camino Á, Umerez M, López-López E, Santos-Zorrozua B, Martín-Guerrero I, de Andoin NG, et al. Involvement of miRNA polymorphism in mucositis development in childhood acute lymphoblastic leukemia treatment. *Pharmacogenomics*. 2018;19(18):1403–12.
40. Aráoz HV, D'Aloi K, Foncuberta ME, Sánchez La Rosa CG, Alonso CN, Chertkoff L, et al. Pharmacogenetic studies in children with acute lymphoblastic leukemia in Argentina. *Leuk Lymphoma*. 2015;56(5):1370–8.
41. Ayad MW, El Naggar AA, El Naggar M. MTHFR C677T polymorphism: association with lymphoid neoplasm and effect on methotrexate therapy. *Eur J Haematol*. 2014 Jul;93(1):63–9.
42. Eissa DS, Ahmed TM. C677T and A1298C polymorphisms of the methylenetetrahydrofolate reductase gene: effect on methotrexate-related toxicity in adult acute lymphoblastic leukaemia. *Blood Coagul Fibrinolysis Int J Haemost Thromb*. 2013;24(2):181–8.
43. Erculj N, Kotnik BF, Debeljak M, Jazbec J, Dolžan V. Influence of folate pathway polymorphisms on high-dose methotrexate-related toxicity and survival in childhood acute lymphoblastic leukemia. *Leuk Lymphoma*. 2012;53(6):1096–104.
44. Erculj N, Kotnik BF, Debeljak M, Jazbec J, Dolžan V. The influence of folate pathway polymorphisms on high-dose methotrexate-related toxicity and survival in children with non-Hodgkin malignant lymphoma. *Radiol Oncol*. 2014;48(3):289–92.
45. Frikha R, Rebai T, Lobna BM, Frikha F, Mdhaftar M, Frikha I, et al. Comprehensive analysis of Methylenetetrahydrofolate reductase C677T in younger acute lymphoblastic leukemia patients: A single-center experience. *J Oncol Pharm Pract Off Publ Int Soc Oncol Pharm Pract*. 2019 Jul;25(5):1182–6.
46. Gemmati D, Ongaro A, Tognazzo S, Catozzi L, Federici F, Mauro E, et al. Methylenetetrahydrofolate reductase C677T and A1298C gene variants in adult non-Hodgkin's lymphoma patients: Association with toxicity and survival. *Haematologica*. 2007;92(4):478–85.
47. Giletti A, Vital M, Lorenzo M, Cardozo P, Borelli G, Gabus R, et al. Methotrexate pharmacogenetics in Uruguayan adults with hematological malignant diseases. *Eur J Pharm Sci Off J Eur Fed Pharm Sci*. 2017;109(c7r, 9317982):480–5.
48. Haase R, Elsner K, Merkel N, Stiefel M, Mauz-Korholz C, Kramm CM, et al. High dose methotrexate treatment in childhood ALL: pilot study on the impact of the MTHFR 677C>T and 1298A>C polymorphisms on MTX-related toxicity. *Klin Padiatr*. 2012;224(3):156–9.
49. Huang L, Tissing WJE, de Jonge R, van Zelst BD, Pieters R. Polymorphisms in folate-related genes: association with side effects of high-dose methotrexate in childhood acute lymphoblastic leukemia. *Leukemia*. 2008;22(9):1798–800.
50. Jabeen S, Holmboe L, Alnæs GIG, Andersen AM, Hall KS, Kristensen VN. Impact of genetic variants of RFC1, DHFR and MTHFR in osteosarcoma patients treated with high-dose methotrexate. *Pharmacogenomics J*. 2015;15(5):385–90.

51. Mahmoud LB, Mdhaffar M, Frikha R, Ghazzi H, Hakim A, Sahnoun Z, et al. Use of MTHFR C677T polymorphism and plasma pharmacokinetics to predict methotrexate toxicity in patients with acute lymphoblastic leukemia. *Adv Clin Exp Med Off Organ Wroclaw Med Univ.* 2018 Aug;27(8):1061–8.
52. Roy Moulik N, Kumar A, Agrawal S, Awasthi S, Mahdi AA, Kumar A. Role of folate status and methylenetetrahydrofolate reductase genotype on the toxicity and outcome of induction chemotherapy in children with acute lymphoblastic leukemia. *Leuk Lymphoma.* 2015;56(5):1379–84.
53. Moulik N, Kumar A, Agrawal S, Mahdi A, Kumar A. Effect of folate status and methylenetetrahydrofolate reductase genotypes on the complications and outcome of high dose methotrexate chemotherapy in north Indian children with acute lymphoblastic leukemia. *Indian J Med Paediatr Oncol.* 2016;37(2):85–9.
54. Pakakasama S, Kanchanakamhaeng K, Kajanachumpol S, Udomsubpayakul U, Sirachainan N, Thithapandha A, et al. Genetic polymorphisms of folate metabolic enzymes and toxicities of high dose methotrexate in children with acute lymphoblastic leukemia. *Ann Hematol.* 2007;86(8):609–11.
55. Park JA, Shin HY. Influence of genetic polymorphisms in the folate pathway on toxicity after high-dose methotrexate treatment in pediatric osteosarcoma. *Blood Res.* 2016 Mar 1;51(1):50–7.
56. Radtke S, Zolk O, Renner B, Paulides M, Zimmermann M, Moricke A, et al. Germline genetic variations in methotrexate candidate genes are associated with pharmacokinetics, toxicity, and outcome in childhood acute lymphoblastic leukemia. *Blood.* 2013;121(26):5145–53.
57. Ramírez-Pacheco A, Moreno-Guerrero S, Alamillo I, Medina-Sanson A, Lopez B, Moreno-Galván M. Mexican Childhood Acute Lymphoblastic Leukemia: A Pilot Study of the MDR1 and MTHFR Gene Polymorphisms and Their Associations with Clinical Outcomes. *Genet Test Mol Biomark.* 2016;20(10):597–602.
58. Ruiz-Argüelles GJ, Coconi-Linares LN, Garcés-Eisele J, Reyes-Núñez V. Methotrexate-induced mucositis in acute leukemia patients is not associated with the MTHFR 677T allele in Mexico. *Hematol Amst Neth.* 2007 Oct;12(5):387–91.
59. Salazar J, Altes A, del Rio E, Estella J, Rives S, Tasso M, et al. Methotrexate consolidation treatment according to pharmacogenetics of MTHFR ameliorates event-free survival in childhood acute lymphoblastic leukaemia. *Pharmacogenomics J.* 2012;12(5):379–85.
60. Seidemann K, Book M, Zimmermann M, Meyer U, Welte K, Stanulla M, et al. MTHFR 677 (C-->T) polymorphism is not relevant for prognosis or therapy-associated toxicity in pediatric NHL: results from 484 patients of multicenter trial NHL-BFM 95. *Ann Hematol.* 2006;85(5):291–300.
61. Suthandiram S, Gan G-G, Zain SM, Bee P-C, Lian L-H, Chang K-M, et al. Effect of polymorphisms within methotrexate pathway genes on methotrexate toxicity and plasma levels in adults with hematological malignancies. *Pharmacogenomics.* 2014 Aug;15(11):1479–94.

62. Tantawy AAG, El-Bostany EA, Adly AAM, Abou El Asrar M, El-Ghouroury EA, Abdulghaffar EE. Methylene tetrahydrofolate reductase gene polymorphism in Egyptian children with acute lymphoblastic leukemia. *Blood Coagul Fibrinolysis*. 2010;21(1):28–34.
63. Yazıcıoğlu B, Kaya Z, Ergun SG, Perçin F, Koçak Ü, Yenicesu İ, et al. Influence of folate-related gene polymorphisms on high-dose methotrexate-related toxicity and prognosis in Turkish children with acute lymphoblastic leukemia. *Turk J Hematol*. 2017;34(2):143–50.
64. Choi YJ, Park H, Lee JS, Lee J-Y, Kim S, Kim TW, et al. Methotrexate elimination and toxicity: MTHFR 677C>T polymorphism in patients with primary CNS lymphoma treated with high-dose methotrexate. *Hematol Oncol*. 2017;35(4):504–9.
65. Oosterom N, Berrevoets M, den Hoed MAH, Zolk O, Hoerning S, Pluijm SMF, et al. The role of genetic polymorphisms in the thymidylate synthase (TYMS) gene in methotrexate-induced oral mucositis in children with acute lymphoblastic leukemia. *Pharmacogenet Genomics*. 2018;28(10):223–9.
66. Sharifi MJ, Bahoush G, Zaker F, Ansari S, Rafsanjani KA, Sharafi H. Association of -24CT, 1249GA, and 3972CT ABCB2 gene polymorphisms with methotrexate serum levels and toxic side effects in children with acute lymphoblastic leukemia. *Pediatr Hematol Oncol*. 2014;31(2):169–77.
67. Liu S-G, Gao C, Zhang R-D, Zhao X-X, Cui L, Li W-J, et al. Polymorphisms in methotrexate transporters and their relationship to plasma methotrexate levels, toxicity of high-dose methotrexate, and outcome of pediatric acute lymphoblastic leukemia. *Oncotarget*. 2017;8(23):37761–72.
68. Leclerc D, Sibani S, Rozen R. Molecular Biology of Methylene tetrahydrofolate Reductase (MTHFR) and Overview of Mutations/Polymorphisms [Internet]. *Madame Curie Bioscience Database* [Internet]. Landes Bioscience; 2013 [cited 2019 Mar 10]. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK6561/>
69. Frosst P, Blom HJ, Milos R, Goyette P, Sheppard CA, Matthews RG, et al. A candidate genetic risk factor for vascular disease: a common mutation in methylenetetrahydrofolate reductase. *Nat Genet*. 1995 May;10(1):111–3.
70. van der Linden IJM, den Heijer M, Afman LA, Gellekink H, Vermeulen SHHM, Kluijtmans LAJ, et al. The methionine synthase reductase 66A>G polymorphism is a maternal risk factor for spina bifida. *J Mol Med Berl Ger*. 2006 Dec;84(12):1047–54.
71. Li W-X, Cheng F, Zhang A-J, Dai S-X, Li G-H, Lv W-W, et al. Folate Deficiency and Gene Polymorphisms of MTHFR, MTR and MTRR Elevate the Hyperhomocysteinemia Risk. *Clin Lab*. 2017 Mar 1;63(3):523–33.
72. Yee SW, Gong L, Badagnani I, Giacomini KM, Klein TE, Altman RB. SLC19A1 pharmacogenomics summary. *Pharmacogenet Genomics*. 2010 Nov;20(11):708–15.
73. Cancer Therapy Evaluation Program (CTEP). Common Terminology Criteria for Adverse Events (CTCAE) - version 5 [Internet]. 2017 [cited 2019 Mar 10]. Available from: https://ctep.cancer.gov/protocoldevelopment/electronic_applications/docs/CTCAE_v5_Quick_Reference_5x7.pdf

74. Kang J, Cho J, Zhao H. Practical issues in building risk-predicting models for complex diseases. *J Biopharm Stat.* 2010 Mar;20(2):415–40.
75. Ioannidis JPA, Trikalinos TA, Khoury MJ. Implications of small effect sizes of individual genetic variants on the design and interpretation of genetic association studies of complex diseases. *Am J Epidemiol.* 2006 Oct 1;164(7):609–14.
76. Relling MV, Evans WE. Pharmacogenomics in the clinic. *Nature.* 2015 Oct 15;526(7573):343–50.
77. Moriyama T, Relling MV, Yang JJ. Inherited genetic variation in childhood acute lymphoblastic leukemia. *Blood.* 2015 Jun 25;125(26):3988–95.

Short biographies of co-authors

Hedy Maagdenberg, PhD, is currently working as an advisor at the National Health Care Institute, Diemen, The Netherlands, and as a post-doctoral researcher for the research group of Dr. Bruce Carleton, Vancouver, Canada. She previously obtained a master's degree in Pharmacy and a doctor's degree from the University of Utrecht, Utrecht, The Netherlands. Her research focuses on pharmacogenetics/genomics in pediatric patients.

Natanja Oosterom, MD PhD, is currently a resident in pediatrics. She previously performed a PhD in the field of pediatric oncology on '(Epi)genetic and biochemical determinants of methotrexate-induced oral mucositis in pediatric acute lymphoblastic leukemia' under supervision of prof. dr. M.M. van den Heuvel-Eibrink, prof. dr. R. Pieters and dr. S.G. Heil.

Jolanda G. Zanen obtained a master's degree in Pharmacy from the University of Utrecht, Utrecht, The Netherlands. She is currently specializing as a hospital pharmacist at the HagaZiekenhuis, The Hague, The Netherlands.

Donato Gemmati is Prof. of Medical Genetics at the Dpt. of Translational Medicine, University of Ferrara, Italy. Interests: molecular medicine, pharmacogenetics/genomics, epigenetics, and gender medicine. Editorial Board of the International Journal of Molecular Science and Genes. Chief of the "Genetic/Epigenetic Mother/Child Dyad Study" collaborative group (GEMCDS) on the mother-fetus cross-talk during pregnancy and pediatric diseases.

Dr Rachael Windsor, BSc, MBBS, MSc, MD Res is a Consultant Paediatric and Adolescent Oncologist in the London Sarcoma Service at University College London Hospitals, London, UK. Her other clinical interest is the long-term effects of cancer treatment and she is a core clinician within the Late Effects Service. She undertook an MD investigating the influence of pharmacogenomics on the response to chemotherapy in osteosarcoma.

Dr. Sandra G. Heil is molecular biologist and leads the Folate and Vitamin Metabolism group within the Department of Clinical Chemistry at the Erasmus MC University Medical Center Rotterdam. The aim of the group is to develop novel biomarkers related to folate- and vitamin B12 metabolism and to assess its association with diseases such as vitamin B12 deficiency and methotrexate efficacy and toxicity in rheumatoid arthritis and leukemia.

Patricia Esperón Percovich (PhD). Full Professor Molecular Genetics (teaching, research and management activities) School of Chemistry, Universidad de la República Montevideo, Uruguay. Head of Molecular Genetic Laboratory of the Tumor Bank of the Hospital Central de las Fuerzas Armadas, Montevideo Uruguay

Shakila Jabeen, PhD. She has both a masters' degree (2006 in field of Cell Biology) and doctor's degree (2018 in field of Breast Onco-immunology) from University in Oslo, Norway. She currently holds a position as researcher at the Clin. Mol. Biology department, Breast cancer group/ Research and Innovation Division, Akershus University Hospital, Lorenskog, Norway. Her research focuses on breast cancer, onco-immunology (cytokines and tumor micro environment), cancer drug resistance relative to SNPs and relative to patients immune profiles.

Guillermo J. Ruiz-Argüelles MD, FRCP(Glasg), MACP, DSc(hc), FRCP is past president of the International Society of Hematology, of the Mexican Society of Hematology and of the Mexican College of Internal Medicine. He has authored 365 papers in peer-reviewed journals. Elected Distinguished Alumnus of the Mayo Clinic and Doctor Honoris Causa by the Universidad Autónoma de San Luis Potosí, his main interests are stem cell transplantation, malignant hematological diseases and thrombophilia.

Oliver Zolk, MD, is Head of the Institute of Clinical Pharmacology and Professor of Clinical Pharmacology at the Brandenburg Medical School Theodor Fontane. His research interests include the various aspects of personalized drug therapy, such as pharmacogenetics, avoiding drug-drug interactions, improving the safety and efficacy of drug therapy, and especially the prevention of late effects of cancer treatment in children.

Dr. Susanne Radtke, MD, is currently in residency training in pediatrics. Her doctoral research focused on pharmacogenetic markers as predictors of efficacy and tolerability of chemotherapy in childhood cancer.

Charlotte Sleurs is a post-doctoral researcher of the Pediatric Oncology research group of KU Leuven, Belgium. Her research focuses on chemotherapy and radiotherapy-induced neurotoxicity, its potential risk factors and predictive biomarkers.

Elixabet López-López has a degree in Biology from the Complutense University of Madrid, Spain and a PhD from the University of the Basque Country (UPV/EHU), where she carries out her research work in the Group Genetics and Epigenetics of complex diseases. Her main line of research is the identification of genetic biomarkers in pediatric cancers and lymphoma, with a special interest in pharmacogenomics.

Mónica Moreno-Galván has a degree in Biology at the Universidad Nacional Autónoma de México, Master In Sciences with a specialty in Molecular Biomedicine, PhD in Medicine Research. Her experience is based on pharmacogenetics and angiogenesis research.

Marry M. van den Heuvel-Eibrink, MD PhD, is a professor in Translational Pediatric Oncology at the Princess Máxima Center for Pediatric Oncology. Her research is dedicated to translational research with a special focus on renal tumors and the genetic variation of early and late toxicity in childhood cancer.

Prof. Dr. A.H. Maitland van der Zee is Professor of precision medicine in respiratory disease at the Amsterdam UMC since 2016. Her research interest is in finding biomarkers that can optimize diagnosis and treatment for patients with respiratory disease.

Dr. Bruce Carleton is Director of the Pharmaceutical Outcomes Programme at the BC Children's Hospital and Professor of Pediatrics, Medical Genetics, Pharmaceutical Sciences, Population & Public Health at the University of British Columbia. He is the Chief of the Division of Translational Therapeutics, in the Department of Pediatrics and a Senior Clinician Scientist at the BC Children's Hospital Research Institute in Vancouver, Canada.

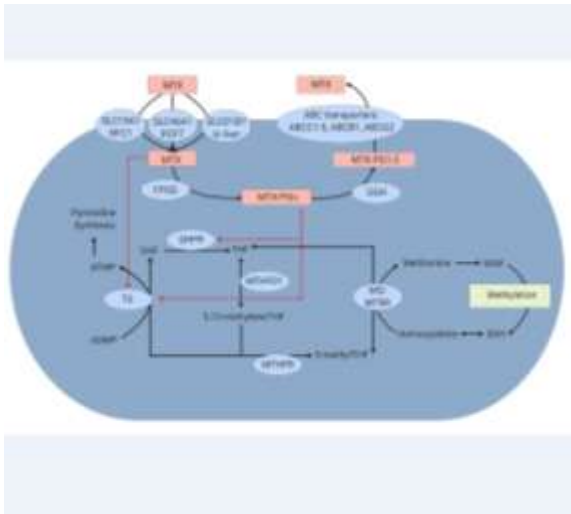


Figure 1. Important transporters and enzymes in MTX- and folate metabolism

Mechanism of action of MTX. MTX is transported into the cell by Reduced Folate Carrier 1 (SLC19A1/RFC1) / Proton-Coupled Folate Transporter (SLC49A1/PCFT). In the liver, MTX is transported by SLCO1B1. MTX is then polyglutamated by Folylpolyglutamate Synthetase (FPGS) and inhibits Dihydrofolate Reductase (DHFR) / Thymidylate Synthase (TS). MTX is de-polyglutamated by gamma-glutamyl hydrolase (GGH) and then transported out of the cell by ABCC1-5, ABCB1 and ABCG2. Methylene tetrahydrofolate reductase (MTHFR), Methylenetetrahydrofolate Dehydrogenase, Cyclohydrolase And Formyltetrahydrofolate Synthetase 1 (MTHFD1) and Methionine Synthase / Methionine Synthase Reductase (MS/MTRR) are important enzymes in the folate / one-carbon metabolism cycle.

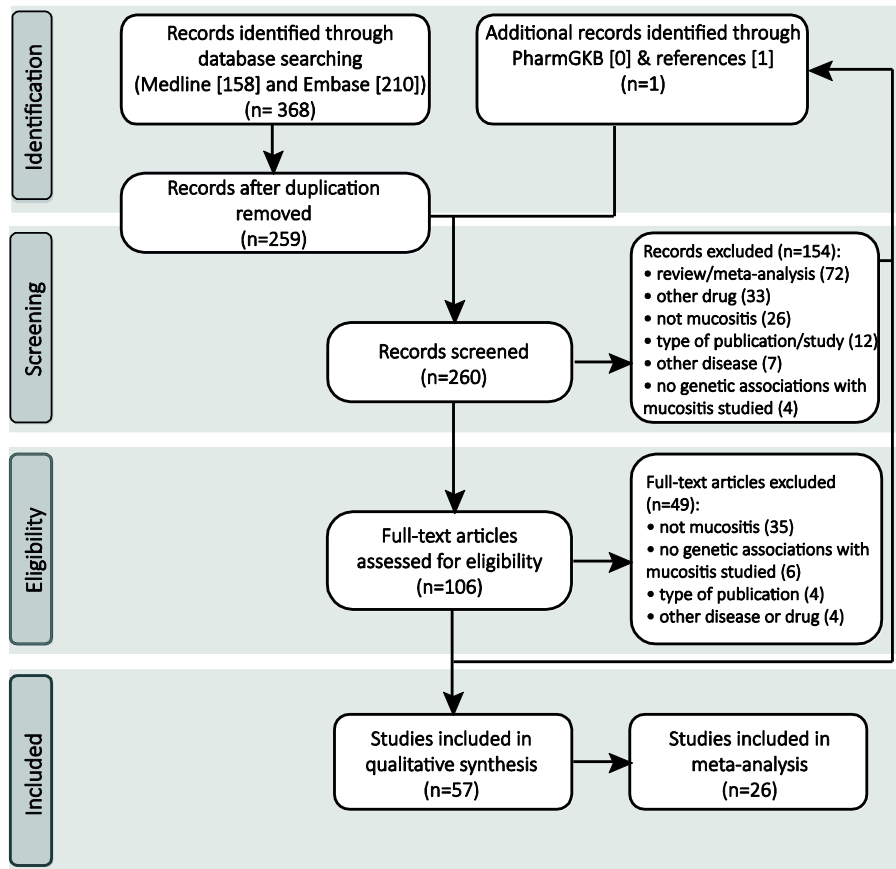


Figure 2. Flowchart

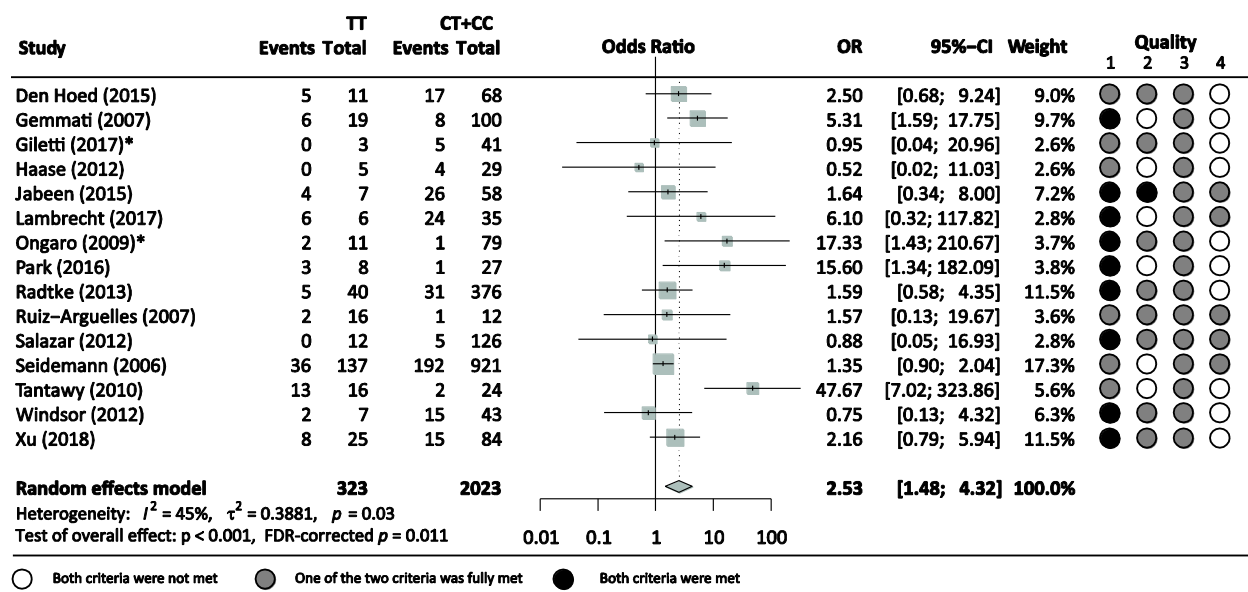


Figure 3. Meta-analysis results for *MTHFR* c.677C>T (rs1801133) grade ≥3 versus grade 0-2 mucositis or gastrointestinal toxicity

Abbreviations: OR odds ratio, CI confidence interval, * outcome was gastrointestinal toxicity, not mucositis.

Quality: item 1 = quality of clinical information; item 2 = quality of genotyping; item 3 = quality in reporting of study population origin; item 4 = quality in terms of sample size calculation and statistical correction for multiple testing (for more detailed information, see table S3 and S5).

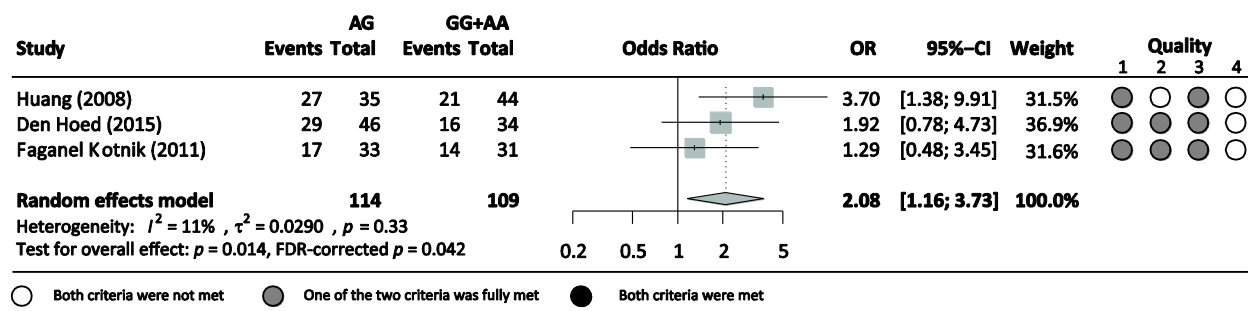


Figure 4. Meta-analysis results for *MTRR* c.66A>G (rs1801394) grade ≥1 versus grade 0 mucositis or gastrointestinal toxicity

Abbreviations: OR odds ratio, CI confidence interval. Quality: item 1 = quality of clinical information; item 2 = quality of genotyping; item 3 = quality in reporting of study population origin; item 4 = quality in terms of sample size calculation and statistical correction for multiple testing (for more detailed information, see table S3 and S5).

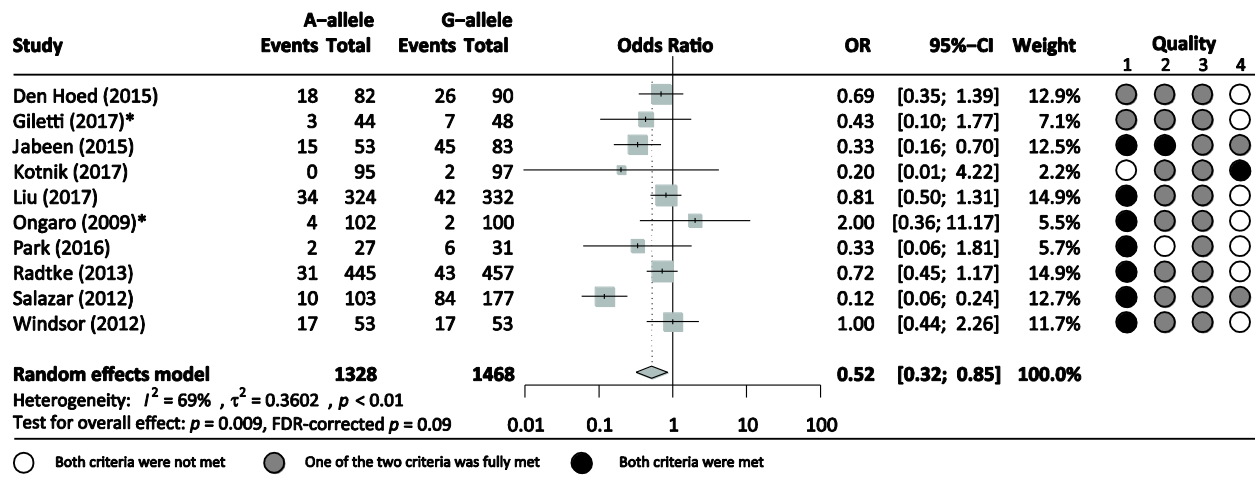


Figure 5. Meta-analysis results for *SL19A1* c.80G>A (*rs1051266*) grade ≥ 3 versus grade 0-2 mucositis or gastrointestinal toxicity

Abbreviations: OR odds ratio, CI confidence interval, * outcome was gastrointestinal toxicity, not mucositis.

Quality: item 1 = quality of clinical information; item 2 = quality of genotyping; item 3 = quality in reporting of study population origin; item 4 = quality in terms of sample size calculation and statistical correction for multiple testing (for more detailed information, see table S3 and S5).

Table 1. SNPs with at least one statistically significant replication

SNP	Number of studies that found a statistically significant association	Total number of studies [ref.]
<i>MTHFR</i> c.677C>T (<i>rs1801133</i>)	8	38 [1,4,5,24,27–29,31,34–38,40–64]
<i>MTRR</i> c.66A>G (<i>rs1801394</i>)	2	4 [1,27,49,63]
<i>TYMS</i> 2R>3R (<i>rs34743033</i>)	2	12 [27,29,34,35,43,44,47,49,54,56,63,65]
<i>ABCC2</i> c.24C>T (<i>rs717620</i>)	2	6 [1,12,33,35,56,66]
<i>SLC19A1</i> c.80G>A (<i>rs1051266</i>)	3	18 [1,4,12,26,27,29,30,34,35,47,49,50,54–56,59,61,67]
<i>SLCO1B1</i> c.1865+4846T>C (<i>rs11045879</i>)	2	5 [12,22,23,56,67]
<i>miR-1206</i> G>A (<i>rs2114358</i>)	2	2 [8,9]

Table 2. SNPs assessed in one study and with a statistically significant association

Gene name/miRNA and rs number	Genotype comparison	OR (95%CI)	p-value (corrected p-value)	Study
<i>DHFR</i> (rs1650723)	NR	NR	0.005	[50]
<i>CNOT1</i> (rs11866002)	TT vs CC + CT	6.68 (1.74-25.70)	0.0056 (0.3956)	[8]
hsa-mir 4268 (rs4674470)	AA + AG vs GG	0.31 (0.12-0.80)	0.0093 (0.011)	[39]
hsa-mir 3683 (rs6977967)	AA vs AG + GG	0 (0.0)	0.0098 (0.003)	[39]
<i>GGH</i> (rs11545078)	CT+TT vs CC	0.11 (0.02-0.56)	0.010	[25]
<i>ABCG2</i> (rs2231135)	NR	NR	0.01	[50]
<i>ABCC1</i> (rs2230671)	NR	NR	0.0105	[12]
<i>XPO5</i> (rs7755135)	AG + AA vs GG	2.68 (1.14-6.32)	0.0251	[8]
<i>EIF2C1</i> (rs595961)	NR	NR	0.0294	[8]
miR-146a (rs2910164)	CG + CC vs GG	2.54 (1.08-5.97)	0.0309	[8]
miR-1307 (rs7911488)	AG + GG vs AA	2.80 (1.04-7.54)	0.0311	[8]
<i>TNRC68</i> (rs139919)	NR	NR	0.0327	[8]
<i>SND1</i> (rs322825)	CT + TT vs CC	0.41 (0.17-0.96)	0.0394	[8]
<i>SLC22A6</i> (rs4149172)	NR	NR	0.0397	[12]
<i>ABCC3</i> (rs9895420)	AA + AT vs TT	0.22 (0.05-0.97)	0.044	[38]

Abbreviations: OR *odds ratio*, CI *confidence interval*, NR *not reported*.